

## 2 Biological background<sup>1</sup>

---

With P. Macklin

In this chapter, we present some of the key biological concepts necessary to motivate, develop, and understand the tumor models introduced in this book. We introduce the molecular and cellular biology of noncancerous tissue (Section 2.1) and then discuss how this biology is altered during cancer progression (Section 2.2). The discussion may in some areas be more detailed than is necessary for the models that we present; the intent is to offer a sampling of the rich world of molecular and cellular biology, helping the reader to consider how these and details may need to be incorporated in the work of cancer modeling. For greater depth on any of the topics, please refer to such excellent texts as [12] for molecular and cellular biology, as well as [386] for cancer cell biology.

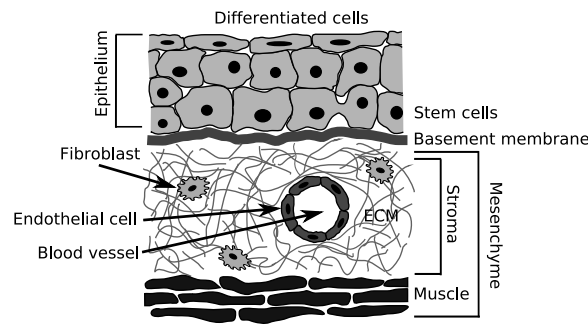
### 2.1 Key molecular and cellular biology

We focus upon the molecular and cellular biology of epithelial cells, the stroma, and the mesenchymal cells that create and maintain the stroma (Section 2.1.1). Specific and often anisotropic adhesive forces help to maintain tissue architecture (Section 2.1.2). Epithelial and stromal cells have the same basic subcellular structure (Section 2.1.3) and share much in common. They progress through a cell cycle when preparing to divide, can control their entry into and exit from the cycle, and can self-terminate (apoptose) when they detect irreparable DNA errors or other damage (Section 2.1.4). Their behavior is governed by a signaling network that integrates genetic and proteomic information with extracellular signals received through membrane-bound receptors (Section 2.1.5). Sometimes, cells respond to signaling events by moving within the stroma or along the basement membrane (Section 2.1.6). In pathologic conditions leading to hypoxia, cells can respond through a variety of mechanisms, or can succumb to necrosis; in some cases, necrotic cellular debris is calcified (Section 2.1.7).

<sup>1</sup> This introduction to cancer biology updates and expands the original exposition in [433].

### 2.1.1 Tissue microarchitecture and maintenance

*Epithelium* is composed of sheets of tightly-adhered epithelial cells that cover organ surfaces and often perform specialized functions. The epithelium is supported by the *stroma*, a loose connective tissue. The main component of the stroma is the *extracellular matrix* (ECM), a scaffolding of fibers (collagen, elastin, fibronectin, etc.) embedded in a mixture of water and glycoproteins. The ECM is secreted and maintained by *stromal cells*, specialized mesenchymal cells that can freely move within the stroma as they maintain the tissue; fibroblasts are the primary stromal cells in loose connective tissue (epithelial stroma). The stroma is interlaced by blood vessels, nerves, and lymphatic vessels, and it may rest on an additional layer of muscle or bone, depending upon the location. A thin, semi-permeable *basement membrane* (BM: a specialized type of ECM) separates the epithelium from the stroma. See Figure 2.1.



**Figure 2.1** Typical tissue structure showing epithelium separated from the stroma by a basement membrane.

This complex tissue structure is maintained by careful regulation of the cell population and a specific balance of adhesive forces. These processes are often tied together through cell signaling. For further information on tissue and organ structure, please see [220], [12], and the references therein.

#### Population dynamics:

Each cell type population must be regulated by balancing proliferation and apoptosis. When a differentiated cell dies, a *somatic stem cell* may divide either symmetrically into two new stem cells or asymmetrically into a stem cell and a *progenitor cell*. The progenitor cell either further divides or terminally differentiates into the desired cell type, migrates or is pushed to the correct position, and assumes its function. This process is tightly regulated by intercellular communication via biochemical signals (growth factors) and mechanics; stromal cells help maintain this signaling environment [425, 496, 728]. Each cell's response to the microenvironment is governed by surface receptors that interact with an internal signaling network. We note that stem cell dynamics are not fully understood; please see the excellent overviews in [73, 728].

### Epithelial cell polarity and adhesion:

Epithelium can be broadly classified as *simple* or *stratified* based upon its cell arrangement. In simple epithelium, cells are arranged in a single layer along the basement membrane. The cells are *polarized*, with a well-defined base adhering to the BM and an apex exposed to the *lumen* (e.g., a cavity in an organ); the apical side of the cell is often used to release secretory products. The epithelial cells adhere tightly to one another along their non-apical, non-basal sides. See Figure 2.2: left. In stratified epithelium, a single cell layer adheres to the BM (similarly to simple epithelium), with additional layers above. The cells in the upper layers adhere to the layers above and below them and tend to be flattened. See Figure 2.2: right. Overall, the careful orchestration of cell-BM and cell-cell adhesion helps determine the tissue geometry [352, 303, 688]. In fact, heterogeneities in the balance of cell-cell and cell-BM adhesion can lead to epithelium invagination [402], folding [675], and other nontrivial geometries [636]. The molecular mechanisms of adhesion are further explored in Section 2.1.2. More information on epithelial cell polarization can be found in standard biology texts, such as [12].

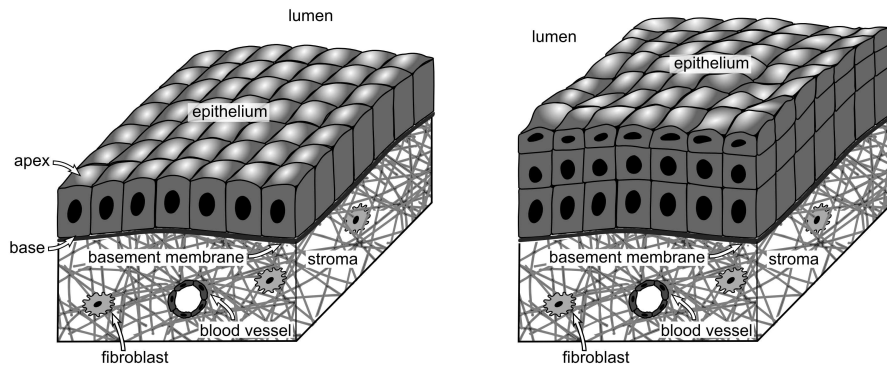


Figure 2.2 Simple (left) and stratified (right) cuboidal epithelium.

### Interaction between cell adhesion and population dynamics:

Cell adhesion and population dynamics are, in fact, linked to one another. Epithelial cell cycle progression and proliferation are controlled in part by cell-cell adhesion: when an epithelial cell is in (adhesive) contact with many neighbors, its cell cycle and proliferation are suppressed. This helps to maintain the epithelial cell population by reducing proliferation when the epithelium is fully populated, and by increasing proliferation near gaps in the epithelium (e.g., due to apoptosis) [144, 303, 688]. Hence, cell-cell contact-dependent proliferation helps prevent overproliferation. This theme is further discussed in Section 2.1.5.

Cell populations are also controlled by contact with the extracellular matrix and basement membrane. Polarized epithelial cells often become apoptotic after losing adhesive contact with the BM [246, 332, 278, 643, 680]; this specialized

type of apoptosis, termed *anoikis*, helps prevent overproliferation of unattached cells into the lumen [164]. The ECM also plays a major role in regulating stromal cells [278]. For example, ECM-bound proteoglycans control the proliferation, differentiation, and apoptosis of bone marrow stromal cells [70], and integrin ligands in the ECM regulate endometrial stromal cells [601].

### 2.1.2 Cellular adhesion and cell sorting

Adhesion is essential to multicellular arrangement and motility: cell-cell, cell-ECM, and cell-BM adhesion are responsible for maintaining the tissue arrangement, while cell-BM and cell-ECM are essential for traction during motility.

#### **Adhesion**

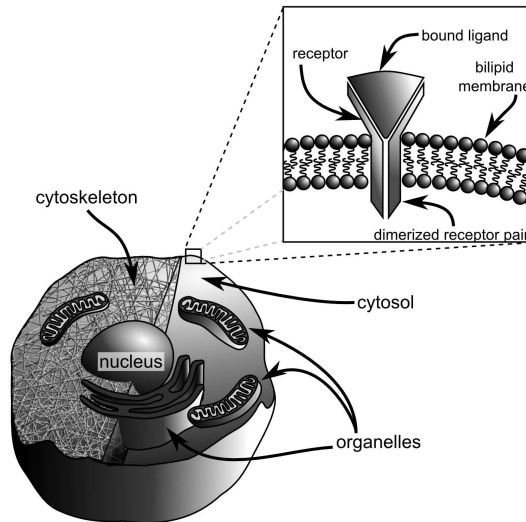
Cells can exhibit both homophilic and heterophilic adhesion. In *homophilic* adhesion, adhesion receptor molecules on the cell surface bond to identical ligands (a receptor's "target" molecules) on neighboring cells (in cell-cell adhesion) or in the microenvironment (in cell-ECM or cell-BM adhesion). This is the mode of E-cadherin-mediated cell-cell adhesion in epithelial cells, including carcinoma [524]. In *heterophilic* adhesion, surface adhesion molecules of one type bond to unlike ligand molecules in the extracellular matrix, on the basement membrane, or on neighboring cells. Cell-ECM and cell-BM adhesion are heterophilic between integrin molecules on the cell surface and ligands such as laminin and fibronectin in the microenvironment [92]. Heterophilic cell-cell adhesion is also observed, such as in T-cell lymphocytes via immunoglobulin-integrin bonds [633, 657, 429].

#### **Cell adhesion and cell sorting**

While epithelial cell-cell adhesion is generally homophilic and mediated by E-cadherin, other cadherins complicate the picture. For example, E-cadherin binds with greatest strength and specificity to E-cadherin, but can also bind to N-cadherin [524] and certain integrins [363]. Hence, the mixture of adhesion molecules on two cells' surfaces (and the specificity and kinetics of the bonds between the molecules) will determine the strength of their adhesion. Adhesive differences between cell types can lead to self-sorting behavior based upon adhesion gradients, which contributes to epithelial cell organization in tissues [554]. Such cell sorting has been observed experimentally [43].

### 2.1.3 Subcellular structure

A cell is composed of a well-defined nucleus containing its DNA, surrounded by cytosol (the liquid in the cell) and enveloped in a bilipid cell membrane. The cytoplasm contains organelles that carry out the cell's functions, such as the mitochondria (which synthesize adenosine triphosphate (ATP) from glucose and oxygen to provide energy to the cell) and endoplasmic reticulum (which provides ideal conditions for protein synthesis, folding, and transport), all sup-

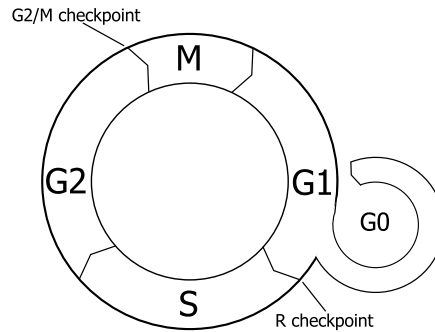


**Figure 2.3 Diagram of a eukaryotic cell:** A bilipid cell membrane contains the cytosol, nucleus, and organelles, all supported by a cytoskeleton. *inset:* Membrane-embedded receptors transmit microenvironmental information to the cell interior.

ported by a *cytoskeleton* of microtubules and actin polymer fibers. See Figure 2.3. The bilipid membrane separates the cell from the microenvironment. It is permeable to passive diffusion of small molecular species such as oxygen and glucose, actively pumps other molecular species (e.g., potassium and sodium) to maintain the cell's internal pH and chemical composition, and is impermeable to other, larger molecules such as growth factors. Embedded in the membrane are a variety of macromolecules that pump smaller molecules (e.g., potassium) against gradients; exchange mechanical forces with the extracellular matrix, basement membrane, and other cells; and transmit microenvironmental information to the cell interior.

#### 2.1.4 Cell cycle, proliferation, and apoptosis

Cell division is regulated by a highly regimented series of stages known as the *cell cycle*. In the first stage in the cell cycle, G1 (gap 1), the cell physically grows, proteins are synthesized, new organelles are constructed, and the cell prepares for DNA replication. In the following S (synthesis) phase, the DNA is copied, and in the G2 (gap 2) phase, final preparations are made within the cell nucleus for the division of the cell. In the final M (mitosis) phase, the two copies of the DNA are separated and incorporated into two nuclei (*mitosis*), and the cytoplasm and the organelles are divided into two daughter cells (*cytokinesis*). See Figure 2.4.



**Figure 2.4** The cell cycle.

The cell cycle contains numerous checkpoints that allow the cell to check for and repair DNA damage, as well as to control or halt cycle progression. At the R (restriction) checkpoint late in the G1 phase, the cell either commits to division (and progresses to the S phase) or exits the cell cycle (and enters the G0 *quiescent state*) [720, 72]. Most noncancerous somatic cells stay in this “resting” state due to microenvironmental signals received prior to the R checkpoint, maintaining homeostasis; after the R checkpoint, cells are committed to division and are less responsive to environmental signals to halt the cycle [609].

There are numerous checkpoints in the S and G2 phases to detect and repair DNA damage (e.g., between G2 and M). See Figure 2.4. Cells that fail to repair DNA damage at such checkpoints induce apoptosis [140]. In the process, “executioner” proteins (Caspases) in the cytoplasm break down the organelles, degrade the cytoskeleton, and fragment the DNA. The cell shrinks, and the degraded cell contents are released as harmless (i.e., chemically inert) vesicles known as *apoptotic bodies*, which are ingested (phagocytosed) by specialized immune cells as well as neighboring epithelial cells [371, 391].

A cell’s speed cell cycle progression is regulated by the production and balance of internal chemical signals, principally *cyclins* and *cyclin-dependent kinases* (CDKs). Surface receptors help control gene expression levels through complex signaling pathways. The gene expression pattern, in turn, determines the production and balance of proteins (including cyclins and CDKs). Hence, cell cycle progression is regulated by a complex interaction between the cell’s internal biomachinery and its surrounding environment [140].

### 2.1.5 Genetics, gene expression, and cell signaling

#### **Oncogenes and tumor suppressor genes**

The correct interpretation of growth and inhibitory signals is key to maintaining healthy tissues. If the cell receives both growth-promoting and -inhibiting signals, its behavior is determined by the balance of the signals and the resulting gene expression pattern. Two types of genes are particularly relevant to regulating cell

proliferation. *Oncogenes* respond to or create growth signals and promote cell cycle progression. *Tumor suppressor genes* (TSGs) respond to inhibitory signals, retard or halt the cell cycle, ensure proper DNA repair, and may trigger apoptosis under certain circumstances. Cancer initiation, or *carcinogenesis*, starts with the malfunction of one or more of these types of genes [302].

Genetic mutations can cause overactivity in oncogenes and impair the function of tumor suppressor genes. Sometimes, a single uncorrected point mutation is sufficient to affect the function of an oncogene [451] or functionally neutralize a tumor suppressor gene [325]. In other cases, cell division errors (e.g., during M phase) can create a mutant fusion oncogene, where the protein coding portion of an oncogene is mistakenly fused with the triggering portion of another, frequently expressed gene. As a result, signals are “misrouted” to the oncogene, thus boosting its activity. See [396], which describes the activation of the *MYC* oncogene by translocation with an immunoglobulin gene.

