

# Lecture 1: Cancer Biology for Modellers

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# Motivation

- To be effective biomathematicians, we need to “speak biology” as well mathematics and programming.
  - Helps in communicating with team members
  - Biological understanding helps inform:
    - Model formulation and analysis
    - Evaluation of modelling predictions
    - Parameter estimation
    - Calibration
      - relates to image processing
      - Need to understand what you’re seeing (histopathology)

# Lecture Outline

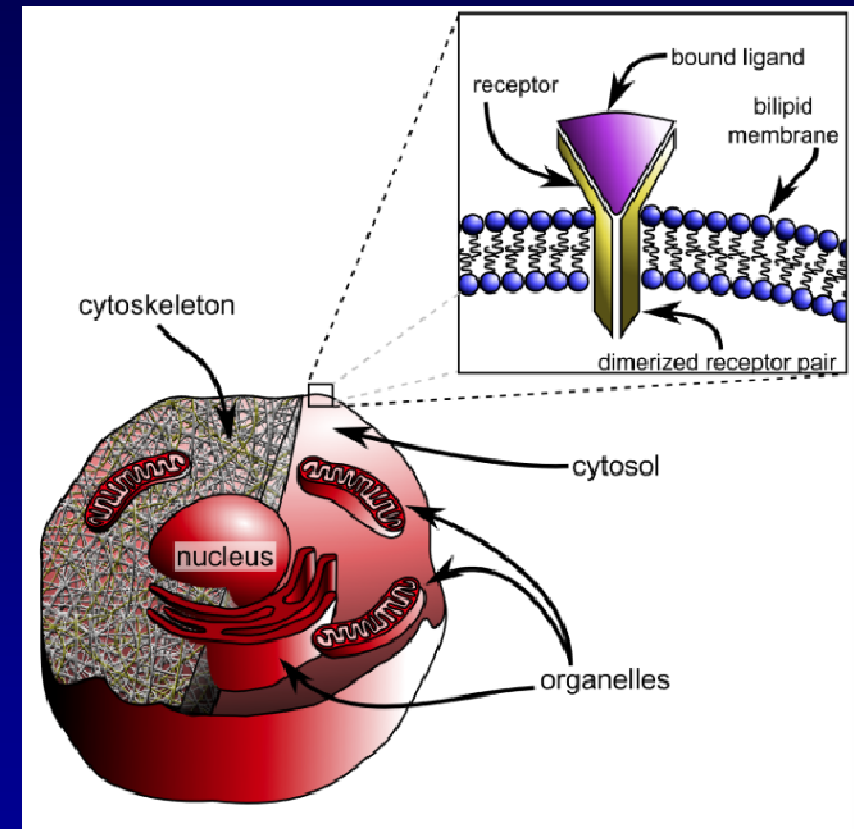
- Cell organisation
- Tissue organisation
- Maintaining tissue structure
- Cell birth and death
- Cell signalling
- Cell motility
- Oncogenes and tumour suppressor genes
- Abusing the system: cancer progression
- Coming next
- References and resources

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# Cell Organisation

- Bilipid layer surrounding cytoplasm
  - Permeable to small molecules
  - Requires active transport of others
    - Ion pumps control cell volume, pH, etc.
  - Impermeable to larger proteins
- Cytoskeleton provides structure
- Organelles embedded in the cytoplasm to perform specialised functions
- Relatively rigid nucleus in the centre
- Receptors on cell surface:
  - Mediate cell-microenvironment communication by binding to ligands
  - Mechanically link cell cytoskeleton to membrane and microenvironment

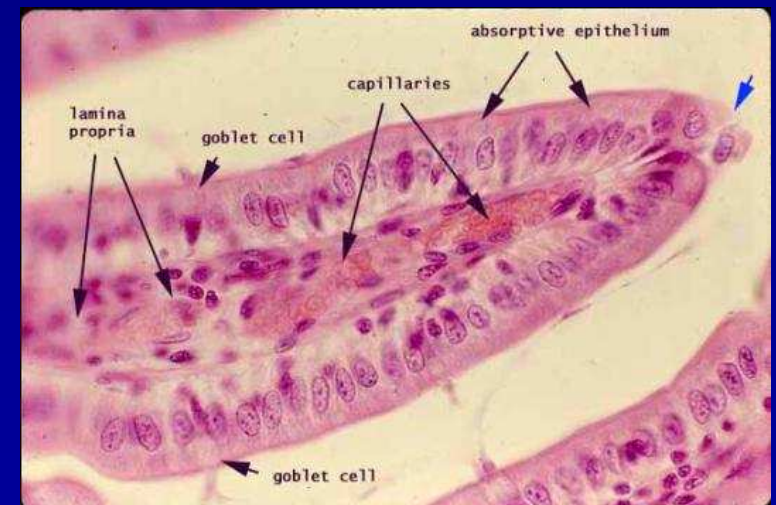
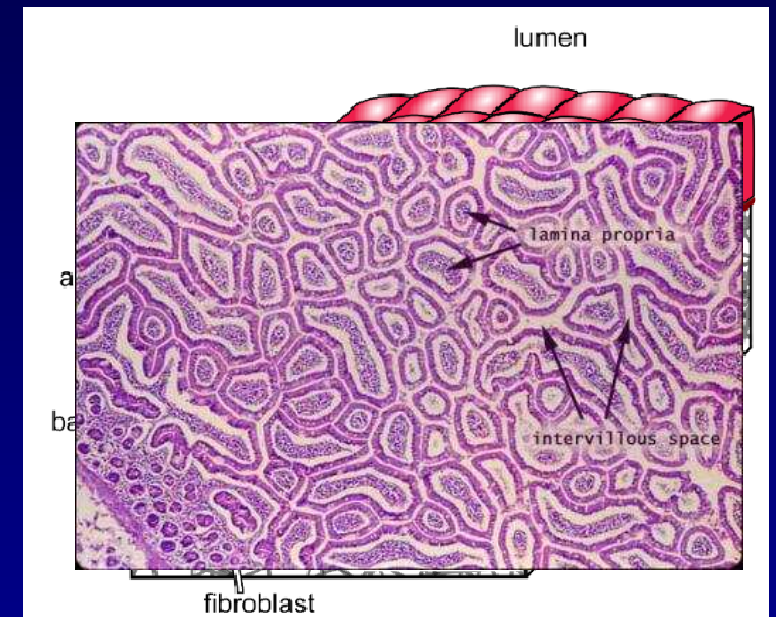