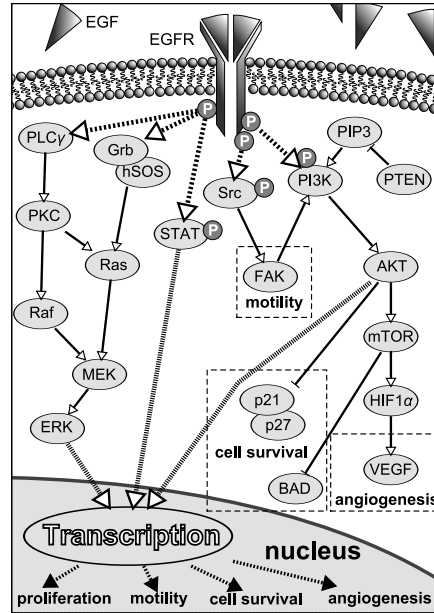
Other errors during cell division may cause a daughter cell to mistakenly receive extra copies of an oncogene (e.g., [111, 119, 217]) or too few copies of a TSG. Because normal cells possess two copies of each tumor suppressor gene, both copies must be damaged for a total loss of function of the gene. (See the Knudson two-hit model [388, 389], which led to the first discovered TSG [245].) While the probability of independent mutations in both copies of the TSG is ordinarily small, *loss of heterozygosity* (two damaged copies of the TSG are passed to a daughter cell) can significantly accelerate the process [503]. Furthermore, the loss of just one TSG copy can significantly impair its activity and increase the probability of completing a multi-step carcinogenesis pathway [558].

### Changes in gene expression

Gene expression is essential to maintaining proper cell function. Recent research has examined the over- and underexpression of genes, rather than outright genetic damage, as a potential contributor to unchecked cell proliferation. Viral infections (e.g., human papillomavirus can induce cervical cancer [695]) and microenvironmental signals (e.g., hypoxia; see Section 2.1.5 and references therein) can also induce changes in gene expression. Because gene expression patterns can be heritable, such changes can potentially affect a cell’s malignant transformation (e.g., by disabling a tumor suppressor gene) in the same way as a genetic mutation [356]. Lastly, we note that the biochemistry of gene expression is very complicated and is beyond the scope of this introduction; see [357, 356, 425, 191] for more on this topic.

### Cell signaling networks

Gene expression is controlled by cell surface receptors after activation by various signaling factors. Internal chemical species (e.g., oxygen) can also affect gene expression. The cell integrates such information with its genetic and proteomic state using a complex signaling network to determine its phenotype. Aberrant



**Figure 2.5 Simplified EGFR Signaling:** Dimerized EGFR can transmit signals through a variety of molecular pathways that trigger proliferation, motility, and increased resistance to apoptosis

cell signaling is often implicated in cancer, making it a key topic to molecular and cellular cancer biology. We illustrate with a few examples:

*Example: HIF-1 $\alpha$  signaling:*

A cell's response to hypoxia (low oxygen levels) is a key example of how internal protein levels can affect gene expression without need for additional receptor signaling. All cells create HIF-1 $\alpha$  (a hypoxia-inducible factor) that is ordinarily degraded in the presence of oxygen [87, 602, 247, 289]. When a cell experiences hypoxia, HIF-1 $\alpha$  accumulates and activates downstream "target" genes. Among targets of importance to cancer biology, HIF-1 $\alpha$  upregulates motility, secretion of angiogenic-promoting factors, and anaerobic glycolysis (an inefficient metabolism attained by reacting glucose with glucose, rather than oxygen); downregulates cell-cell and cell-ECM adhesion; and reduces sensitivity to apoptotic signals [305, 716, 13, 543]. We discuss the significance of this signaling pathway in cancer biology in Sections 2.1.7 and 2.2.2.

*Example: EGF signaling:*

Epidermal growth factor (EGF) can bind to and subsequently activate EGF receptors (EGFR). When two activated EGFRs bind to one another (dimerize), they can transmit signals leading to increased HIF-1 $\alpha$  secretion, increased cell proliferation, increased cell motility, and reduced sensitivity to apoptosis. See



Figure 2.5 and the excellent reviews in [314, 505, 134]. Malfunctions in this signaling process have been implicated in several cancers. For example, a mutant form of EGFR (HER2) commonly found in breast cancer is constitutively (i.e., permanently) active and does not require EGF binding for signaling activity; moreover, HER2 can bind to activated EGFR to provide a “shortcut” in the EGFR signaling cascade and thus increase EGFR signaling activity [192, 134]. In non-small cell lung carcinoma (NSCLC), downstream targets of EGFR are often mutated, most notably a constitutively-active form of K-ras that can function independently of upstream EGFR signals. Indeed, NSCLC with K-ras mutations are generally resistant to therapies that target EGFR [193, 525]. Both these mutations effectively activate downstream targets of EGFR independent of receptor activity; i.e., the EGFR pathway switch is “stuck in the ON position,” leading to excessive proliferation and other cancer-promoting activity.

*Example: E-Cadherin/ $\beta$ -Catenin signaling:*

Some receptors have multiple, simultaneous roles. E-cadherin mediates homophilic epithelial cell-cell adhesion (Section 2.1.2). The intracellular domain of E-cadherin binds to  $\alpha$ -catenin (using  $\beta$ -catenin as an adapter protein) to mechanically couple an adhered cell to its actin cytoskeleton [387, 189]. Ligated E-cadherin also binds to  $\beta$ -catenin, which sequesters it at the cell membrane and prevents its downstream signaling. Unsequestered  $\beta$ -catenin would otherwise promote cell cycle progression by triggering transcription of Cyclin D1, c-myc, and Axin2. Hence, E-cadherin not only plays a mechanical role in cell-cell interactions, but also a signaling role by inhibiting cell cycle progression when physically adhered to epithelial cells [71, 600, 430, 315]. This signaling pathway plays a key role in maintaining normal epithelial tissue microarchitecture [144, 303, 688]; see Section 2.1.1. In many cancers (e.g., breast cancer [419]), the E-cadherin/ $\beta$ -catenin signaling pathway can be disrupted, leading to increased downstream oncogenic activity (e.g., increased cell cycle progression due to Cyclin D1 overexpression [464]).

### 2.1.6 Cell motility

Motile cells demonstrate directed motion by a complex interaction between cell signaling, their cytoskeleton, and adhesion with the ECM or BM. We describe here the key aspects of this process; more detail can be found in [145, 280, 405].

Gradients in microenvironmental signaling molecules (e.g., EGF) can be amplified by the multiple steps in signaling networks, leading to pronounced interal signaling gradients [379]. A key downstream effect of motility signaling is actin polymerization (the formation of linked chains of actin monomer that extend the actin cytoskeleton) and depolymerization (the spontaneous degradation of actin polymers). This process takes place within a thin region just below the cell membrane [359, 405]. Wherever polymerization exceeds depolymerization, there is net outward growth of the cell’s cytoskeleton, which, in turn, deforms and

extends the cell membrane. If net actin polymerization continues in a consistent direction, the cell forms a pseudopod (i.e., a “false foot”) that extends from its leading edge into the microenvironment. Net actin depolymerization at the cell’s trailing edge, along with internal microtubule activity, leads to cell contraction [145, 280, 405]. The signaling network creates and maintains this bias in actin polymerization. For example, dimerized EGFR can activate Src, which, in turn, can mediate the formation of Arp2/3-N-WASP complexes that nucleate actin polymerization; microenvironmental EGF gradients thus create internal polymerization gradients towards the cell’s leading edge [596, 145, 481, 692, 691].

Cell motility requires mechanical interaction between cell membrane protrusions and the microenvironment. Individual cells may move through the stroma (in 3D) in an amoeboid motion by squeezing between ECM fibers (e.g., T-lymphocyte migration [703]) or by extending a slender, finger-like pseudopod (a *filopodium*) that forms focal adhesions with the ECM to exert traction [405]. The latter, which occurs during cancer cell invasion of the stroma [235, 702], requires directed, coordinated degradation of the ECM to create space for motion, and is accomplished by forming tiny invadopodia on the filopodium surface that secrete proteases to degrade the ECM [367, 174, 687]. In other cases, cells may move along a surface by extending a sheet-like pseudopod (a *lamellipodium*) that focally adheres to the surface for traction [405]. This has been observed in Paget’s disease of the breast (cancerous epithelial cells chemotax along the breast duct basement membrane towards the nipple [78]), wound healing (keratinocytes crawl along the top of granular tissue [403]), and fibrosarcoma metastasis (cancer cells crawl along lymph vessel walls [712]). Following membrane protrusion, non-amoeboid motility requires the release of integrin bonds along the cell’s trailing edge and subsequent cell contraction, allowing net forward motion [405]. Directed cell motility also requires active intracellular transport of actin monomer [689], integrins [280], and other cytoskeletal components between the cell’s trailing and leading edges [405].

### 2.1.7 Hypoxia, necrosis, and calcification

In Section 2.1.5, we discussed some of the cellular adaptations to hypoxia. Sustained hypoxia (as well as sustained hypoglycemia), such as that encountered in ischemic tissue [385, 252, 583] and in larger tumors [114, 214], can lead to ATP depletion and consequently cell death. This unplanned cell death is referred to as *necrosis*.

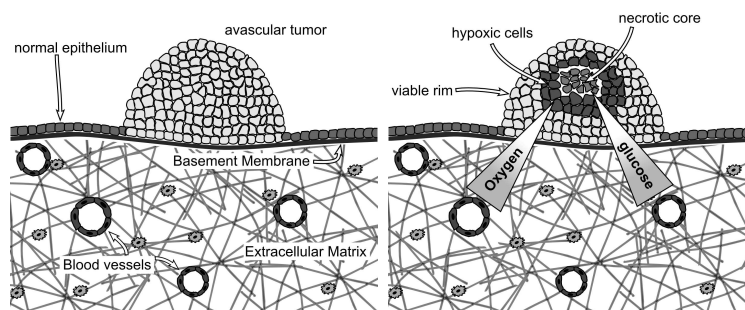
When a cell becomes necrotic, its surface ion pumps cease to function, resulting in osmosis of water into the cell, cell swelling, and subsequent bursting [51]. This differs from apoptosis, where the volume loss is orderly and the intracellular contents are contained in apoptotic bodies [51]. In necrotic cells, the remaining solid cell fraction is generally not phagocytosed by surrounding cells, as they themselves are typically also necrotic. In some cancers (e.g., breast cancer [641], liver cancer [310], ovarian cancer [631], and lymphoma [336, 137]) and other

pathologic conditions (e.g., tuberculosis [54] and abscesses [398, 701]), necrotic tissue can undergo calcification: the solid cell components are replaced by calcium phosphate and/or calcium oxalate molecules that bond together to form calcite crystals that grow into hard *microcalcifications* [446].

## 2.2 The biology of cancer

Most simply stated, cancer occurs when defective genes cause cells to malfunction and interact with the body in an aberrant, hyperproliferative manner (either by increased cell proliferation or reduced cell apoptosis). We now examine how the molecular and cellular biology previously introduced in Section 2.1 can break down, leading to cancer. Our discussion primarily focuses upon *carcinoma* (cancers arising from epithelial cells) rather than *sarcoma* (cancers arising from mesenchymal cells).

### 2.2.1 Carcinogenesis



**Figure 2.6** **Left:** Initial avascular tumor. **Right:** Substrate gradients lead to hypoxia and central necrosis

Carcinogenesis is a multistage process thought to begin with a genetic mutation or epigenetic alteration that overexpresses an oncogene or underexpresses a tumor suppressor gene in one or a small number of cells. If the cell survives and the mutation escapes its DNA repair mechanisms, the cell (or its descendants) may over time acquire further mutations to ignore growth-inhibiting signals from its neighbors, bypass its internal controls and checkpoints, and form a colony of hyperproliferative, aberrant cells. This accumulation of mutations may require years to progress, but can be accelerated by exposure to carcinogens and other harsh, DNA-damaging environmental effects.

Differentiated cells can only divide a limited number of times before reaching *senescence*: the point at which they permanently arrest in G0 or apoptose. Thus,

differentiated cells alone cannot drive unlimited tumor growth without additional mutations to overcome senescence. Recent studies suggest that cancer may arise from mutated somatic stem cells rather than differentiated cells [58, 425, 608]. In this scenario, the tumor is a mixed cell population whose overgrowth is driven by a small sub-population of cancer stem cells, rather than by differentiated cells that have overcome senescence. With or without cancer stem cells, the result is the same at the multicell and tissue scales: a mass of hyperproliferative cells that fail to respond to ordinary physiologic limits to their growth (Figure 2.6: left).

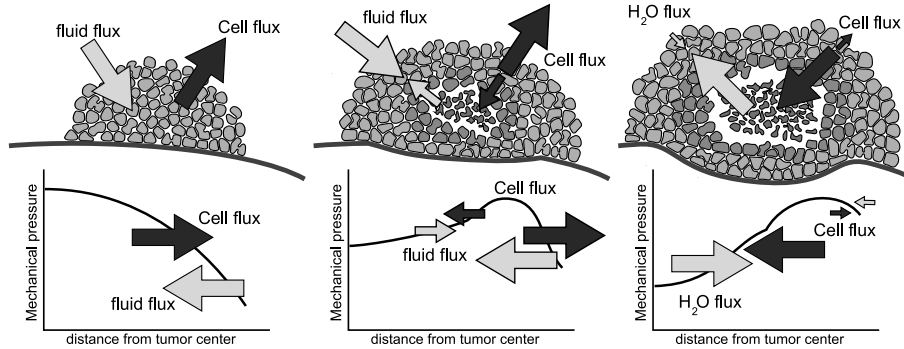
### 2.2.2 Avascular solid tumor growth

Once a tumor has established a foothold in its host tissue, it begins an early period of growth as it becomes an *in situ* cancer. Epithelial cells are generally constrained by the basement membrane.

#### **The limiting role of oxygen and nutrient diffusion, hypoxia, and necrosis**

In this early stage of cancer, the tumor has no vascular system of its own, and so it must rely upon the host vasculature in the nearby stroma for crucial oxygen, nutrients, and growth factors; we refer to these generically as “substrates.” Substrates diffuse from the surrounding vascularized tissue, enter the tumor, and are uptaken by proliferating tumor cells. This motion of substrates from external sources (the host vasculature) to internal sinks (the metabolically active tumor cells) causes substrate gradients to form within the tumor. Of particular importance is oxygen, which generally diffuses on the order of 100–200  $\mu\text{m}$  into tissue before dropping to levels insufficient for cellular metabolism [114, 151, 214, 437, 439]. Interior tumor cells experience hypoxia and respond to their harsher microenvironment in a variety of ways (Section 2.1.5). Deeper within the tumor, oxygen and glucose levels drop to critically low levels that cause the tumor cells to necrose. These dynamics are manifested as an outer tumor viable rim of proliferating cells, an interior band of hypoxic cells, and a central necrotic core. See Figure 2.6: right.

This affects the tumor mechanically. Prior to the formation of a necrotic core, proliferation throughout the tumor causes a net outward cell flux that expands the tumor (Figure 2.7: left). Simultaneously, the proliferating tumor cells absorb fluid from the interstitium to fuel their growth and eventual division, resulting in a net fluid flux into the tumor. Once a necrotic core has formed, cell lysis reduces the tumor cell volume and releases fluid that leaves the necrotic core and enters the proliferative rim interstitium. The subsequent reduction in mechanical pressure in the necrotic core redirects some of the viable rim cell flux towards the tumor interior (Figure 2.7: middle). As the tumor grows, the volume of its necrotic core increases, thus accentuating its cell volume sink effect. Once the tumor grows large enough, the cell flux resulting from proliferation balances with the fluid flux stemming from necrosis, leading to zero outward cell flux. This gives rise to a steady-state tumor spheroid (Figure 2.7: right).



**Figure 2.7** Cell and fluid flux in early (**left**), later (**middle**), and long-time (**right**) tumor growth.

### 2.2.3 Interaction with the microenvironment

As the nascent tumor grows in its host tissue, it interacts with the surrounding microenvironment in a variety of ways. It mechanically displaces and compresses the surrounding tissue, including the basement membrane (Figure 2.7: middle and right). The tumor degrades and remodels the extracellular matrix (ECM), both biomechanically and biochemically by the secretion of matrix degrading enzymes such as matrix metalloproteinases (MMPs) that degrade the ECM. The degraded ECM, in turn, can release ECM-associated growth factors that fuel further tumor growth [647]. The degradation of the ECM by the MMPs increases the ability of the tumor to push into the surrounding tissue, both by reducing the mechanical rigidity of the surrounding tissue and by creating extra space for the growing tumor [327]. The combination of proliferation-induced pressure and proteolytic degradation of the surrounding tissue results in *tissue invasion*: the invasion of sheets or “fingers” of tumor cells into the surrounding tissue along paths of least mechanical resistance. *Acidosis* (a decreased microenvironmental pH resulting from anaerobic glycolysis in hypoxic tumor cells) has also been hypothesized to play a role in tumor invasion, by inducing apoptosis in the surrounding normal epithelium, by giving invasive tumor cells a selective advantage over tumor cells that have not adapted to acidity, and by contributing to ECM degradation (due to proteases released by apoptotic cells) [283, 529, 264, 266, 629, 265, 630, 627, 282, 281, 268, 628, 209, 284, 267].

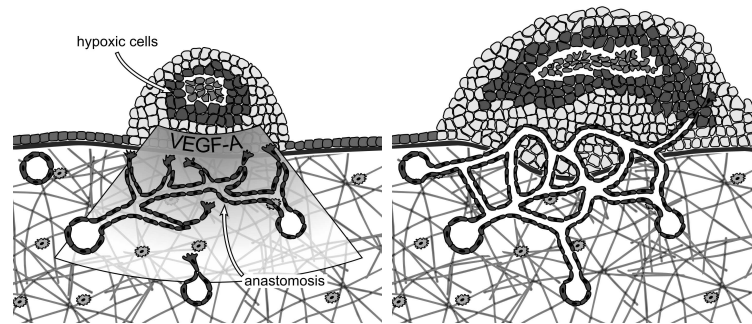
There is recent evidence that tumors induce changes in gene expression in the nearby stroma that help sustain tumor growth [328, 728]. For instance, carcinomas may release signaling molecules (e.g.,  $IL-1\beta$ ) that stimulate fibroblasts to secrete hepatocyte growth factor (HGF). The HGF, in turn, promotes tumor cell growth, decreases cell-cell adhesion, and increases MMP secretion [459]. Tumors may also alter gene expression in nearby, non-cancerous epithelial cells [335].

## 2.2.4 Vascular growth and metastasis

The next stage in cancer development can be viewed as a response to hypoxia. The ultimate result is *angiogenesis*, where the tumor induces endothelial cells to form a new vasculature that directly supplies the tumor with the nutrients, enabling further expansion. Some of the same mechanisms responsible for angiogenesis play a role in *metastasis*, the spread of tumor cells to distant locations.

### Angiogenesis

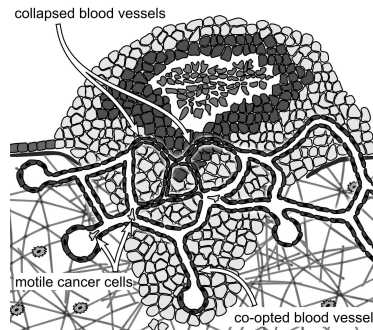
As discussed in Section 2.1.5, hypoxia-inducible factors (e.g., HIF-1 $\alpha$ ) accumulate in hypoxic cells, which can trigger numerous downstream genetic targets. In particular, the hypoxic cells secrete tumor angiogenic growth factors (TAFs) such as vascular endothelial growth factor (VEGF) [716, 364, 13, 543]. These TAFs diffuse outward from the hypoxic regions of the tumor and eventually reach nearby blood vessels. See Figure 2.8: left.



**Figure 2.8 Left:** Angiogenic growth factors such as VEGF-A are secreted by hypoxic tumor cells, leading to angiogenesis. **Right:** The fresh nutrient supply allows for renewed tumor expansion.

Blood vessels are composed of tightly connected squamous (flat and scale-like) endothelial cells that are surrounded by a basement membrane and other supporting cells, including smooth muscle cells and pericytes [426]. When the endothelial cells detect the TAF gradient emanating from the tumor, they secrete MMPs that degrade the basement membrane and extracellular matrix [48] (Figure 2.8: left). This allows the endothelial cells to migrate away from the blood vessel and toward the TAF source in the tumor. The leading endothelial cells are referred to as *sprout tips*; immediately behind the sprout tips, other endothelial cells divide, migrate, align, and form tubes of polarized endothelial cells surrounding a vascular lumen [494]. The vessels then link with one another to form a network of loops in a process called *anastomosis* (Figure 2.8: left). It can take on the order of 10 to 21 days for new vessels to form and connect to the parent vessels [285, 48, 489].

The end result is a *neovasculature* that provides the tumor with a direct supply of oxygen and nutrients. The configuration of the neovasculature is determined by the balance of pro- and anti-angiogenic growth factors, as well as by the mechanical pressures from the growing tumor and flow stresses within the nascent blood vessels [409, 653, 557, 287, 215]. The fresh nutrient supply allows a new stage of rapid tumor growth into the surrounding tissue (Figure 2.9).



**Figure 2.9** Invasive tumor growth into the stroma. The tumor grows to co-opt the neovasculature, leading to collapse of some vessels and renewed hypoxia.

Angiogenesis is not unique to tumor growth, but is also a key part of wound healing, the menstrual cycle, and embryonic development [114, 214]. However, we note that tumor angiogenesis is pathological in nature, and the resulting vasculature is inefficient in a number of ways: the vessels are often “leaky” due to large gaps between endothelial cells; the newly formed vessels are not as stiff and rigid as mature vessels and may collapse when subjected to tissue stress (such as that created by rapidly growing tumors); the basement membrane around the new vessels may not be fully formed; some of the newly formed vessel walls may be composed of a mosaic of tumor and endothelial cells; and the tumor neovascular network tends to be much more tortuous than regular vascular networks [218, 114]. See Figure 2.9. This inefficiency may hinder drug delivery within tumors [342, 623], as well as lead to the development of new hypoxic regions within the tumor and additional sessions of angiogenesis.

### Tissue invasion and metastasis

A particularly damaging aspect of advanced cancer is *metastasis*, the spread of tumor cells to form secondary tumors in distant locations. Metastasis occurs most commonly in breast, prostate, and lung cancers [66], and it is estimated that over 90% of all deaths from solid tumors result from metastasis [300]. In spite of the great clinical importance of metastasis, it is poorly understood [362].

Metastasis is a complex phenomenon involving several mechanisms that are closely related to tissue invasion. Genetic instability, intrinsic limits (e.g., senescence), and extrinsic selective pressures (e.g., limited nutrients,

immune system attacks) lead to competition within heterogeneous tumor cell populations and the eventual selection for pro-metastatic genes [300]. Hypoxia creates a strong selective pressure, leading to increasing internal HIF-1 $\alpha$  levels in the tumor cells and the expression of genes responsible for increased motility, glycolysis, reduced response to apoptotic pathways, and increased production of MMPs [305]. The selective pressures also lead to increased expression of genes responsible for locomotion [544]. As a result, tumor cells degrade the BM and ECM and invade the stroma, either individually, as small clumps of cells (emboli), or in cohort motion of sheets of cells linked by cell-cell adhesion [490, 300]. Eventually, invasive tumor cells can enter the vasculature or lymphatic system. See Figure 2.9.

For sarcomas (which already reside in the stroma), this is accomplished by the proteolytic degradation of the ECM and BM surrounding the stromal vessels, followed by direct entry into the vessels. For carcinomas (which are separated from the stroma by the BM), entry into the vasculature could also indirect via the lymphatic system [200]. The mesenchymally-derived sarcoma cells move with built-in cellular machinery in a contractile manner: by first degrading the ECM on their leading edge, adhering to the ECM, and contracting, followed by rebuilding the ECM on the trailing edge [672]; see Section 2.1.6. Epithelial-derived carcinoma cells initially lack this locomotive ability, but mutations and altered gene expression can restore these locomotive mechanisms; the process is often referred to as the *epithelial-mesenchymal transition* (EMT) [544, 672].

Once the metastatic tumor cells have reached the vasculature, they circulate in the blood. Initially, survival of the circulating tumor cells is inhibited by the immune system, which kills most of the individual cells; emboli consisting of 5 to 10 cells are more likely to escape attack by the immune system [200]. Note that the complex role of the immune system is poorly understood and may both promote and inhibit metastasis. Circulating tumor cells that do survive can eventually lodge in the capillary bed of distant organs; the most frequent destinations include the liver, lungs, and bones [66].

However, without further tumor-host interaction, the destination microenvironment will not support the newly arrived metastatic tumor cells. Different types of tumor cells tend to metastasize to specific tissues. This “seed and soil” idea, that only specific tissues are suitable to each tumor cell line, was first formulated by Stephen Paget in 1889 when studying breast cancer metastases [519, 172, 487]. The reasons for this are only now being elucidated in an emerging area of cancer research. The theory is that tumors release cytokines, VEGF, and other chemical signals into the circulatory system that recruit progenitor and endothelial cells from the bone marrow and vasculature to assist in creating a *pre-metastatic niche*: a modified microenvironment in a distant host tissue that is suitable for tumor metastasis [300]. In the process, the chemical signals alter the gene expression in the endothelial cells in capillary walls at the destination tissue, which then express additional adhesion molecules and secrete MMPs to



degrade the basement membrane surrounding the capillaries [316, 200, 362]. The newly-expressed adhesion molecules on the inner surface of the capillary bed improve the ability of the metastatic tumor cells to arrest at the destination, and the degraded BM assists in the extravasation of the tumor cells from capillaries into the destination tissue.

Once the metastatic tumor cells successfully invade the destination tissue, they secrete growth factors that induce additional changes in the new location. Growth is similar to the mechanisms of tissue invasion that were introduced earlier, but with additional elements. Tumor-induced changes in the stromal cells cause them to degrade and remodel the matrix, even as the tumor cells also secrete MMPs to degrade the matrix. Growth-promoting molecules that were previously sequestered in the ECM fuel further tumor growth [200]. With ample room to grow and a favorable microenvironment, these tumor cells can develop into secondary tumors. Because the metastatic tumor cells have been selected for their invasive phenotype, they are capable of expressing pro-angiogenic growth factors to initiate angiogenesis and enter vascularized growth. The tissue specificity of this process is likely due to the combination and balance of cytokines and chemicals secreted by the tumors, which, in turn, depends upon the genetic makeup of the tumors [626]. It is thought that only a small fraction of the cells in the primary tumor have the ability to recruit the proper progenitor and endothelial cells to build the pre-metastatic niche [300].

The scientific understanding of metastasis is advancing rapidly, and the reader is encouraged to read the reviews by [200, 300, 362, 521]. The reviews on bone metastases by [421, 66] provide well-written, concrete examples of the process, and they give an excellent overview of the latest in metastasis research.

### 2.3 Concluding remarks

In this chapter, we presented a simplified overview of the major topics in biology that relate to cancer. Cancer modelers may wish to keep these topics in mind as they study and extend the models presented in this book, and to explore the excellent references cited in this chapter and elsewhere to learn more about these biological themes in greater depth. In the following chapters of Part I, we present state-of-the-art continuum, discrete, and hybrid models that incorporate a broad spectrum of the tumor progression and behavior presented in this chapter.

## References

- [1] A. C. Abajian and J. S. Lowengrug. An agent-based hybrid model for avascular tumor growth. *UCI Undergrad. Res. J.*, 11, 2008.
- [2] R. Abbott, S. Forrest, and K. Pienta. Simulating the hallmarks of cancer. *Art. Lif*, 12(4):617–34, 2006.
- [3] H. Acker, J. Carlsson, W. Mueller-Klieser, and R. Sutherland. Comparative po2 measurements in cell spheroids cultured with different techniques. *Br. J. Cancer*, 56:325–327, 1987.
- [4] D. Adalsteinsson and J. A. Sethian. The Fast Construction of Extension Velocities in Level Set Methods. *J. Comput. Phys.*, 148(1):2–22, 1999.
- [5] J. Adam. General aspects of modeling tumor growth and the immune response. In J. Adam and N. Bellomo, editors, *A Survey of Models on Tumor Immune Systems Dynamics*, pages 15–87. Birkhaeuser, Boston, 1996.
- [6] T. L. Adamovich and R. M. Simmons. Ductal carcinoma in situ with microinvasion. *Am. J. Surg.*, 186(2):112–6, 2003.
- [7] B. Addison-Smith, D. McElwain, and P. Maini. A simple mechanistic model of sprout spacing in tumour-associated angiogenesis. *J. Theor. Biol.*, 250:1–15, 2008.
- [8] L. Ai, W.-J. Kim, T.-Y. Kim, C. R. Fields, N. A. Massoll, K. D. Robertson, and K. D. Brown. Epigenetic silencing of the tumor suppressor cystatin m occurs during breast cancer progression. *Canc. Res.*, 66:7899–909, 2006.
- [9] T. Alarcón, H. Byrne, and P. Maini. A cellular automaton model for tumour growth in inhomogeneous environment. *J. Theor. Biol.*, 225:257–274, 2003.
- [10] T. Alarcón, H. Byrne, and P. Maini. A multiple scale model for tumor growth. *Multiscale Model. Sim.*, 3:440–475, 2005.
- [11] M. Alber, M. Kiskowski, J. Glazier, and Y. Jiang. On cellular automaton approaches to modeling biological cells. In R. Rosenthal and D. Gilliam, editors, *IMA Series on Mathematical systems theory in biology, communication and finance*, volume 142, pages 1–40. Springer, New York, 2002.
- [12] B. Alberts, A. Johnson, J. Lewis, M. Raff, K. Roberts, and P. Walter. *Molecular Biology of the Cell*. Garland Science, New York, fifth edition, 2007.
- [13] J. W. Allen, S. R. Khetani, R. S. Johnson, and S. Bhatia. *In Vitro* Liver Tissue Model Established from Transgenic Mice: Role of HIF-1alpha on Hypoxic Gene Expression. *Tissue Engineering*, 12(11):3135–47, 2006.
- [14] M. Amar and A. Goriely. Growth and instability in soft tissues. *J. Mech. Phys. Solids*, 53:2284–2319, 2005.
- [15] D. Ambrosi, F. Bussolino, and L. Preziosi. A review of vasculogenesis models. *Comp. Math. Meth. Med.*, 6:1–19, 2005.