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# Tissue Organisation

- **Typical tissue microstructure in an organ:**
  - **Epithelial tissue (epithelium)**
    - Sheets of specialised cells that perform the work of the organ (secretory products, filtration, etc.)
  - **Loose connective tissue (stroma)**
    - Extracellular matrix (ECM) supports the organ
    - Contains blood vessels, lymphatics, nerves
  - **Tissues separated by basement membrane (BM)**
  - **Often a *lumen* (fluid- or air-filled cavity)**
    - Transports secretory products from/to epithelium
    - stroma-BM-epithelium microstructure: designed to maximise surface area of epithelium-lumen interface
- **Notable biophysics and transport:**
  - **BM is a physical barrier to cell motion between tissues**
  - **Oxygen, glucose, and growth factors can only reach epithelium by diffusion**



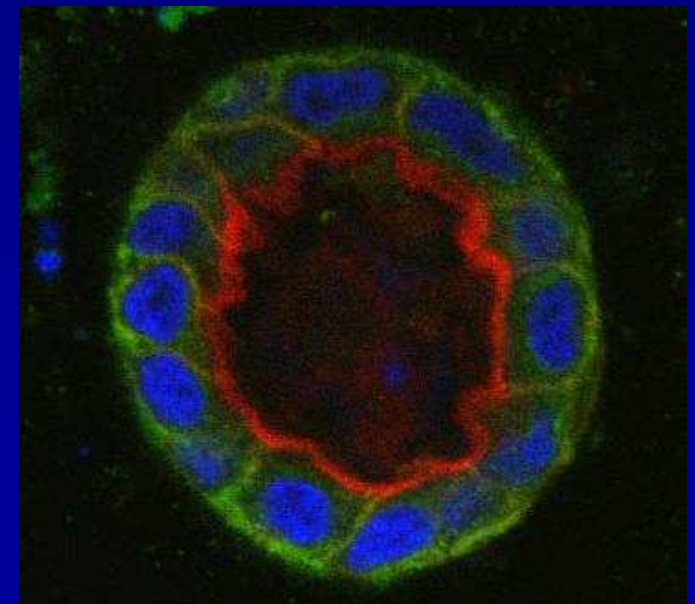
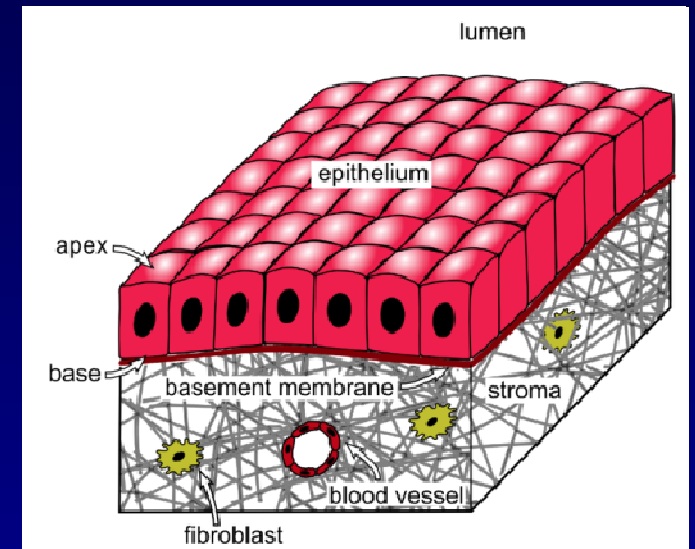
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- **Maintaining tissue structure**
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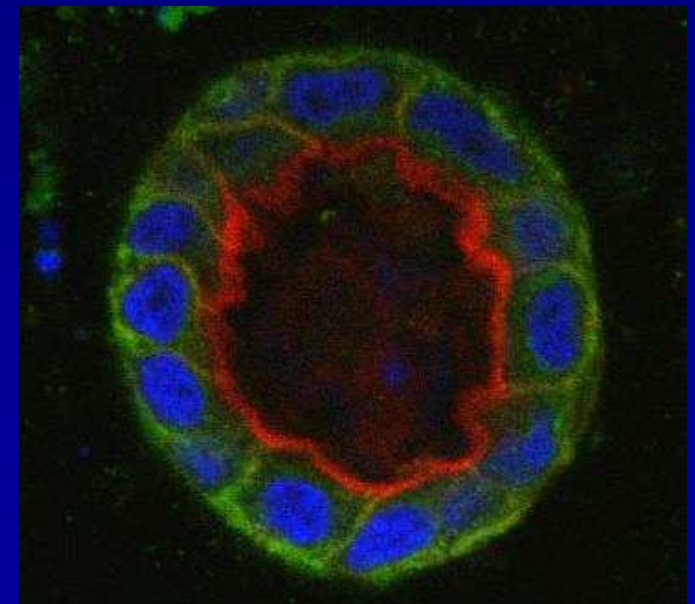
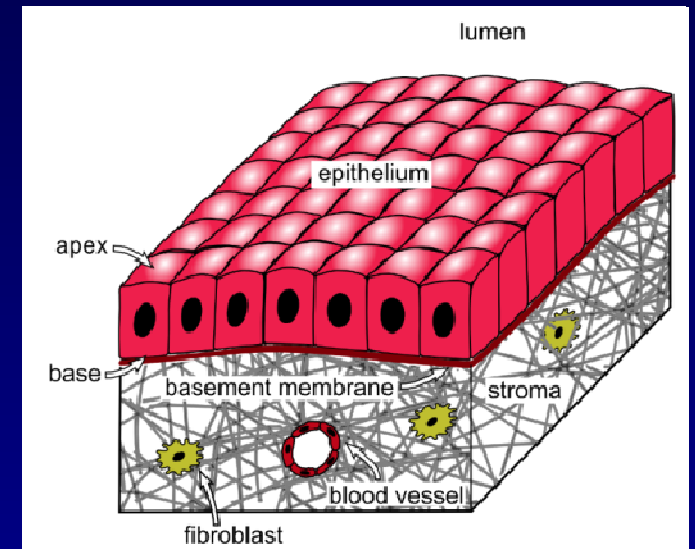
# Maintaining Tissue Structure: Mechanics