- 
- [16] D. Ambrosi, A. Duperray, V. Peschetola, and C. Verdier. Traction patterns of tumor cells. *J. Math. Biol.*, 58(1-2):163–81, 2009.
- [17] D. Ambrosi, A. Gamba, and G. Serini. Cell directional and chemotaxis in vascular morphogenesis. *Bull. Math. Biol.*, 66:1851–1873, 2004.
- [18] D. Ambrosi and F. Guana. Mechanical aspects of growth in soft tissues. *Boll. Unione Mat. Ital.*, 7:775–781, 2004.
- [19] D. Ambrosi and F. Guana. Stress-modulated growth. *Math. Mech. Solids*, 12:319–343, 2007.
- [20] D. Ambrosi, A. Guillou, and E. DiMartino. Stress-modulated remodeling of a non-homogeneous body. *Biomech. Model. Mechanobiol.*, 7:63–76, 2008.
- [21] D. Ambrosi and F. Mollica. On the mechanics of a growing tumor. *Int. J. Eng. Sci.*, 40:1297–1316, 2002.
- [22] D. Ambrosi and F. Mollica. The role of stress in the growth of a multicell spheroid. *J. Math. Biol.*, 48:477–499, 2004.
- [23] D. Ambrosi and L. Preziosi. On the closure of mass balance models for tumor growth. *Math. Mod. Meth. Appl. Sci.*, 12:737–754, 2002.
- [24] D. Ambrosi and L. Preziosi. Cell adhesion mechanisms and elasto-viscoplastic mechanics of tumours. *Mech. Model. Mechanobiol.*, 8:397–413, 2009. 10.1007/s10237-008-145-y.
- [25] American Cancer Society. American cancer society breast cancer facts and figures 2007-2008. *Atlanta: American Cancer Society, Inc.*, 2007. <http://www.cancer.org/downloads/STT/BCFF-Final.pdf>.
- [26] A. Anderson. A hybrid mathematical model of solid tumour invasion: The importance of cell adhesion. *Math. Med. Biol.*, 22:163–186, 2005.
- [27] A. Anderson and M. Chaplain. Continuous and discrete mathematical models of tumor-induced angiogenesis. *Bull. Math. Biol.*, 60:857–900, 1998.
- [28] A. Anderson, M. Chaplain, E. Newman, R. Steele, and A. Thompson. Mathematical modeling of tumour invasion and metastasis. *J. Theor. Med.*, 2:129–154, 2000.
- [29] A. Anderson, M. Chaplain, K. Rejniak, and J. Fozard. Single-cell based models in biology and medicine. *Math. Med. Biol.*, 25(2):185–6, 2008.
- [30] A. Anderson and V. Quaranta. Integrative mathematical oncology. *Nature Reviews Cancer*, 8:227–244, 2008.
- [31] A. Anderson, K. Rejniak, P. Gerlee, and V. Quaranta. Microenvironment driven invasion: a multiscale multimodel investigation. *J. Math. Biol.*, 58:579–624, 2009.
- [32] A. Anderson, A. Weaver, P. Commmings, and V. Quaranta. Tumor morphology and phenotypic evolution driven by selective pressure from the microenvironment. *Cell*, 127:905–915, 2006.
- [33] E. Anderson. Cellular homeostasis and the breast. *Maturitas*, 48(S1):13–7, 2004.
- [34] M. Anderson, D. Srolovitz, G. Grest, and P. Sahini. Computer simulation of grain growth- i. kinetics. *Acta Metall.*, 32:783–791, 1984.
- [35] E. D. Angelis and L. Preziosi. Advection-diffusion models for solid tumour evolution in vivo and related free boundary problem. *Math. Models Meth. Appl. Sci.*, 10:379–407, 2000.
- [36] R. Araujo and D. McElwain. A history of the study of solid tumour growth: The contribution of mathematical modelling. *Bull. Math. Biol.*, 66:1039–1091, 2004.
- [37] R. Araujo and D. McElwain. A linear-elastic model of anisotropic tumor growth. *Eur. J. Appl. Math.*, 15:365–384, 2004.

- 
- [38] R. Araujo and D. McElwain. New insights into vascular collapse and growth dynamics in solid tumors. *J. Theor. Biol.*, 228:335–346, 2004.
- [39] R. Araujo and D. McElwain. A mixture theory for the genesis of residual stresses in growing tissues I: A general formulation. *SIAM J. Appl. Math.*, 65:1261–1284, 2005.
- [40] R. Araujo and D. McElwain. A mixture theory for the genesis of residual stresses in growing tissues II: Solutions to the biphasic equations for a multicell spheroid. *SIAM J. Appl. Math.*, 66:447–467, 2005.
- [41] R. Araujo and D. McElwain. The nature of the stresses induced during tissue growth. *Appl. Math. Lett.*, 18:1081–1088, 2005.
- [42] N. Armstrong, K. Painter, and J. Sherratt. A continuum approach to modeling cell-cell adhesion. *J. Theor. Biol.*, 243:98–113, 2006.
- [43] P. Armstrong. Light and electron microscope studies of cell sorting in combinations of chick embryo and neural retina and retinal pigment epithelium. *Willhelm Roux Archiv.*, 168:125–141, 1971.
- [44] S. Astanin and L. Preziosi. Multiphase models of tumour growth. In N. Bellomo, M. Chaplain, and E. DeAngelis, editors, *Selected Topics on Cancer Modelling: Genesis - Evolution - Immune Competition - Therapy*. Birkhaeuser, Boston, 2007.
- [45] C. Athale and T. Deisboeck. The effects of egf-receptor density on multiscale tumor growth patterns. *J. Theor. Biol.*, 238:771–779, 2006.
- [46] C. Athale, Y. Mansury, and T. Deisboeck. Simulating the impact of a molecular 'decision-process' on cellular phenotype and multicellular patterns in brain tumors. *J. Theor. Biol.*, 233:469–481, 2005.
- [47] H. Augustin. Tubes, branches, and pillars: The many ways of forming a new vasculature. *Circ. Research*, 89:645–647, 2001.
- [48] D. H. Ausprunk and J. Folkman. Migration and proliferation of endothelial cells in preformed and newly formed blood vessels during tumour angiogenesis. *Microvasc. Res.*, 14(1):53–65, 1977.
- [49] D. Balding and D. McElwain. A mathematical model of tumor-induced capillary growth. *J. Theor. Biol.*, 114:53–73, 1985.
- [50] A. Bardelli, M. Basile, E. Audero, S. Giordano, S. Wennström, S. Ménard, P. Comoglio, and C. Ponzetto. Concomitant activation of pathways downstream of grb2 and pi 3-kinase is required for met-mediated metastasis. *Oncogene*, 18:1139–1146, 1999.
- [51] L. F. Barros, T. Hermosilla, and J. Castro. Necrotic volume increase and the early physiology of necrosis. *Comp. Biochem. Physiol. A. Mol. Integr. Physiol.*, 130:401–9, 2001.
- [52] U. Bartels, C. Hawkins, M. Jing, M. Ho, P. Dirks, J. Rutka, D. Stephens, and E. Bouffet. Vascularity and angiogenesis as predictors of growth in optic pathway/hypothalamic gliomas. *J. Neurosurg.*, 104:314–320, 2006.
- [53] K. Bartha and H. Rieger. Vascular network remodeling via vessel cooption, regression and growth in tumors. *J. Theor. Biol.*, 241:903–918, 2006.
- [54] R. J. Basaraba, H. Bielefeldt-Ohmann, E. K. Eschelbach, C. Reisenhauer, A. E. Tolnay, L. C. Taraba, C. A. Shanley, E. A. Smith, C. L. Bedwell, E. A. Chlipala, and I. A. Orme. Increased expression of host iron-binding proteins precedes iron accumulation and calcification of primary lung lesions in experimental tuberculosis in the guinea pig. *Tuberculosis*, 88(1):69–79, 2008.
- [55] A. Bauer, T. Jackson, and Y. Jiang. A cell-based model exhibiting branching and anastomosis during tumor-induced angiogenesis. *Biophys. J.*, 92:3105–3121, 2007.

- [56] F. O. Baxter, K. Neoh, and M. C. Tevendale. The beginning of the end: Death signaling in early involution. *J. Mamm. Gland Biol. Neoplas.*, 12(1):3–13, 2007.
- [57] B. Bazaliy and A. Friedman. A free boundary problem for an elliptic-parabolic system: Application to a model of tumor growth. *Comm. Partial Diff. Eq.*, 28:517–560, 2003.
- [58] P. A. Beachy, S. S. Karhadkar, and D. M. Berman. Tissue repair and stem cell renewal in carcinogenesis. *Nature*, 432(7015):324–331, 2004.
- [59] E. Bearer, J. Lowengrub, Y. Chuang, H. Frieboes, F. Jin, S. Wise, M. Ferrari, D. Agus, and V. Cristini. Multiparameter computational modeling of tumor invasion. *Cancer Res.*, 69:4493–4501, 2009.
- [60] L. Bello, V. Lucini, F. Costa, M. Pluderi, C. Giussani, F. Acerbi, G. Carrabba, M. Pan-nacci, D. Caronzolo, S. Grosso, S. Shinkaruk, F. Colleoni, X. Canron, G. Tomei, G. Deleris, and A. Bikfalvi. Combinatorial administration of molecules that simul-taneously inhibit angiogenesis and invasion leads to increased therapeutic efficacy in mouse models of malignant glioma. *Clin. Cancer Res.*, 10:4527–4537, 2004.
- [61] N. Bellomo, E. de Angelis, and L. Preziosi. Multiscale modeling and mathematical problems related to tumor evolution and medical therapy. *J. Theor. Medicine*, 5:111–136, 2003.
- [62] N. Bellomo, N. Li, and P. Maini. On the foundations of cancer modelling: selected topics, speculations, and perspectives. *Math. Models Meth. Appl. Sci.*, 4:593–646, 2008.
- [63] N. Bellomo and L. Preziosi. Modelling and mathematical problems related to tumor evo-lution and its interaction with the immune system. *Math. Comput. Modelling*, 32:413–542, 2000.
- [64] M. Ben-Amar and A. Gorielly. Growth and instability in elastic tissues. *J. Mech. Phys. Solids*, 53:2284–2319, 2005.
- [65] R. Benjamin, J. Capparella, and A. Brown. Classification of glioblastoma multiforme in adults by molecular genetics. *The Cancer Journal*, 9:82–90, 2003.
- [66] J. R. Berenson, L. Rajdev, and M. Broder. Pathophysiology of Bone Metastases. *Cancer Biol. Ther.*, 5(9):1078–1081, 2006.
- [67] M. Berger and I. Rigoutsos. An algorithm for point clustering and grid generation. *IEEE Trans. Syst. Man. Cybern.*, 21:1278–1286, 1991.
- [68] H. Bernsen and A. van der Kogel. Antiangiogenic therapy in brain tumor models. *J. Neuro-oncology*, 45:247–255, 1999.
- [69] R. Betteridge, M. Owen, H. Byrne, T. Alarcón, and P. Maini. The impact of cell crowding and active cell movement on vascular tumour growth. *Networks Heterogen. Media*, 1:515–535, 2006.
- [70] Y. Bi, C. H. Stuelten, T. Kilts, S. Wadhwa, R. V. Iozzo, P. G. Robey, X.-D. Chen, and M. F. Young. Extracellular matrix proteoglycans control the fate of bone marrow stromal cells. *J. Biol. Chem.*, 280:30481–9, 2005.
- [71] M. Bienz and H. Clevers. Linking colorectal cancer to Wnt signaling. *Cell*, 103:311–20, 2000.
- [72] M. V. Blagosklonny and A. B. Pardee. The restriction point of the cell cycle. *Cell Cycle*, 1(2):103–110, 2002.
- [73] C. Blanpain and E. Fuchs. Epidermal Stem Cells of the Skin. *Annu. Rev. Cell Dev. Biol.*, 22:339–73, 2006.
- [74] H. Bloemendal, T. Logtenberg, and E. Voest. New strategies in anti-vascular cancer therapy. *Euro J. Clin. Invest.*, 29:802–809, 1999.

- 
- [75] C. Boccaccio, M. Andò, L. Tamagnone, A. Bardelli, P. Michieli, C. Battistini, and P. Comoglio. Induction of epithelial tubules by growth factor hgf depends on the stat pathway. *Nature*, 391:285–288, 1998.
- [76] K. Bold, Y. Zou, I. Kevrekidis, and M. Henson. An equation-free approach to analyzing heterogeneous cell population dynamics. *J. Math. Biol.*, 55:331–352, 2007.
- [77] A. Brandt. Multi-level adaptive solutions to boundary-value problems. *Math. Comput.*, 31:333–390, 1977.
- [78] B. Brandt, D. Kemming, J. Packeisen, R. Simon, M. Helms, U. Feldmann, A. Matuschek, C. Kersting, B. Hinrichs, J.-M. Midart, D. Bellet, K. Bartkowiak, N. Dankbar, T. Dittmar, G. Sauter, W. Boecker, and H. Buerger. Expression of early placenta insulin-like growth factor in breast cancer cells provides an autocrine loop that predominantly enhances invasiveness and motility. *Endocrine-Related Canc.*, 12(4):823–7, 2005.
- [79] A. Bredel-Geissler, U. Karbach, S. Walenta, L. Vollrath, and W. Mueller-Klieser. Proliferation-associated oxygen consumption and morphology of tumor cells in monolayer and spheroid culture. *J. Cell Phys.*, 153:44–52, 1992.
- [80] D. Bresch, T. Colin, E. Grenier, and B. Ribba. A viscoelastic model for avascular tumor growth. inria-00267292, version 2, 2008.
- [81] D. Bresch, T. Colin, E. Grenier, B. Ribba, and O. Saut. Computational modeling of solid tumor growth: the avascular stage. unpublished, 2007.
- [82] C. Beward, H. Byrne, and C. Lewis. The role of cell-cell interactions in a two-phase model for avascular tumour growth. *J. Math. Biol.*, 45:125–152, 2002.
- [83] C. Beward, H. Byrne, and C. Lewis. A multiphase model describing vascular tumor growth. *Bull. Math. Biol.*, 65:609–640, 2003.
- [84] D. Brizel, S. Scully, J. Harrelson, L. Layfield, J. Bean, L. Prosnitz, and M. Dewhirst. Tumor oxygenation predicts for the likelihood of distant metastases in human soft tissue sarcoma. *Cancer Res.*, 56:941–943, 1996.
- [85] D. Brizel, G. Sibley, L. Prosnitz, R. Scher, and M. Dewhirst. Tumor hypoxia adversely affects the prognosis of carcinoma of the head and neck. *Int. J. Radiat. Oncol. Biol. Phys.*, 38:285–289, 1997.
- [86] J. Brown and L. Lowe. Multigrid elliptic equation solver with adaptive mesh refinement. *J. Comput. Phys.*, 209:582–598, 2005.
- [87] R. K. Bruick and S. L. McKnight. A conserved family of Prolyl-4-Hydroxylases that modify HIF. *Science*, 294(5545):1337–40, 2001.
- [88] H. Bueno, G. Ercole, and A. Zumpano. Asymptotic behaviour of quasi-stationary solutions of a nonlinear problem modelling the growth of tumours. *Nonlinearity*, 18:1629–1642, 2005.
- [89] E. Bullitt, D. Zeng, G. Gerig, S. Aylward, S. Joshi, J. Smith, W. Lin, and M. Ewend. Vessel tortuosity and brain tumor malignance: a blinded study. *Acad. Radiol.*, 12:1232–1240, 2005.
- [90] A. Burton. Rate of growth of solid tumours as a problem of diffusion. *Growth*, 30:157–176, 1966.
- [91] F. Bussolino, M. Arese, E. Audero, E. Giraudo, S. Marchiò, S. Mitola, L. Primo, and G. Serini. *Cancer Modelling and Simulation*, chapter 1: Biological Aspects of Tumour Angiogenesis, pages 1–22. Chapman and Hall/CRC, London, 2003.
- [92] L. M. Butler, S. Khan, G. E. Rainger, and G. B. Nash. Effects of endothelial basement membrane on neutrophil adhesion and migration. *Cell. Immun.*, 251:56–61, 2008.

- [93] S. Byers, C. Sommers, B. Hoxter, A. Mercurio, and A. Tozeren. Role of e-cadherin in the response of tumor cell aggregates to lymphatic, venous and arterial flow: measurement of cell-cell adhesion strength. *J. Cell Sci.*, 108:2053–2064, 1995.
- [94] H. Byrne. The effect of time delays on the dynamics of avascular tumor growth. *Math. Biosci.*, 144:83–117, 1997.
- [95] H. Byrne. The importance of intercellular adhesion in the development of carcinomas. *IMA J. Math. Med. Biol.*, 14:305–323, 1997.
- [96] H. Byrne. A comparison of the roles of localized and nonlocalised growth factors in solid tumour growth. *Math. Models Meth. Appl. Sci.*, 9:541–568, 1999.
- [97] H. Byrne. A weakly nonlinear analysis of a model of avascular solid tumour growth. *J. Math. Biol.*, 39:59–89, 1999.
- [98] H. Byrne, T. Alarcón, M. Owen, S. Webb, and P. Maini. Modeling aspects of cancer dynamics: A review. *Phi. Trans. R. Soc. A*, 364:1563–1578, 2006.
- [99] H. Byrne and M. Chaplain. Growth of nonnecrotic tumors in the presence and absence of inhibitors. *Mathl. Biosci.*, 130:151–181, 1995.
- [100] H. Byrne and M. Chaplain. Mathematical models for tumour angiogenesis: Numerical simulations and nonlinear wave solutions. *Bull. Math. Biol.*, 57:461–486, 1995.
- [101] H. Byrne and M. Chaplain. Growth of necrotic tumors in the presence and absence of inhibitors. *Mathl. Biosci.*, 135:187–216, 1996.
- [102] H. Byrne and M. Chaplain. Modelling the role of cell-cell adhesion in the growth and development of carcinomas. *Mathl. Comput. Modelling*, 24:1–17, 1996.
- [103] H. Byrne and M. Chaplain. Free boundary value problems associated with the growth and development of multicellular spheroids. *Eur. J. Appl. Math.*, 8:639–658, 1997.
- [104] H. Byrne and D. Drasdo. Individual-based and continuum models of growing cell populations: A comparison. *J. Math. Biol.*, 58(4-5):657–87, 2009.
- [105] H. Byrne, J. King, D. McElwain, and L. Preziosi. A two-phase model of solid tumour growth. *Appl. Math. Letters*, 16:567–573, 2003.
- [106] H. Byrne and P. Matthews. Asymmetric growth of models of avascular solid tumors: exploiting symmetries. *IMA J. Math. Appl. Med. Biol.*, 19:1–29, 2002.
- [107] H. Byrne and L. Preziosi. Modelling solid tumour growth using the theory of mixtures. *Math. Med. Biol.*, 20:341–366, 2003.
- [108] N. Cabioglu, K. K. Hunt, A. A. Sahin, H. M. Kuerer, G. V. Babiera, S. E. Singletary, G. J. Whitman, M. I. Ross, F. C. Ames, B. W. Feig, T. A. Buchholz, and F. Meric-Bernstam. Role for intraoperative margin assessment in patients undergoing breast-conserving surgery. *Ann. Surg. Oncol.*, 14(4):1458–71, 2007.
- [109] J. Cahn and J. Hilliard. Free energy of a nonuniform system. i. interfacial free energy. *J. Chem. Phys.*, 28:258–267, 1958.
- [110] R. Cairns, T. Kalliomaki, and R. Hill. Acute (cyclic) hypoxia enhances spontaneous metastasis of kht murine tumors. *Cancer Res.*, 61:8903–8908, 2001.
- [111] D. Q. Calcagno, M. F. Leal, A. D. Seabra, A. S. Khayat, E. S. Chen, S. Demachki, P. P. Assumpcao, M. H. G. Faria, S. H. B. Rabenhorst, M. V. P. Ferreira, M. D. C. Smith, and R. R. Burbano. Interrelationship between chromosome 8 aneuploidy, C-MYC amplification and increased expression in individuals from northern Brazil with gastric adenocarcinoma. *World J. Gastroenterol.*, 12(38):6027–211, 2006.
- [112] V. Capasso and D. Morale. Stochastic modelling of tumour-induced angiogenesis. *J. Math. Biol.*, 58:219–233, 2009.

- 
- [113] J. Carlsson and H. Acker. Relations ph, oxygen partial pressure and growth in cultured cell spheroids. *Int. J. Cancer*, 42:715–720, 1988.
- [114] P. Carmeliot and R. Jain. Angiogenesis in cancer and other diseases. *Nature*, 407:249–257, 2000.
- [115] L. Carreras, P. Macklin, J. Kim, S. Sanga, V. Cristini, and M. E. Edgerton. Oxygen uptake in quiescent versus cycling cells in a model of DCIS. *U.S. Canadian Acad. Path.*, 2010 Annual Meeting, 2010. (submitted).
- [116] J. Casciari, S. Sotirchos, and R. Sutherland. Glucose diffusivity in multicellular tumor spheroids. *Cancer Res.*, 48:3905–3909, 1988.
- [117] J. Casciari, S. Sotirchos, and R. Sutherland. Variations in tumor cell growth rates and metabolism with oxygen concentration, glucose concentration, and extracellular ph. *J. Cell. Physiol.*, 151:386–394, 1992.
- [118] M. Castro, C. Molina-Paris, and T. Deisboeck. Tumor growth instability and the onset of invasion. *Phys. Rev. E*, 72:041907, 2005.
- [119] P. Castro, P. Soares, L. Gusmo, R. Seruca, and M. Sobrinho-Simes. H-RAS 81 polymorphism is significantly associated with aneuploidy in follicular tumors of the thyroid. *Oncogene*, 25:4620–7, 2006.
- [120] M. Chaplain. Reaction-diffusion prepatterning and its potential role in tumour invasion. *J. Biol. Sys.*, 3:929–936, 1995.
- [121] M. Chaplain. Avascular growth, angiogenesis and vascular growth in solid tumours: The mathematical modelling of the stages of tumour development. *Mathl. Comput. Modelling*, 23:47–87, 1996.
- [122] M. Chaplain. Pattern formation in cancer. In M. Chaplain, G. Singh, and J. MacLachlan, editors, *On Growth and Form: Spatio-Temporal Pattern Formation in Biology*. Wiley, New York, 2000.
- [123] M. Chaplain, M. Ganesh, and I. Graham. Spatio-temporal pattern formation on spherical surfaces: numerical simulation and application to solid tumour growth. *J. Math. Biol.*, 42:387–423, 2001.
- [124] M. Chaplain, L. Graziano, and L. Preziosi. Mathematical modelling of the loss of tissue compression responsiveness and its role in solid tumor development. *Math. Med. Biol.*, 23:197–229, 2006.
- [125] M. Chaplain and G. Lolas. Mathematical modeling of cancer cell invasion of tissue: The role of the urokinase plasminogen activation system. *Math. Models Meth. Appl. Sci.*, 15:1685–1734, 2005.
- [126] M. Chaplain, S. McDougall, and A. Anderson. Mathematical modeling of tumor-induced angiogenesis. *Ann. Rev. Biomed. Eng.*, 8:233–257, 2006.
- [127] M. Chaplain and B. Sleeman. Modelling the growth of solid tumours and incorporating a method for their classification using nonlinear elasticity theory. *J. Math. Biol.*, 31:431–479, 1993.
- [128] M. Chaplain and A. Stuart. A model mechanism for the chemotactic response of endothelial cells to tumor angiogenesis factor. *IMA J. Math. Appl. Med. Biol.*, 10:149–168, 1993.
- [129] A. Chauvier and L. Preziosi. A mathematical framework to model migration of a cell population in the extracellular matrix. In A. Chauvier, L. Preziosi, and C. Verdier, editors, *Cell mechanics: From single cell scale-based models to multiscale modeling*. Chapman and Hall/CRC Press, Boca Raton, 2009.



- [130] A. Chauviere, T. Hillen, and L. Preziosi. Modeling cell movement in anisotropic and heterogeneous tissues. *Networks Hetero. Media*, 2:333–357, 2007.
- [131] A. Chauviere, L. Preziosi, and T. Hillen. Modeling the motion of a cell population in the extracellular matrix. *Discrete Cont. Dyn. Syst. B*, Supp:250–259, 2007.
- [132] C. Chen, H. Byrne, and J. King. The influence of growth-induced stress from the surrounding medium on the development of multicell spheroids. *J. Math. Biol.*, 43:191–220, 2001.
- [133] L. L. Chen, L. Zhang, J. Yoon, and T. S. Deisboeck. Cancer cell motility: optimizing spatial search strategies. *Biosys.*, 95(3):234–42, 2009.
- [134] W. W. Chen, B. Schoeberl, P. J. Jasper, M. Niepel, D. A. Nielsen, U B andLauffenburger, et al. Input-output behavior of ErbB signaling pathways as revealed by a mass action model trained against dynamic data. *Mol. Syst. Biol.*, 5:239ff, 2009.
- [135] X. Chen, S. Cui, and A. Friedman. A hyperbolic free boundary problem modeling tumor growth: asymptotic behavior. *Trans. Am. Math. Soc.*, 357:4771–4804, 2005.
- [136] L. Cheng, N. K. Al-Kaisi, N. H. Gordon, A. Y. Liu, F. Gebrail, and R. R. Shenk. Relationship between the size and margin status of ductal carcinoma in situ of the breast and residual disease. *J. Natl. Cancer Inst.*, 89(18):1356–60, 1997.
- [137] J. H. Choi, Y. Y. Jeong, S. S. Shin, H. S. Lim, and H. K. Kang. Primary calcified T-cell lymphoma of the urinary bladder: a case report. *Korean J. Radiol.*, 4(4):252–4, 2003.
- [138] Y.-L. Chuang, M. E. Edgerton, P. Macklin, S. Wise, J. S. Lowengrub, and V. Cristini. Clinical predictions of bulk DCIS properties based on a duct-scale mixture model. (in preparation), 2009.
- [139] S. Ciatto, S. Bianchi, and V. Vezzosi. Mammographic appearance of calcifications as a predictor of intraductal carcinoma histologic subtype. *Eur. Radiology*, 4(1):23–6, 1994.
- [140] R. G. Clyde, J. L. Brown, T. R. Hupp, N. Zhelev, and J. W. Crawford. The role of modelling in identifying drug targets for diseases of the cell cycle. *J. R. Soc. Interface*, 3(10):617–627, 2006.
- [141] D. Coffey. Self-organization, complexity and chaos: the new biology for medicine. *Nature Med.*, 4:882, 1998.
- [142] P. Colella, D. T. Graves, T. J. Ligocki, D. F. Martin, D. Modiano, D. B. Serafini, and B. V. Straalen. CHOMBO software package for AMR applications: design document. Technical report, Lawrence Berkeley National Laboratory, Applied Numerical Algorithms Group; NERSC Division, Berkeley, CA, USA, 2003.
- [143] B. Coleman and W. Noll. Thermodynamics of elastic materials with conduction and viscosity. *Arch. Rat. Mech.*, 13:167–178, 1963.
- [144] M. Conacci-Sorrell, J. Zhurinsky, and A. Ben-Zeév. The cadherin-catenin adhesion system in signaling and cancer. *J. Clin. Invest.*, 109(8):987–91, 2002.
- [145] J. Condeelis, R. Singer, and J. Segall. The great escape: When cancer cells hijack the genes for chemotaxis and motility. *Annu. Rev. Cell Dev. Biol.*, 21:695–718, 2005.
- [146] A. Coniglio, A. deCandia, S. DiTalia, and A. Gamba. Percolation and burgers’ dynamics in a model of capillary formation. *Phys. Rev. E*, 69:051910, 2004.
- [147] V. Cristini, H. Frieboes, R. Gatenby, S. Caserta, M. Ferrari, and J. Sinek. Morphologic instability and cancer invasion. *Clin. Cancer Res.*, 11:6772–6779, 2005.
- [148] V. Cristini, H. Frieboes, X. Li, J. Lowengrub, P. Macklin, S. Sanga, S. Wise, and X. Zheng. Nonlinear modeling and simulation of tumor growth. In N. Bellomo, M. Chaplain, and E. de Angelis, editors, *Modelling and simulation in science, engineering and technology*. Birkhäuser, Boston, 2008.

- 
- [149] V. Cristini, X. Li, J. Lowengrub, and S. Wise. Nonlinear simulations of solid tumor growth using a mixture model: invasion and branching. *J. Math. Biol.*, 58(4-5):723–763, 2009.
- [150] V. Cristini and J. Lowengrub. Three-dimensional crystal growth. i. linear analysis and self-similar evolution. *J. Crystal Growth*, 240:267–276, 2002.
- [151] V. Cristini, J. Lowengrub, and Q. Nie. Nonlinear simulation of tumor growth. *J. Math. Biol.*, 46:191–224, 2003.
- [152] S. Cui. Analysis of a mathematical model for the growth of tumors under the action of external inhibitors. *J. Math. Biol.*, 44:395–426, 2002.
- [153] S. Cui. Analysis of a free boundary problem modeling tumor growth. *Acta Math. Sinica*, 21:1071–1082, 2005.
- [154] S. Cui. Well-posedness of a multidimensional free boundary problem modelling the growth of nonnecrotic tumors. *J. Func. Analysis*, 245:1–18, 2007.
- [155] S. Cui. Lie group action and stability analysis of stationary solutions for a free boundary problem modelling tumor growth. *J. Diff. Eq.*, 246(5):1845–1882, 2009.
- [156] S. Cui and J. Escher. Asymptotic behaviour of solutions of a multidimensional moving boundary problem modeling tumor growth. *Comm. Partial Diff. Equations*, 33:636–655, 2008.
- [157] S. Cui and J. Escher. Well-posedness and stability of a multi-dimensional tumor growth model. *Arch. Rat. Mech. Analysis*, 191:173–193, 2009.
- [158] S. Cui and A. Friedman. Analysis of a mathematical model of the effect of inhibitors on the growth of tumors. *Math. Biosci.*, 164:103–137, 2000.
- [159] S. Cui and A. Friedman. A free boundary problem for a singular system of differential equations: An application to a model of tumor growth. *Trans. Amer. Math. Soc.*, 255:3537–3590, 2003.
- [160] S. Cui and A. Friedman. Formation of necrotic cores in the growth of tumors: analytic results. *Acta Mathematica Scientia*, 26:781–796, 2006.
- [161] S. Cui and X. Wei. Global existence for a parabolic-hyperbolic free boundary problem modelling tumor growth. *Acta Math. Appl. Sinica*, 21:597–614, 2005.
- [162] S. Cui and S. Xu. Analysis of mathematical models for the growth of tumors with time delays in cell proliferation. *J. Math. Analysis Appl.*, 336:523–541, 2007.
- [163] J. Dallon and H. Othmer. How cellular movement determines the collective force generated by the dictyostelium discoideum slug. *J. Theor. Biol.*, 231:299–306, 2004.
- [164] C. G. Danes, S. L. Wyszomierski, J. Lu, C. L. Neal, W. Yang, and D. Yu. 14-3-3 $\zeta$  down-regulates p53 in mammary epithelial cells and confers luminal filling. *Canc. Res.*, 68:1760–7, 2008.
- [165] K. Date, K. Matsumoto, K. Kuba, H. Shimura, M. Tanaka, and T. Nakamura. Inhibition of tumor growth and invasion by a four-kringle antagonist (hgf/nk4) for hepatocyte growth factor. *Oncogene*, 17:3045–3054, 1998.
- [166] C. R. De Potter, I. Eeckhout, A.-M. Schelfhout, M.-L. Geerts, and H. J. Roelsh. Keratinocyte induced chemotaxis in the pathogenesis of Paget’s disease of the breast. *Histopath.*, 24(4):349–56, 1994.
- [167] J. Debnath and J. Brugge. Modelling glandular epithelial cancers in three-dimensional cultures. *Nature Rev. Cancer*, 5:675–688, 2005.
- [168] J. Debnath, K. Mills, N. Collins, M. Reginato, S. Muthuswamy, and J. Brugge. The role of apoptosis in creating and maintaining luminal space within normal and oncogene-expressing mammary acini. *Cell*, 111:29–40, 2002.