- Mesenchymal cells (e.g., fibroblasts) can move freely in the stroma, secrete and degrade ECM
- Epithelial cells are polarised:
  - Anisotropic adhesion receptor distribution
  - Integrins on base for cell-BM adhesion
    - Heterophilic adhesion
  - No adhesion receptors on apex
  - E-cadherins on basolateral sides for cell-cell adhesion
    - Homophilic adhesion
- Mechanics determine tissue geometry:
  - Balance of cell-cell and cell-BM strength
  - Distribution of cell receptors likely help in determine curvature
  - Mechanics of cell-BM adhesion, stresses, BM-to-ECM coupling likely determines BM curvature.



# Maintaining Tissue Structure: Population Dynamics

- Cell populations must be maintained in homeostasis
  - Proliferation to replace aged cells
  - Apoptosis to remove damaged cells, or those out of place
- Adhesion receptors are not merely mechanical:
  - E-cadherin helps detect presence/absence of neighbours
    - Trigger proliferation when missing a neighbour (E-cadherin/ $\beta$ -catenin)
    - Suppress cell cycle when attached to neighbours (block Cyclin D1, etc.)
  - Integrins detect detachment from BM
    - Trigger apoptosis (anoikis)
  - (This is why they're receptors. 😊)

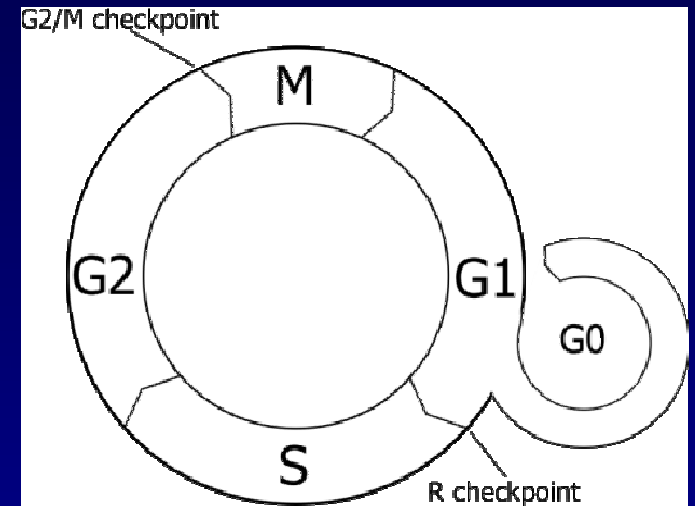


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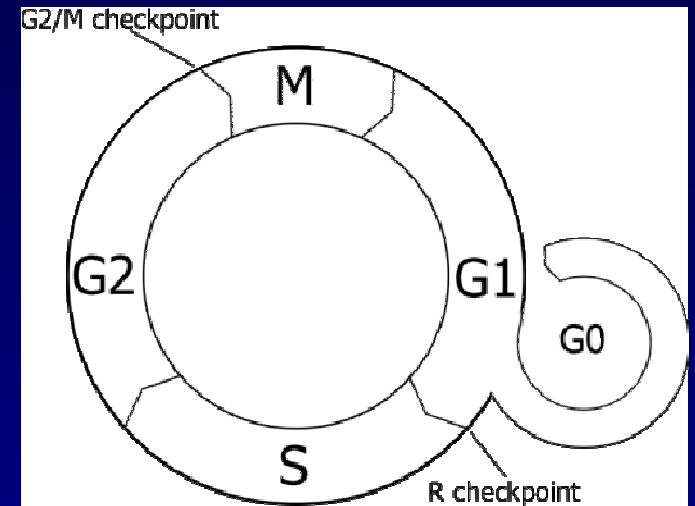
# Cell Birth and Death: Proliferation

- **Tightly-controlled cell-cycle:**
  - **G<sub>0</sub>:** quiescent “resting state”
  - **S:** Synthesis of new DNA
  - **G<sub>2</sub>:** “gap” phase – final prep for division
  - **M:** mitosis phase
    - DNA divided into two daughter nuclei (mitosis)
    - Cytoplasm and organelles divided into daughter cells (cytokinesis)
  - **G<sub>1</sub>:** “gap” or “growth” phase – daughter cells grow in volume, then exit cycle
  - **Note 1:** Some biologists treat G<sub>0</sub>+G<sub>1</sub> as long, variable-length G<sub>1</sub> phase (often seen in flow cytometry)
  - **Note 2:** Others treat G<sub>1</sub> as relatively fixed length, with G<sub>0</sub> of variable length (useful for Ki-67 matching)