- [169] T. Deisboeck, M. Berens, A. Kansal, S. Torquato, A. Stemmer-Rachamimov, and E. Chiocca. Pattern of self-organization in tumour systems: Complex growth dynamics in a novel brain tumour spherical model. *Cell Proliferation*, 34:115–134, 2001.
- [170] T. Deisboeck, L. Zhang, J. Yoon, and J. Costa. In silico cancer modeling: is it ready for prime time? *Nature Clin. Practice Oncol.*, 6(1):34–42, 2009.
- [171] K. DeJaeger, M. Kavanagh, and R. Hill. Relationship of hypoxia to metastatic ability in rodent tumors. *Br. J. Cancer*, 84:1280–1285, 2001.
- [172] H. D. Dell. Milestone 1 (1889) Seed and soil hypothesis: Observations from a ploughman. *Nat. Rev. Cancer*, 6:S7, 1989. <http://www.nature.com/milestones/milecancer/index.html>.
- [173] R. Demicheli, G. Pratesi, and R. Foroni. The exponential-gompertzian growth model : data from six tumor cell lines in vitro and in vivo. estimate of the transition point from exponential to gompertzian growth and potential clinical applications. *Tumori*, 77:189–195, 1991.
- [174] B. Desai, T. Ma, and M. A. Chellaiah. Invadopodia and matrix degradation, a new property of prostate cancer cells during migration and invasion. *J. Biol. Chem.*, 283(20):13856–66, 2008.
- [175] A. Deutsch and S. Dormann. *Cellular automaton modeling of biological pattern formation*. Birkhäuser, New York, 2005.
- [176] A. E. DeWitt, J. Y. Dong, H. S. Wiley, and D. A. Lauffenburger. Quantitative analysis of the EGF receptor autocrine system reveals cryptic regulation of cell response by ligand capture. *J. Cell. Sci.*, 114(Pt 12):2301–13, 2001.
- [177] J. Diaz and J. Tello. On the mathematical controllability in a simple growth tumors model by the internal localized action of inhibitors. *Nonlinear Analysis*, 4:109–125, 2003.
- [178] A. DiCarlo and S. Quiligotti. Growth and balance. *Mech. Res. Comm.*, 29:449–456, 2002.
- [179] M. F. Dillon, A. A. Maguire, E. W. McDermott, C. Myers, A. D. Hill, A. O’Doherty, and C. M. Quinn. Needle core biopsy characteristics identify patients at risk of compromised margins in breast conservation surgery. *Mod. Pathol.*, 21(1):39–45, 2008.
- [180] M. F. Dillon, E. W. McDermott, A. O’Doherty, C. M. Quinn, A. D. Hill, and N. O’Higgins. Factors affecting successful breast conservation for ductal carcinoma in situ. *Ann. Surg. Oncol.*, 14(5):1618–28, 2007.
- [181] R. Dillon, M. Owen, and K. Painter. A single-cell based model of multicellular growth using the immersed interface method. In B. Khoo, Z. Li, and P. Lin, editors, *Contemporary mathematics: Moving interface problems and applications in fluid dynamics*, volume 466, chapter 1, pages 1–15. AMS, Providence, 2008.
- [182] S. Dormann and A. Deutsch. Modeling of self-organized avascular tumor growth with a hybrid cellular automaton. *In Silico Biology*, 2:393–406, 2002.
- [183] D. Drasdo. On selected individual-based approaches to the dynamics of multicellular systems. In W. Alt, M. Chaplain, and M. Griebel, editors, *Multiscale modeling*. Birkhaeuser, Basel, 2003.
- [184] D. Drasdo. Coarse graining in simulated cell populations. *Adv. Complex Sys.*, 8:319–363, 2005.
- [185] D. Drasdo and S. Höhme. Individual-based approaches to birth and death in avascular tumors. *Math. Comput. Modelling*, 37:1163–1175, 2003.

- 
- [186] D. Drasdo and S. Höhme. A single-scale-based model of tumor growth in vitro: monolayers and spheroids. *Phys. Biol.*, 2:133–147, 2005.
- [187] D. Drasdo and S. Höhme. On the role of physics in the growth and pattern of multicellular systems: What we learn from individual-cell based models? *J. Stat. Phys.*, 128:287–345, 2007.
- [188] D. Drasdo, R. Kree, and J. McCaskill. Monte-carlo approach to tissue cell populations. *Phys. Rev. E*, 52:6635–6657, 1995.
- [189] F. Drees, S. Pokutta, S. Yamada, W. J. Nelson, and W. T. Weis. Alpha-catenin is a molecular switch that binds E-cadherin-beta-catenin and regulates actin-filament assembly. *Cell*, 123:903–15, 2005.
- [190] W. R. Duan, D. S. Garner, S. D. Williams, C. L. Funckes-Shippy, I. S. Spath, and E. A. G. Blomme. Comparison of immunohistochemistry for activated caspase-3 and cleaved cytokeratin 18 with the TUNEL method for quantification of apoptosis in histological sections of PC-3 subcutaneous xenografts. *J. Pathol.*, 199(2):221–8, 2003.
- [191] M. Ducasse and M. A. Brown. Epigenetic aberrations and cancer. *Mol. Cancer*, 5:60, 2006.
- [192] H. S. Earp, T. L. Dawson, X. Li, and H. Yu. Heterodimerization and functional interaction between EGF receptor family members: a new signaling paradigm with implications for breast cancer research. *Breast Canc. Res. Treat.*, 35(1):115–132, 1995.
- [193] D. A. Eberhard, B. E. Johnson, L. C. Amler, et al. Mutations in the epidermal growth factor receptor and in KRAS are predictive and prognostic indicators in patients with non-small-cell lung cancer treated with chemotherapy alone and in combination with erlotinib. *J. Clin. Oncol.*, 23:5900–09, 2005.
- [194] M. Edgerton, Y.-L. Chuang, J. Kim, G. Tomaiuolo, P. Macklin, S. Sanga, W. Yang, A. Broom, K.-A. Do, and V. Cristini. Using mathematical models to understand the time dependence of the growth of ductal carcinoma in situ. *31st Annual San Antonio Breast Cancer Symposium.*, Supplement to Volume 68(24):Abstract 1165, 2008.
- [195] M. Edgerton, Y.-L. Chuang, J. Kim, G. Tomaiuolo, P. Macklin, S. Sanga, W. Yang, A. Broom, K.-A. Do, and V. Cristini. Using mathematical models to understand time-dependence of growth DCIS: Implications for clinical detection and IDC. (in preparation), 2009.
- [196] M. Edgerton, P. Macklin, Y.-L. Chuang, G. Tomaiuolo, W. Yang, J. Kim, S. Sanga, A. Broom, K.-A. Do, and V. Cristini. A multiscale model of ductal carcinoma in situ. *Cancer Res.*, 2010. (submitted).
- [197] A. W. El-Kareh and T. W. Secomb. A mathematical model for cisplatin cellular pharmacodynamics. *Neoplasia*, 5(2):161–9, 2003.
- [198] A. W. El-Kareh and T. W. Secomb. Two-mechanism peak concentration model for cellular pharmacodynamics of doxorubicin. *Neoplasia*, 7(7):705–13, 2005.
- [199] C. Elliot and S. Luckhaus. A generalized diffusion equation for phase separation of a multi-component mixture with interfacial free energy. Technical report, Univ. Sussex and Univ. Bonn, 1991. Inst. Math. Appl., report 887.
- [200] Y. I. Elshimali and W. W. Grody. The Clinical Significance of Circulating Tumor Cells in the Peripheral Blood. *Diagn. Mol. Pathol.*, 15(4):187–194, 2006.
- [201] P. Elvin and A. Garner. Tumour invasion and metastasis: challenges facing drug discovery. *Curr. Opin. Pharmacol.*, 5:374–381, 2005.
- [202] S. Enam, M. Rosenblum, and K. Edvardsen. Role of extracellular matrix in tumor invasion: migration of glioma cells along fibronectinpositive mesenchymal cell processes.

- Neurosurgery*, 42:599–608, 1998.
- [203] R. Erban, I. Kevrekidis, and H. Othmer. An equation-free computational approach for extracting population-level behavior from individual-based models of biological dispersal. *Physica D*, 215:1–24, 2006.
- [204] R. Erban and H. Othmer. From individual to collective behavior in bacterial chemotaxis. *SIAM J. Appl. Math.*, 65:361–391, 2004.
- [205] R. Erban and H. Othmer. From signal transduction to spatial pattern formation in *E. coli*: A paradigm for multi-scale modeling in biology. *Multiscale Model. Simul.*, 3:362–394, 2005.
- [206] B. Erbas, E. Provenzano, J. Armes, and D. Gertig. The natural history of ductal carcinoma *in situ* of the breast: a review. *Breast Canc. Res. Treat.*, 97(2):135–44, 2006.
- [207] J. Erler, K. Bennewith, M. Nicolau, N. Dornhoefer, C. Kong, Q.-T. Le, J.-T. Chi, S. Jeffrey, and A. Giaccia. Lysyl oxidase is essential for hypoxia-induced metastasis. *Nature*, 440:1222–1226, 2006.
- [208] M. Esteban and P. Maxwell. If, a missing link between metabolism and cancer. *Nature Med.*, 11:1047–1048, 2005.
- [209] J. Fang, R. Gillies, and R. Gatenby. Adaptation to hypoxia and acidosis in carcinogenesis and tumor progression. *Semin. Cancer Biol.*, 18:330–337, 2008.
- [210] A. Fasano, A. Bertuzzi, and A. Gandolfi. *Complex systems in biomedicine*, chapter Mathematical modelling of tumour growth and treatment, pages 71–108. Springer, Milan, 2006.
- [211] L. Ferrante, S. Bompadre, L. Possati, and L. Leone. Parameter estimation in a gompertzian stochastic model for tumor growth. *Biometrics*, 56:1076–1081, 2000.
- [212] M. Ferrari. Cancer nanotechnology: opportunities and challenges. *Nature Rev. Cancer*, 5:161–171, 2005.
- [213] R. Filion and A. Popel. A reaction-diffusion model of basic fibroblast growth factor interactions with cell surface receptors. *Annals Biomed. Eng.*, 32:645–663, 2004.
- [214] I. Fischer, J.-p. Gagner, M. Law, E. W. Newcomb, and D. Zagzag. Angiogenesis in Gliomas: Biology and Molecular Pathophysiology. *Brain Pathol.*, 15(4):297–310, 2005.
- [215] A. B. Fisher, S. Chien, A. I. Barakat, and R. M. Nerem. Endothelial cellular response to altered shear stress. *Am. J. Physiol. Heart Circ. Physiol.*, 281(3):L529–L533, 2001.
- [216] B. Flaherty, J. P. McGarry, and P. E. McHugh. Mathematical models of cell motility. *Cell. Biochem. Biophys.*, 49(1):14–28, 2007.
- [217] G. B. Fogarty, N. M. Conus, J. Chu, and G. McArthur. Characterization of the expression and activation of the epidermal growth factor receptor in squamous cell carcinoma of the skin. *Brit. J. Derm.*, 156(1):92–98, 2007.
- [218] J. Folkman. Angiogenesis in cancer, vascular, rheumatoid and other disease. *Nat. Med.*, 1:27–30, 1995.
- [219] J. Forsythe, B. Jiang, N. Iyer, S. L. F. Agani, R. Koos, and G. Semenza. Activation of vascular endothelial growth factor gene transcription by hypoxia-inducible factor 1. *Mol. Cell. Biol.*, 16:4604–4613, 1996.
- [220] L. M. Franks and M. A. Knowles. What is Cancer? In Knowles and Selby [386], chapter 1, pages 1–24.
- [221] S. Franks, H. Byrne, J. King, J. Underwood, and C. Lewis. Modeling the early growth of ductal carcinoma *in situ* of the breast. *J. Math. Biol.*, 47:424–452, 2003.
- [222] S. Franks, H. Byrne, H. Mudhar, J. Underwood, and C. Lewis. Mathematical modeling of comedo ductal carcinoma *in situ* of the breast. *Math. Med. Biol.*, 20:277–308, 2003.

- 
- [223] S. Franks and J. King. Interactions between a uniformly proliferating tumor and its surrounding. uniform material properties. *Math. Med. Biol.*, 20:47–89, 2003.
- [224] J. Freyer and R. Sutherland. Determination of diffusion constants for metabolites in multicell tumor spheroids. *Adv. Exp. Med. Biol.*, 159:463–475, 1983.
- [225] J. Freyer and R. Sutherland. A reduction in the in situ rates of oxygen and glucose consumption of cells in emt6/ro spheroids during growth. *J. Cell. Physiol.*, 124:516–524, 1985.
- [226] J. Freyer and R. Sutherland. Regulation of growth saturation and development of necrosis in emt6/ro multicellular spheroids by the glucose and oxygen supply. *Cancer Res.*, 46:3504–3512, 1986.
- [227] J. P. Freyer. Decreased mitochondrial function in quiescent cells isolated from multicellular tumor spheroids. *J. Cell. Physiol.*, 176(1):138–49, 1998.
- [228] H. Frieboes, M. Edgerton, J. Fruehauf, F. Rose, L. Worrall, R. Gatenby, M. Ferrari, and V. Cristini. Prediction of drug response in breast cancer using integrative experimental/computational modeling. *Cancer Research*, 69:4484–4492, 2009.
- [229] H. Frieboes, J. Fang, Y.-L. Chuang, S. Wise, J. Lowengrub, and V. Cristini. Nonlinear simulations of three-dimensional multispecies tumor growth -ii: Tumor invasion and angiogenesis. *J. Theor. Biol.*, in review.
- [230] H. Frieboes, J. Lowengrub, S. Wise, X. Zheng, P. Macklin, E. Bearer, and V. Cristini. Computer simulation of glioma growth and morphology. *NeuroImage*, 37:S59–S70, 2007.
- [231] H. Frieboes, X. Zheng, C.-H. Sun, B. Tromberg, R. Gatenby, and V. Cristini. An integrated computational/experimental model of tumor invasion. *Cancer Res.*, 66:1597–1604, 2006.
- [232] P. Friedl. Prespecification and plasticity: shifting mechanisms of cell migration. *Curr. Opin. Cell Biol.*, 16:14–23, 2004.
- [233] P. Friedl, E. Brocker, and K. Zanker. Integrins, cell matrix interactions and cell migration strategies: fundamental differences in leukocytes and tumor cells. *Cell Adhes. Commun.*, 6:225–236, 1998.
- [234] P. Friedl, Y. Hegerfeldt, and M. Tilisch. Collective cell migration in morphogenesis and cancer. *Int. J. Dev. Biol.*, 48:441–449, 2004.
- [235] P. Friedl, P. Noble, P. Walton, D. Laird, P. Chauvin, R. Tabah, M. Black, and K. Zaenker. Migration of coordinated cell clusters in mesenchymal and epithelial cancer explants in vitro. *Cancer Res.*, 55:4557–4560, 1995.
- [236] P. Friedl and A. Wolf. Tumor cell invasion and migration: diversity and escape mechanisms. *Nat. Rev. Cancer*, 3:362–374, 2003.
- [237] A. Friedman. A hierarchy of cancer models and their mathematical challenges. *Discrete Cont. Dyn. Systems Ser. B*, 4:147–159, 2004.
- [238] A. Friedman, N. Bellomo, and P. Maini. Mathematical analysis and challenges arising from models of tumor growth. *Math. Models Meth. Appl. Sci.*, 17:1751–1772, 2007.
- [239] A. Friedman and B. Hu. Asymptotic stability for a free boundary problem arising in a tumor model. *J. Diff. Equations*, 227:598–639, 2005.
- [240] A. Friedman and B. Hu. Bifurcation from stability to instability for a free boundary problem arising in a tumor model. *Arch. Rat. Mech. Analysis*, 180:293–330, 2006.
- [241] A. Friedman and B. Hu. Bifurcation from stability to instability for a free boundary problem modeling tumor growth by stokes equation. *J. Math. Analysis Appl.*, 327:643–664, 2007.