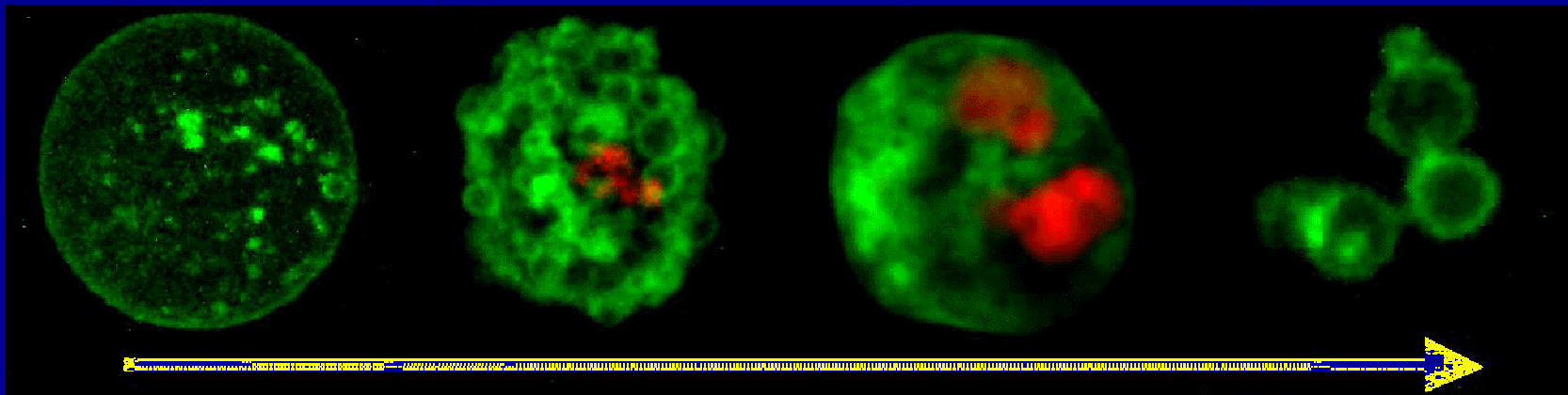
# Cell Birth and Death: Proliferation

- **Checkpoints provide opportunities for:**
  - **Cycle arrest: Restriction checkpoint R at G1/S**
  - **DNA error checking and repair**
  - **Apoptosis for irreparable damage**
  
- **Cycle progression controlled by intracellular signalling**
  - **Cyclins and cyclin-dependent kinases (CDKs)**
  - **Connected to other signalling networks**



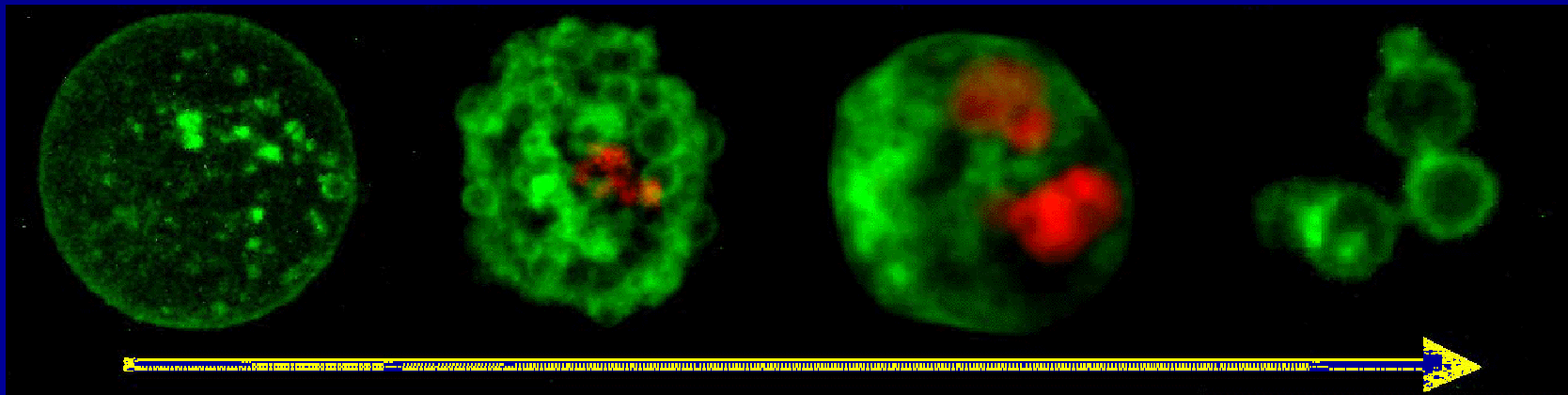
# Cell Birth and Death: Apoptosis

- **Apoptosis: orderly cell death**
  - Self-regulated, in response to signalling events
  - Early processes:
    - Mitochondria lose membrane potential, integrity
    - Pre-positioned caspases are activated (cleaved), begin degrading cell
  - Orderly cell volume shrinking
    - requires active  $\text{Na}^+$ ,  $\text{K}^+$  pumps!



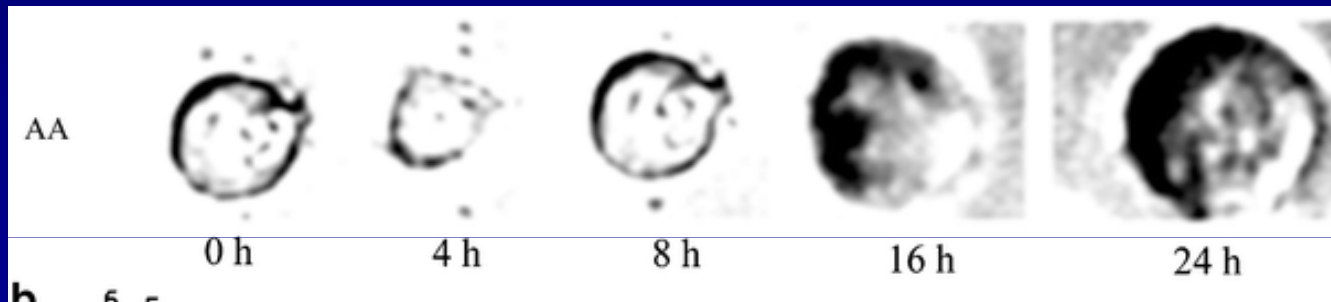
# Cell Birth and Death: Apoptosis

- **Apoptosis: orderly cell death (continued)**
  - DNA is chopped into pieces
  - Organelles disassembled
  - Cell contents encapsulated into apoptotic bodies
    - Protects surrounding cells from otherwise damaging reactions
  - Cell lyses to release apoptotic bodies
  - Apoptotic bodies phagocytosed
  - Entire process requires energy!



# Cell Birth and Death: Necrosis and Calcification

- **Necrosis: uncontrolled cell death**
  - Can occur due to energy depletion (hypoxia or hypoglycemia), mechanical injury, chemical stressors, failure during apoptosis, etc.
  - Uncontrolled cell volume
    - No energy for  $\text{Na}^+$ ,  $\text{K}^+$  pumps
    - Cell swells, then bursts
    - Cell contents released into microenvironment



- No orderly disassembly of nucleus, organelles, etc.
- Generally no phagocytosis
- Calcification:
  - $\text{Ca}^+$  pumps not active
  - Solid cell fraction replaced by calcium phosphate and oxalate crystals
  - Hard microcalcifications results





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# Cell Signalling

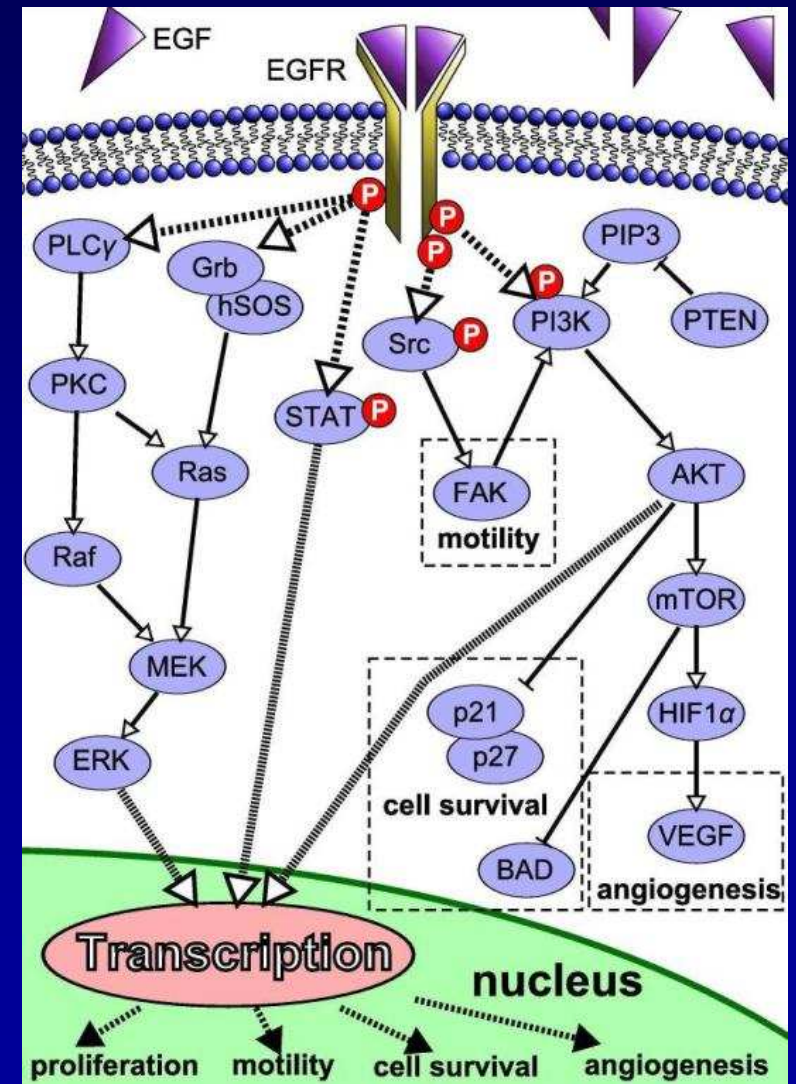
- Cells are regulated by internal signalling networks
  - DNA transcribes RNA
  - RNA encodes proteins
  - Proteins:
    - Assemble into structures
    - Perform “duties”
    - Transmit information through reactions
  - Network:
    - Redundancies
    - Feedback loops
    - Signal amplification

# Cell Signalling: Two examples

- HIF signalling:
  - Cells create hypoxia-inducible factors (HIFs)
    - Big example: HIF-1 $\alpha$
  - Ordinarily, O<sub>2</sub> tags these for degradation
  - During hypoxia, HIFs accumulate → HIFs as O<sub>2</sub> sensors
  - Trigger downstream transcription
    - Decreased cell adhesion
    - Increased motility
    - Glycolysis
    - Temporary resistance to apoptosis
    - VEGF secretion

# Cell Signalling: Two examples

- EGFR signalling
  - Epidermal growth factor (EGF) binds to EGFR receptor
  - Ligated EGFR receptors dimerise
  - Dimerised EGFR receptors phosphorylate intracellular proteins
  - Downstream actions:
    - Transcription
    - Increased motility
    - Increased proliferation
    - Cell survival