- [242] A. Friedman and F. Reitich. Analysis of a mathematical model for the growth of tumors. *J. Math. Biol.*, 38:262–284, 1999.
- [243] A. Friedman and F. Reitich. On the existence of spatially patterned dormant malignancies in a model for the growth of non-necrotic vascular tumors. *Math. Models Meth. Appl. Sci.*, 11:601–625, 2001.
- [244] A. Friedman and F. Reitich. Symmetry-breaking bifurcation of analytic solutions to free boundary problems: an application to a model of tumor growth. *Trans. Am. Math. Soc.*, 353:1587–1634, 2001.
- [245] S. H. Friend, R. Bernards, S. Rogelj, R. A. Weinberg, J. M. Rapaport, D. M. Albert, and T. P. Dryja. A human DNA segment with properties of the gene that predisposes to retinoblastoma and osteosarcoma. *Nature*, 323:643–6, 1986.
- [246] S. M. Frisch and H. Francis. Disruption of epithelial cell-matrix interactions induces apoptosis. *J. Cell Biol.*, 124:619–26, 1994.
- [247] R. Fukuda, H. Zhang, J.-W. Kim, L. Shimoda, C. V. Dang, and G. L. Semenza. HIF-1 regulates cytochrome oxidase subunits to optimize efficiency of respiration in hypoxic cells. *Cell*, 129(1):111–22, 2007.
- [248] D. Fukumura and R. Jain. Tumor microenvironment abnormalities: causes, consequences, and strategies to normalize. *J. Cell. Biochem.*, 101:937–949, 2007.
- [249] Y. Fung. *Biomechanics: motion, flow, stress and growth*. Springer, New York, 1990.
- [250] Y. Fung. *Biomechanics: material properties of living tissues*. Springer, New York, 1993.
- [251] A.-P. Gadeau, H. Chaulet, D. Daret, M. Kockx, J.-M. Daniel-Lamazière, and C. Desgranges. Time course of osteopontin, osteocalcin, and osteonectin accumulation and calcification after acute vessel wall injury. *J. Histochem. Cytochem.*, 49:79–86, 2001.
- [252] D. Galaris, A. Barbouti, and P. Korantzopoulos. Oxidative stress in hepatic ischemia-reperfusion injury: The role of antioxidants and iron chelating compounds. *Curr. Pharma. Design*, 12:2875–2890, 2006.
- [253] J. Galle, G. Aust, G. Schaller, T. Beyer, and D. Drasdo. Individual cell-based models of the spatial temporal organization of multicellular systems- achievements and limitations. *Cytometry*, 69A:704–710, 2006.
- [254] J. Galle, M. Hoffmann, and G. Aust. From single cells to tissue architecture-a bottom-up approach to modeling the spatio-temporal organization of complex multicellular systems. *J. Math. Biol.*, 58:261–283, 2009.
- [255] J. Galle, M. Loeffler, and D. Drasdo. Modeling the effect of deregulated proliferation and apoptosis on the growth dynamics of epithelial cell populations in vitro. *Biophys. J.*, 88:62–75, 2005.
- [256] J. Galle, L. Preziosi, and A. Tosin. Contact inhibition of growth described using a multiphase model and an individual cell based model. *Appl. Math. Lett.*, 2009. in press.
- [257] A. Gamba, D. Ambrosi, A. Coniglio, A. deCandia, S. DiTalia, E. Giraudo, G. Serini, L. Preziosi, and F. Bussolino. Percolation, morphogenesis and burgers dynamics in blood vessels formation. *Phys. Rev. Lett.*, 90:118101, 2003.
- [258] J. Ganghoffer. Some issues related to growth and goal functions for continuum biological systems. *Phil. Mag.*, 85:4353–4391, 2005.
- [259] M. Garbey and G. Zouridakis. Modeling tumor growth: from differential deformable models to growth prediction of tumors detected in pet images. *Eng. Med. Biol. Soc.*, 3:2687–2690, 2003.
- [260] H. Garcke, M. Rumpf, and U. Weikard. The Cahn-Hilliard equation with elasticity: finite element approximation and qualitative studies. *Interfaces Free Bound*, pages 101–

- 118, 2001.
- [261] M. Gardner. The fantastic combinations of john conway's new solitaire game "life". *Scientific American*, 223:120–3, Oct. 1970.
- [262] K. Garikipati, E. Arruda, K. Grosh, H. Narayanan, and S. Calve. A continuum treatment of growth in biological tissue: the coupling of mass transport and mechanics. *J. Mech. Phys. Solids*, 52:1595–1625, 2004.
- [263] L. Garner, Y. Lau, T. Jackson, M. Uhler, D. Jordan, and R. Gilgenbach. Incorporating spatial dependence into a multicellular tumor spheroid growth model. *J. Appl. Phys.*, 98:124701, 2005.
- [264] R. Gatenby and E. Gawlinski. The glycolytic phenotype in carcinogenesis and tumor invasion: insights through mathematical models. *Cancer Res.*, 63:3847–3854, 2003.
- [265] R. Gatenby, E. Gawlinski, A. Gmitro, B. Kaylor, and R. Gillies. Acid-mediated tumor invasion: a multidisciplinary study. *Cancer Res.*, 66:5216–5223, 2006.
- [266] R. Gatenby and E. Gawlinsky. *The tumour microenvironment*, chapter Mathematical models of tumour invasion mediated by transformation-induced alteration of microenvironmental pH, pages 85–99. Wiley, London, 2003.
- [267] R. Gatenby and R. Gillies. A microenvironmental model of carcinogenesis. *Nat. Rev. Cancer*, 8:56–61, 2008.
- [268] R. Gatenby, K. Smallbone, P. Maini, F. Rose, J. Averill, R. Nagle, L. Worrall, and R. Gillies. Cellular adaptations to hypoxia and acidosis during somatic evolution of breast cancer. *Br. J. Cancer*, 97:646–653, 2007.
- [269] R. Gatenby and T. Vincent. An evolutionary model of carcinogenesis. *Cancer Res.*, 63:6212–6220, 2003.
- [270] C. W. Gear and I. G. Kevrekidis. Projective methods for stiff differential equations: problems with gaps in the eigenvalue spectrum. *SIAM J. Sci. Comput.*, 24:1091–1106, 2003.
- [271] C. W. Gear, I. G. Kevrekidis, and C. Theodoropoulos. 'Coarse' integration/bifurcation analysis via microscopic simulators: micro-Galerkin methods. *Comput. Chem. Engineer.*, 26:941–963, 2002.
- [272] A. Gerisch and M. Chaplain. Mathematical modelling of cancer cell invasion of tissue: local and non-local models and the effect of adhesion. *J. Theor. Biol.*, 250:684–704, 2008.
- [273] P. Gerlee and A. Anderson. An evolutionary hybrid cellular automaton model of solid tumor growth. *J. Theor. Biol.*, 246:583–603, 2007.
- [274] P. Gerlee and A. Anderson. Stability analysis of a hybrid cellular automaton model of cell colony growth. *Phys. Rev. E*, 75:051911, 2007.
- [275] P. Gerlee and A. Anderson. A hybrid cellular automaton model of clonal evolution in cancer: The emergence of the glycolytic phenotype. *J. Theor. Biol.*, 250:705–722, 2008.
- [276] P. Gerlee and A. Anderson. A hybrid cellular automaton model of clonal evolution in cancer: The emergence of the glycolytic phenotype. *J. Theor. Biol.*, 250:705–722, 2008.
- [277] J. Gevertz and S. Torquato. Modeling the effects of vasculature evolution on early brain tumor growth. *J. Theor. Biol.*, 243:517–531, 2006.
- [278] F. G. Giancotti and E. Ruoslahti. Integrin signaling. *Science*, 285(5430):1028–32, 1999.
- [279] J. W. Gibbs. Fourier's series. *Nature*, 59:200–200, 1898.
- [280] M. Z. Gilcrease. Integrin signaling in epithelial cells. *Canc. Lett.*, 247(1):1–25, 2007.
- [281] R. Gillies and R. Gatenby. Adaptive landscapes and emergent phenotypes: why do cancers have high glycolysis? *J. Bioenerg. Biomem.*, 39:251–257, 2007.



- [282] R. Gillies and R. Gatenby. Hypoxia and adaptive landscapes in the evolution of carcinogenesis. *Cancer Metastasis Rev.*, 26:311–317, 2007.
- [283] R. Gillies, Z. Liu, and Z. Bhujwala. 31p-mrs measurements of extracellular ph of tumors using 3- aminopropylphosphonate. *Am. J. Physiol.*, 267:195–203, 1994.
- [284] R. Gillies, I. Robey, and R. Gatenby. Causes and consequences of increased glucose metabolism of cancers. *J. Nuclear Med.*, 49:24S–42S, 2008.
- [285] M. Gimbrone, R. Cotran, S. Leapman, and J. Folkman. Tumor growth and neovascularization: an experimental model using the rabbit cornea. *J. Nat. Cancer Inst.*, 52:413–427, 1974.
- [286] J. Glazier and F. Garner. Simulation of the differential adhesion driven rearrangement of biological cells. *Phys. Rev. E*, 47:2128–2154, 1993.
- [287] R. Godde and H. Kurz. Structural and biophysical simulation of angiogenesis and vascular remodeling. *Dev. Dyn.*, 220(4):387–401, 2001.
- [288] J. J. Going and T. J. Mohun. Human breast duct anatomy, the ‘sick lobe’ hypothesis and intraductal approaches to breast cancer. *Breast. Canc. Res. and Treat.*, 97(3):0167–6806, 2006.
- [289] J. D. Gordan, J. A. Bertout, C.-J. Hu, J. A. Diehl, and M. C. Simon. HIF-2 $\alpha$  promotes hypoxic cell proliferation by enhancing c-Myc transcriptional activity. *Canc. Cell*, 11(4):335–47, 2007.
- [290] V. Gordon, M. Valentine, M. Gardel, D. Andor-Ardó, S. Dennison, A. Bogdanov, D. Weitz, and T. Deisboeck. Measuring the mechanical stress induced by an expanding multicellular tumor system: a case study. *Exp. Cell Res.*, 289:58–66, 2003.
- [291] T. Graeber, C. Osmanian, T. Jacks, and *et al.* Hypoxia-mediated selection of cells with diminished apoptotic potential in solid tumors. *Nature*, 379:88–91, 1996.
- [292] F. Graner and J. Glazier. Simulation of biological cell sorting using a two-dimensional extended potts model. *Phys. Rev. Lett.*, 69:2013–2016, 1992.
- [293] L. Graziano and L. Preziosi. Mechanics in tumor growth. In F. Mollica, L. Preziosi, and K. Rajagopal, editors, *Modeling of Biological Materials*, pages 267–328. Birkhaeuser, New York, 2007.
- [294] H. Greenspan. Models for the growth of a solid tumor by diffusion. *Stud. Appl. Math.*, 51:317–340, 1972.
- [295] H. Greenspan. On the growth and stability of cell cultures and solid tumors. *J. Theor. Biol.*, 56:229–242, 1976.
- [296] A. Grin, G. Horne, M. Ennis, and F. P. O’Malley. Measuring extent of ductal carcinoma in situ in breast excision specimens: a comparison of 4 methods. *Arch. Pathol. Lab. Med.*, 133:31–7, 2009.
- [297] K. Groebe, S. Erz, and W. Mueller-Klieser. Glucose diffusion coefficients determined from concentration profiles in emt6 tumor spheroids incubated in radioactively labeled l-glucose. *Adv. Exp. Med. Biol.*, 361:619–625, 1994.
- [298] J. Guck, S. Schinkinger, B. Lincoln, F. Wottawah, S. Ebert, M. Romeyke, D. Lenz, H. M. Erickson, R. Ananthkrishnan, D. Mitchell, J. Käs, S. Ulvick, and C. Bilby. Optical deformability as an inherent cell marker for testing malignant transformation and metastatic competence. *Biophys. J.*, 88(5):3689–98, 2005.
- [299] C. Guiot, P. D. Santo, and T. Deisboeck. Morphological instability and cancer invasion: a ‘splashing water drop’ analogy. *Theor. Biol. Med. Model.*, 4:4, 2007.
- [300] G. P. Gupta and J. Massagu. Cancer Metastasis: Building a Framework. *Cell*, 127(4):679–695, 2006.

- 
- [301] G. Hamilton. Multicellular spheroids as an in vitro tumor model. *Cancer Letters*, 131:29–34, 1998.
- [302] D. Hanahan and R. Weinberg. The hallmarks of cancer. *Cell*, 100:57–70, 2000.
- [303] R. K. Hansen and M. J. Bissell. Tissue architecture and breast cancer: the role of extracellular matrix and steroid hormones. *Endocrine-Related Cancer*, 7(2):95–113, 2000.
- [304] B. D. Harms, G. M. Bassi, A. R. Horwitz, and D. A. Lauffenburger. Directional persistence of EGF-induced cell migration is associated with stabilization of lamellipodial protrusions. *Biophys. J.*, 88(2):1479–88, 2005.
- [305] A. Harris. Hypoxia—a key regulatory factor in tumor growth. *Nat. Rev. Cancer*, 2:38–47, 2002.
- [306] A. Harten, B. Engquist, S. Osher, and S. R. Chakravarthy. Uniformly high order accurate essentially non-oscillatory schemes, III. *J. Comput. Phys.*, 71:231–303, 1987.
- [307] H. Hashizume, P. Baluk, S. Morikawa, J. McLean, G. Thurston, S. Roberge, R. Jain, and D. McDonald. Openings between defective endothelial cells explain tumor vessel leakiness. *Am. J. Pathol.*, 156:1363–1380, 2000.
- [308] H. Hatzikirou, L. Brusch, and A. Deutsch. From cellular automaton rules to an effective macroscopic mean field description. *Math. Biosci.*, 2009. in review.
- [309] H. Hatzikirou, A. Deutsch, C. Schaller, M. Simon, and K. Swanson. Mathematical modeling of glioblastoma tumour development: A review. *Math. Models Meth. Appl. Sci.*, 15:1779–1794, 2005.
- [310] M. A. Hayat. *Methods of Cancer Diagnosis, Therapy, and Prognosis: Liver Cancer*. Springer, New York, fifth edition, 2009.
- [311] Y. Hegerfeldt, M. Tusch, E. Brocker, and P. Friedl. Collective cell movement in primary melanoma explants: plasticity of cell-cell interaction, 1-integrin function, and migration strategies. *Cancer Res.*, 62:2125–2130, 2002.
- [312] G. Helmlinger, P. Netti, H. Lichtenbeld, R. Melder, and R. Jain. Solid stress inhibits the growth of multicellular tumor spheroids. *Nat. Biotech.*, 15:778–783, 1997.
- [313] G. Helmlinger, F. Yuan, M. Dellian, and R. Jain. Interstitial pH and pO<sub>2</sub> gradients in solid tumors in vivo: high-resolution measurements reveal a lack of correlation. *Nat. Med.*, 3:177–182, 1997.
- [314] R. S. Herbst. Review of epidermal growth factor receptor biology. *Int. J. Rad. Oncol. Biol. Phys.*, 59(2 (S1)):S21–S26, 2004.
- [315] S.-i. Hino, C. Tanji, K. I. Nakayama, and A. Kikuchi. Phosphorylation of  $\beta$ -catenin by cyclic AMP-dependent protein kinase stabilizes  $\beta$ -catenin through inhibition of its ubiquitination. *Molec. Cell. Biol.*, 25(20):9063–72, 2005.
- [316] S. Hiratsuka, K. Nakamura, S. Iwai, M. Murakami, T. Itoh, H. Kijima, J. M. Shipley, R. M. Senior, and M. Shibuya. MMP9 induction by vascular endothelial growth factor receptor-1 is involved in lung-specific metastasis. *Cancer Cell*, 2(4):289–300, 2002.
- [317] M. Höckel, K. Schlenger, B. Aral, M. Mitze, U. Schaffer, and P. Vaupel. Association between tumor hypoxia and malignant progression in advanced cancer of the uterine cervix. *Cancer Res.*, 56:4509–4515, 1996.
- [318] M. Höckel, K. Schlenger, S. Hoekel, and P. Vaupel. Hypoxic cervical cancers with low apoptotic index are highly aggressive. *Cancer Res.*, 59:4525–4528, 1999.
- [319] M. Höckel, K. Schlenger, and P. Vaupel. Tumor hypoxia: definitions and current clinical, biologic, and molecular aspects. *J. Natl. Cancer Inst.*, 93:266–276, 2001.
- [320] C. Hoguea, B. Murray, and J. Sethian. Simulating complex tumor dynamics from avascular to vascular growth using a general level-set method. *J. Math. Biol.*, 53:86–134,