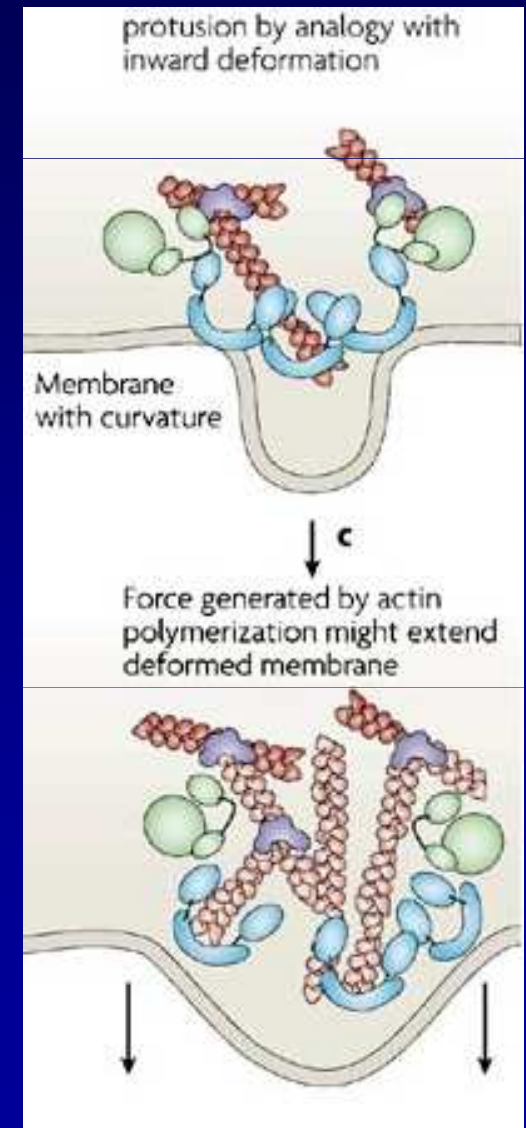
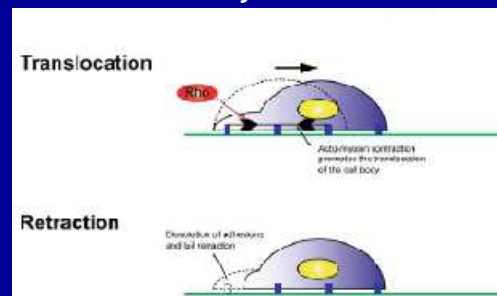
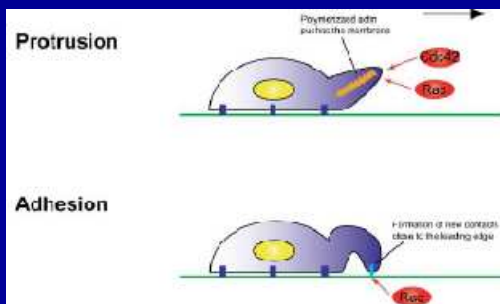


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# Cell Motility

- Actin polymerisation and depolymerisation:
  - Actin monomers join or leave chains to grow or shrink the cytoskeleton
- Cell signalling can nucleate biased polymerisation (e.g., in response to a gradient)
  - EGFR → Src → N-Wasp → Arp2/3 nucleation → ...
- Actin fibres deform and extend cell membrane, forming a pseudopod (“false foot”)
  - Filopodium: finger-like projection
  - Lamellipodium: sheet-like projection
- Cells focally secrete MMPs from invadopodia:
  - Degrade ECM
  - Break integrin bonds
  - Create space for motion
- Cell forms new focal adhesion on leading edge
- Actin polymers broken down in trailing edge (depolymerisation)
- Cell contracts to pull towards leading edge
- Protrusion – Adhesion – Contraction – Retraction cycles



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# Oncogenes and Tumour Suppressor Genes

- Oncogenes:
  - Proliferation-promoting genes : “gas pedal”
  - Examples:
    - Growth factor secretion (including autocrine): EGF, PDGF, VEGF
    - Internal promotion of cell cycle:  $\beta$ -catenin promotes transcription of Cyclin D1
    - Counter-act TSGs: MDM2 degrades p53
    - Growth factor receptors: EGFR / ErbB
    - Downstream regulators of receptor pathways: Ras, Raf, Src
- Tumour suppressor genes (TSGs):
  - Impede proliferation : “brake pedal”
  - Examples:
    - Block oncogenic signals: VHL helps degrade HIF-1 $\alpha$
    - Regulate cell cycle / create inhibitory signals: Rb impedes cell cycle, promotes arrest at G1/S
    - Repair DNA damage: TP53 impedes cell cycle to allow DNA repair at G1/S
    - Promote apoptosis: p53 can trigger apoptosis
  - Knudson 2-hit hypothesis:
    - You have 2 copies of each TSG, so need to eliminate both copies to lose function
      - Discovered in studying Rb (retinoblastoma) TSG
      - Mitigating factor: loss of heterozygosity
      - Mitigating factor: partial loss of function

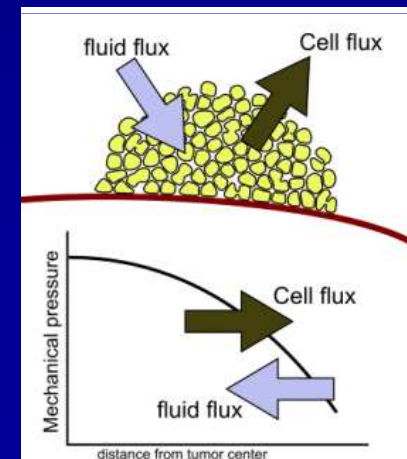
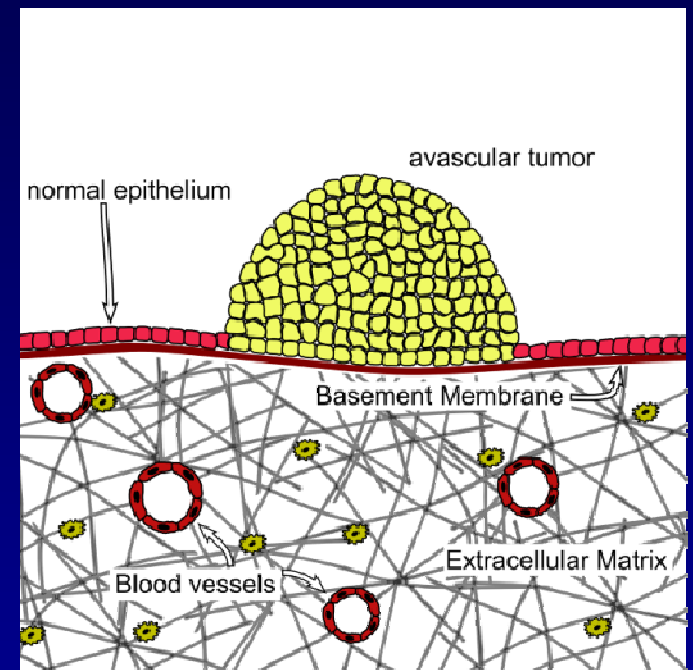


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# Abusing the system: Carcinogenesis

- **Mutations and/or epigenetic events:**
  - **Down-regulate apoptosis (lose TSGs)**
    - No anoikis – cells can survive in the lumen
    - p53 mutation – cells can ignore apoptosis signals
  - **Up-regulate proliferation (oncogenes)**
    - Decreased contact inhibition – proliferate even in the presence of neighbours
    - Increased secretion of growth signals → self-signalling
    - “stuck switches”
      - HER2 – can dimerise without binding ligand
        - » increases signalling activity
      - K-ras mutation – constitutive active, allowing EGFR signalling with EGF
- **Consequences:**
  - Increased survival fitness versus normal cells
  - Forms a colony of hyperproliferative cells
- **Mechanics:**
  - Mechanical pressure, stresses created by proliferating cells
  - Net outward flux of cells as mass grows – Darcy’s law
  - Net inward fluid flux

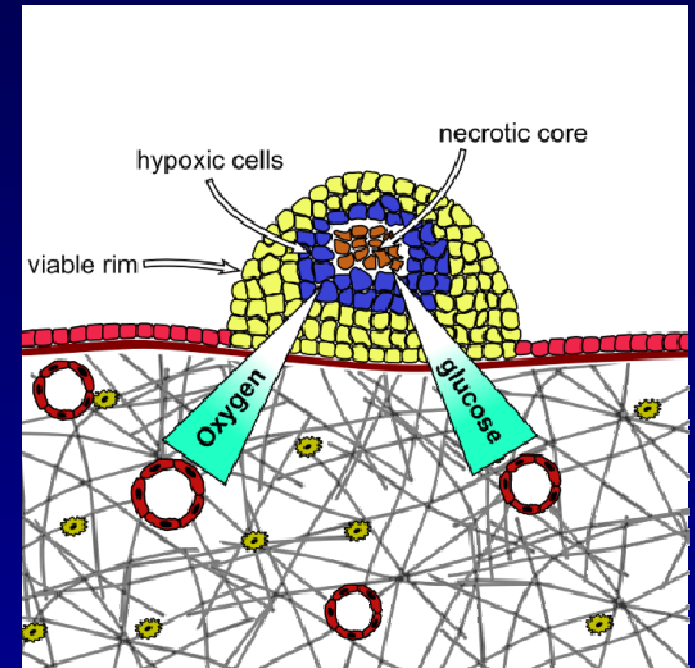


# Abusing the system: Avascular Growth

- Can only receive substrates by diffusion from the stroma
  - Substrate gradients
  - Hypoxia
  - Hyperglycemia

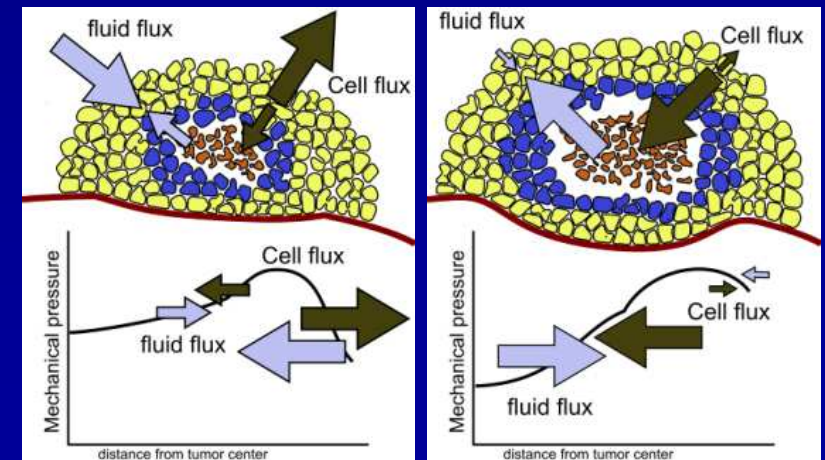
- Consequences:

- Viable rim thickness related to diffusion length scale
- Heterogeneous growth rates – relationship with oxygen and glucose (cell energetics)
- Inner band of hypoxic cells
  - HIF signalling
- Interior necrotic core



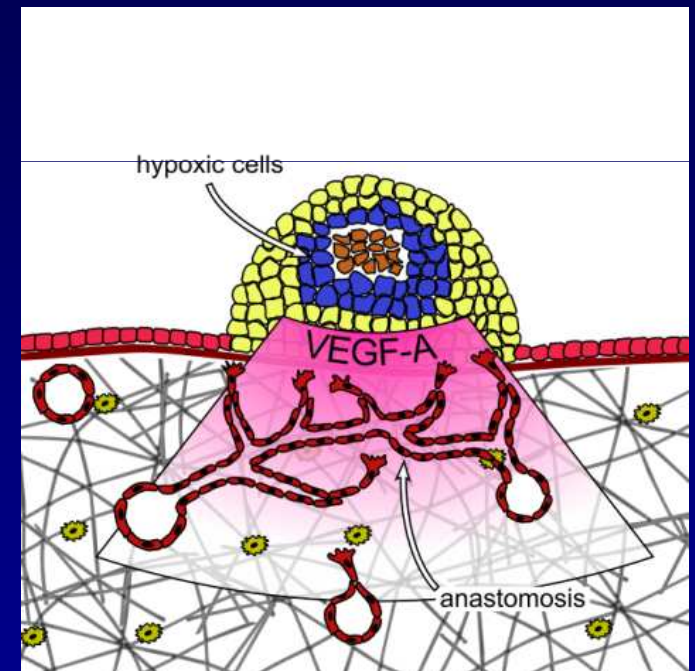
- Mechanics:

- Outward fluid flux from necrotic core due to lysis
- Inward cell flux due to reduced interior strain
- Steady tumour volume:
  - Cell flux out of viable rim  $\approx$  fluid flux from necrotic core



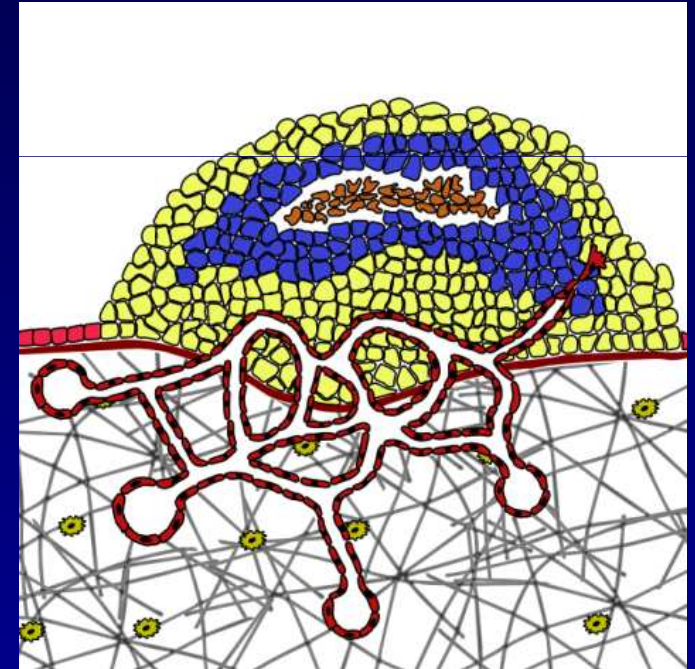
# Abusing the system: Angiogenesis

- Hypoxic cells release angiogenic factors
- VEGF diffuses into stroma
- Endothelial cells respond to VEGF
  - Degrade vessel walls
  - Chemotaxis (up  $\nabla$ VEGF), haptotaxis (up  $\nabla$ ECM)
  - Increased proliferation
  - Temporary suspension of anoikis (to facilitate survival until new basal lamina surrounds mature vessels)
- New vessels grow towards tumour, generally towards the VEGF gradient
- New vessels cross-link (anastomose)
- New blood flow in the vessels – new substrate transport to fuel further tumour growth
- Vessels mature: pericytes recruited, deposit basal lamina around vessels



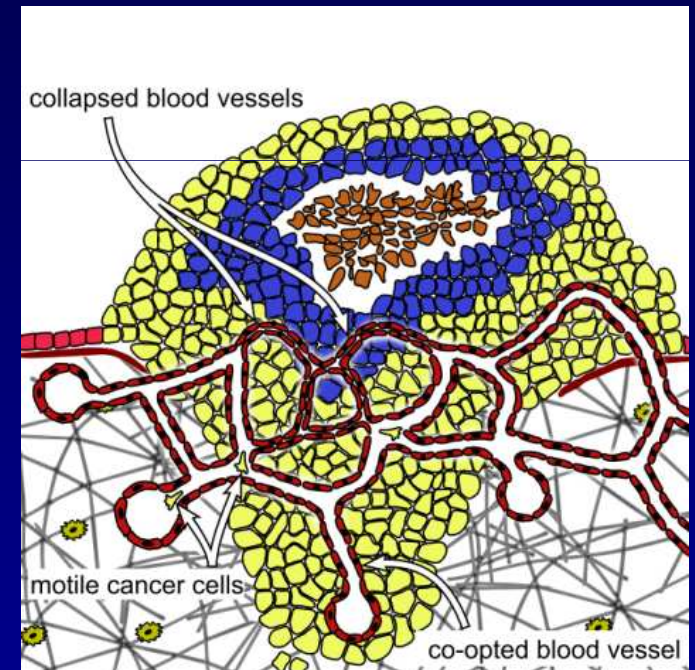
# Abusing the system: Vascular Growth & Invasion

- **New vessels fuel rapid tumour growth**
- **BM deformation and stress**
- **Hypoxic stress remains**
  - Glycolysis
  - Acidosis
  - *Selection pressures*



# Abusing the system: Vascular Growth & Invasion

- **Mutations**
  - Acid-resistant
  - BM and ECM degradation (by MMPs)
  - Motility
- **Invasion into stroma**
  - By growth and by motility
  - Co-option of blood vessels
- **Sustained tissue stress**
  - Collapsed blood vessels
  - New rounds of angiogenesis
- **Metastasis**
  - Travel through blood vessels and lymphatics
  - Extravasation, growth of new tumour
  - Possible role of metastatic niche



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# Coming Next:

- **Lecture 1:**
  - Cancer biology for modellers
- **Lecture 2:**
  - **An agent-based cell model; application to DCIS**
- **Lecture 3:**
  - Parameter estimation, patient-specific calibration
- **Lecture 4:**
  - Numerical method, simulation results



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# Some References

- Some biology texts:
  - P. Macklin. Biological background. In: V. Cristini and J. Lowengrub. *Multiscale Modeling of Cancer*. Cambridge University Press, Cambridge, UK, 2010. Chapter 2, pages 8-24. ISBN 978-0521884426. (in press)
  - B. Alberts et al. *Molecular Biology of the Cell*. Garland Science, New York, NY USA, 5<sup>th</sup> edition, 2007. ISBN 978-0815341116.
  - M. Knowles and P. Selby, eds., *Introduction to the Cellular and Molecular Biology of Cancer*. Oxford Univ. Press, Oxford, UK, 4<sup>th</sup> edition, 2005. ISBN 0-19-852563-X.
- Also see some great modelling texts by Wodarz & Komarova, Anderson et al.
- Some great websites:
  - Cell biology and animations: <http://www.johnkyrk.com/>
  - SIU histology: <http://www.siumed.edu/~dking2/>
  - Zoomified histology:  
<http://www.meddean.luc.edu/lumen/MedEd/Histo/virtualhistology.htm>

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    - (new but under construction)
  - <http://biomathematics.shis.uth.tmc.edu>
    - (old but already built)