- 2006.
- [321] P. Hogeweg. Evolving mechanisms of morphogenesis: On the interplay between differential adhesion and cell-differentiation. *J. Theor. Biol.*, 203:317–333, 2000.
- [322] J. Holash, P. Maisonpierre, D. Compton, P. Boland, C. Alexander, D. Zagzag, G. Yancopoulos, and S. Wiegand. Vessel cooption, regression, and growth in tumors mediated by angiopoietins and vegf. *Science*, 284:1994–1998, 1999.
- [323] J. Holash, S. Wiegand, and G. Yancopoulos. New model of tumor angiogenesis: dynamic balance between vessel regression and growth mediated by angiopoietins and vegf. *Oncogene*, 18:5356–5362, 1999.
- [324] M. Holmes and B. Sleeman. A mathematical model of tumor angiogenesis incorporating cellular traction and viscoelastic effects. *J. Theor. Biol.*, 202:95–112, 2000.
- [325] J. M. Horowitz, D. W. Yandell, S.-H. Park, S. Canning, P. Whyte, K. Buchkovich, E. Harlow, R. A. Weinberg, and T. P. Dryja. Point mutational inactivation of the retinoblastoma antioncogene. *Science*, 243(4893):937–940, 1989.
- [326] D. Horstmann, K. Painter, and H. Othmer. Aggregation under local reinforcement: From lattice to continuum. *Eur. J. Appl. Math.*, 15:545–576, 2004.
- [327] K. B. Hotary, E. D. Allen, P. C. Brooks, N. S. Datta, M. W. Long, and S. J. Weiss. Membrane type 1 matrix metalloproteinase usurps tumour growth control imposed by the three-dimensional extracellular matrix. *Cell*, 114(1):33–45, 2003.
- [328] M. Hu, J. Yao, L. Cai, K. E. Bachman, F. van den Brle, V. Velculescu, and K. Polyak. Distinct epigenetic changes in the stromal cells of breast cancers. *Nat. Genet.*, 37(8):899–905, 2005.
- [329] Z. Hu, K. Yuri, H. Ozawa, H. Lu, and M. Kawata. The *In Vivo* time course for elimination of adrenalectomy-induced apoptotic profiles from the granule cell layer of the rat hippocampus. *J. Neurosci.*, 17(11):3981–9, 1997.
- [330] J. Humphrey. Continuum biomechanics of soft biological tissues. *Proc. Roy. Soc. London A*, 459:3–46, 2003.
- [331] J. Humphrey and K. Rajagopal. A constrained mixture model for growth and remodeling of soft tissues. *Math. Mod. Meth. Appl. Sci.*, 12:407–430, 2002.
- [332] D. Ilic, E. A. Almeida, D. D. Schlaepfer, P. Dazin, S. Aizawa, and C. H. Damsky. Extracellular matrix survival signals transduced by focal adhesion kinase suppress p53-mediated apoptosis. *J. Cell Biol.*, 143:547–60, 1998.
- [333] J. H. Irving and J. G. Kirkwood. The statistical mechanical theory of transport processes. IV. The equations of hydrodynamics. *J. Chem. Phys.*, 18:817–829, 1950.
- [334] N. Ishii, D. Maier, A. Merlo, M. Tada, Y. Sawamura, A. Diserens, and E. V. Meir. Frequent co-alterations of tp53, p16/cdkn2a, p14arf, pten tumor suppressor genes in human glioma cell lines. *Brain Pathol.*, 9:469–479, 1999.
- [335] T. Ishii, J. Murakami, K. Notohara, H. M. Cullings, H. Sasamoto, T. Kambara, Y. Shirakawa, Y. Naomoto, M. Ouchida, K. Shimizu, N. Tanaka, J. R. Jass, and N. Matsubara. Orophageal squamous cell carcinoma may develop within a background of accumulating DNA methylation in normal and dysplastic mucosa. *Gut*, 56(1):13–19, 2007.
- [336] T. Ishikawa, Y. Kobayashi, A. Omoto, Y. Adachi, S. Nakagawa, T. Kaneko, K. Nishida, Y. Miyamoto, Y. Chimori, T. Yoshikawa, and M. Kondo. Calcification in untreated non-Hodgkin’s lymphoma of the jejunum. *Acta Haematol.*, 102(4):185–9, 1999.
- [337] T. Jackson. Intracellular accumulation and mechanism of action of doxorubicin in a spatio-temporal tumor model. *J. Theor. Biol.*, 220:201–213, 2003.

- 
- [338] T. Jackson. A mathematical investigation of the multiple pathways to recurrent prostate cancer: Comparison with experimental data. *Neoplasia*, 6:697–704, 2004.
- [339] T. Jackson. A mathematical model of prostate tumor growth and androgen-independent relapse. *Disc. Cont. Dyn. Sys. B*, 4:187–201, 2004.
- [340] T. Jackson and H. Byrne. A mechanical model of tumor encapsulation and transcapillary spread. *Math. Biosci.*, 180:307–328, 2002.
- [341] R. Jain. Determinants of tumor blood flow: a review. *Cancer Res.*, 48:2641–2658, 1988.
- [342] R. Jain. Physiological barriers to delivery of monoclonal antibodies and other macromolecules in tumors. *Cancer Res.*, 50:814s–819s, 1990.
- [343] R. Jain. Delivery of molecular medicine to solid tumors: lessons from in vivo imaging of gene expression and function. *J. Control. Release*, 74:7–25, 2001.
- [344] R. Jain. Normalizing tumor vasculature with anti-angiogenic therapy: a new paradigm for combination therapy. *Nat. Med.*, 7:987–989, 2001.
- [345] R. Jain. Molecular regulation of vessel maturation. *Nature Med.*, 9:685–693, 2003.
- [346] R. Jain. Normalization of tumor vasculature: An emerging concept in antiangiogenic therapy. *Science*, 307:58–62, 2005.
- [347] K. A. Janes and D. A. Lauffenburger. A biological approach to computational models of proteomic networks. *Curr. Opin. Chem. Biol.*, 10(1):73–80, 2006.
- [348] A. Jemal, R. Siegel, E. Ward, T. Murray, J. Xu, and M. J. Thun. Cancer statistics, 2007. *CA Cancer J. Clin.*, 57(1):43–66, 2007.
- [349] R. Jensen. Hypoxia in the tumorigenesis of gliomas and as a potential target for therapeutic measures. *Neurosurg. Focus*, 20:E24, 2006.
- [350] B. Jian, N. Narula, Q.-Y. Li, E. R. Mohler III, and R. J. Levy. Progression of aortic valve stenosis: TGF- $\beta$ 1 is present in calcified aortic valve cusps and promotes aortic valve interstitial cell calcification via apoptosis. *Ann. Thoracic Surg.*, 75(2):457–65, 2003.
- [351] G.-S. Jiang and C.-W. Shu. Efficient implementation of weighted ENO schemes. *J. Comput. Phys.*, 126:202–228, 1996.
- [352] T. X. Jiang and C. M. Chuong. Mechanism of skin morphogenesis I: Analyses with antibodies to adhesion molecules tenascin, NCAM, and integrin. *Dev. Biol.*, 150:82–98, 1992.
- [353] Y. Jiang, J. Pjesivac-Grbovic, C. Cantrell, and J. Freyer. A multiscale model for avascular tumor growth. *Biophys. J.*, 89:3884–3894, 2005.
- [354] A. Jones, H. Byrne, J. Gibson, and J. Dold. Mathematical model for the stress induced during avascular tumor growth. *J. Math. Biol.*, 40:473–499, 2000.
- [355] P. Jones and B. Sleeman. Angiogenesis-understanding the mathematical challenge. *Angiogenesis*, 9:127–138, 2006.
- [356] P. A. Jones and S. B. Baylin. The fundamental role of epigenetic events in cancer. *Nat. Rev. Genet.*, 3(6):415–28, 2002.
- [357] P. A. Jones and P. W. Laird. Cancer epigenetics comes of age. *Nat. Genet.*, 21(2):163–7, 1999.
- [358] R. E. Jr., K. O’Connor, D. Lacks, D. Schwartz, and R. Dotson. Dynamics of spheroid self-assembly in liquid-overlay culture of du 145 human prostate cancer cells. *Biotech. Bioeng.*, 72:579–591, 2001.
- [359] K. Kaibuchi, S. Kuroda, and M. Amano. Regulation of the cytoskeleton and cell adhesion by the Rho family GTPases in mammalian cells. *Annu. Rev. Biochem.*, 68:459–86, 1999.

- [360] F. Kallinowski, P. Vaupel, S. Runkel, G. Berg, H. Fortmeyer, K. Baessler, K. Wagner, W. Mueller-Klieser, and S. Walenta. Glucose uptake, lactate release, ketone body turnover, metabolic milieu and pH distributions in human cancer xenografts in nude rats. *Cancer Res.*, 48:7264–7272, 1988.
- [361] K. Kaneko, K. Satoh, and A. Masamune. T. myosin light chain kinase inhibitors can block invasion and adhesion of human pancreatic cancer cell lines. *Pancreas*, 24:34–41, 2002.
- [362] R. N. Kaplan, S. Rafii, and D. Lyden. Preparing the “Soil”: The Premetastatic Niche. *Cancer Res.*, 66(23):11089–93, 2006.
- [363] P. I. Karecla, S. J. Green, S. J. Bowden, J. Coadwell, and P. J. Kilshaw. Identification of a binding site for integrin  $\alpha\beta7$  in the N-terminal domain of E-cadherin. *J. Biol. Chem.*, 271:30909–15, 1996.
- [364] B. Kaur, F. Khwaja, E. Severson, S. Matheny, D. Brat, and E. VanMeir. Hypoxia and the hypoxia-inducible-factor pathway in glioma growth and angiogenesis. *Neuro-Oncol.*, 7:134–153, 2005.
- [365] D. Kay and R. Welford. A multigrid finite element solver for the Cahn-Hilliard equation. *J. Comput. Phys.*, 212:288–304, 2006.
- [366] P. Keller, F. Pampaloni, and E. Stelzer. Life sciences require the third dimension. *Curr. Op. Cell Biol.*, 18:117–124, 2006.
- [367] T. Kelly, Y. Yan, R. L. Osborne, A. B. Athota, T. L. Rozypal, J. C. Colclasure, et al. Proteolysis of extracellular matrix by invadopodia facilitates human breast cancer cell invasion and is mediated by matrix metalloproteinases. *Clin. Exp. Metastasis*, 16(6):501–12, 1998.
- [368] P. Kenny, G. Lee, and M. Bissell. Targeting the tumor microenvironment. *Front. Biosci.*, 12:3468–3474, 2007.
- [369] K. Keren, Z. Pincus, G. M. Allen, E. L. Barnhart, G. Marriott, A. Mogilner, et al. Mechanism of shape determination in motile cells. *Nature*, 453(7194):475–80, 2008.
- [370] K. Kerlikowske, A. Molinaro, I. Cha, B. M. Ljung, V. L. Ernster, K. Stewart, K. Chew, D. H. Moore 2nd, and F. Waldman. Characteristics associated with recurrence among women with ductal carcinoma in situ treated by lumpectomy. *J. Natl. Cancer Inst.*, 95(22):1692–702, 2003.
- [371] J. F. R. Kerr, C. M. Winterford, and B. V. Harmon. Apoptosis. its significance in cancer and cancer therapy. *Cancer*, 73(8):2013–26, 1994.
- [372] I. Kevrekidis, C. Gear, J. Hyman, P. Kevrekidis, O. Runborg, and K. Theodoropoulos. Equation-free, coarse-grained multiscale computation: Enabling microscopic simulators to perform system-level analysis. *Comm. Math. Sci.*, 1:715–762, 2003.
- [373] P. Kevrekidis, N. Whitaker, D. Good, and G. Herring. Minimal model for tumor angiogenesis. *Phys. Rev. E*, 73:061926, 2006.
- [374] E. Khain and L. Sander. Generalized cahn-hilliard equation for biological applications. *Phys. Rev. E*, 77:051129, 2008.
- [375] E. Khain, L. Sander, and C. Schneider-Mizell. The role of cell-cell adhesion in wound healing. *J. Stat. Phys.*, 128:209–218, 2007.
- [376] S. Khan, M. Rogers, K. Khurana, M. Meguid, and P. Numann. Estrogen receptor expression in benign breast epithelium and breast cancer risk. *J. Natl. Canc. Inst.*, 90:37–42, 1998.
- [377] S. Khan, A. Sachdeva, S. Naim, M. Meguid, W. Marx, H. Simon, et al. The normal breast epithelium of women with breast cancer displays an aberrant response to estro-

- diol. *Canc. Epidemiol. Biomarkers Prev.*, 8:867–72, 1999.
- [378] S. Kharait, S. Hautaniemi, S. Wu, A. Iwabu, D. A. Lauffenburger, and A. Wells. Decision tree modeling predicts effects of inhibiting contractility signaling on cell motility. *BMC Syst. Biol.*, 1:9ff, 2007.
- [379] B. N. Kholodenko, O. V. Demin, G. Moehren, and J. B. Hoek. Quantification of short term signaling by the epidermal growth factor receptor. *J. Biol. Chem.*, 274(42):30169–81, 1999.
- [380] J. Kim. Three-dimensional tissue culture models in cancer biology. *J. Biomol. Screening*, 15:365–77, 2005.
- [381] J. Kim, K. Kang, and J. Lowengrub. Conservative multigrid methods for Cahn-Hilliard fluids. *J. Comput. Phys.*, 193:511–543, 2003.
- [382] J. Kim, K. Kang, and J. Lowengrub. Conservative multigrid methods for ternary Cahn-Hilliard systems. *Comm. Math. Sci.*, 2:53–77, 2004.
- [383] J. Kim and J. Lowengrub. Phase field modeling and simulation of three phase flows. *Int. Free Bound.*, 7:435–466, 2005.
- [384] Y. Kim, M. Stolarska, and H. Othmer. A hybrid model for tumor spheroid growth in vitro i: Theoretical development and early results. *Math. Meth. App. Sci.*, 17:1773–1798, 2007.
- [385] R. Kloner and R. Jennings. Consequences of brief ischemia: stunning, preconditioning, and their clinical implications: part 1. *Circulation*, 104:2981–2989, 2001.
- [386] M. Knowles and P. Selby, editors. *Introduction to the Cellular and Molecular Biology of Cancer*. Oxford University Press, Oxford, UK, fourth edition, 2005.
- [387] K. A. Knudsen, A. P. Soler, K. R. Johnson, and M. J. Wheelock. Interaction of  $\alpha$ -actin with the cadherin/catenin cell-cell adhesion complex via  $\alpha$ -catenin. *J. Cell. Biol.*, 130:66–77, 1995.
- [388] A. G. Knudson. Mutation and cancer: statistical study of retinoblastoma. *Proc. Natl. Acad. Sci. USA*, 68(4):820–3, 1971.
- [389] A. G. Knudson. Two genetic hits (more or less) to cancer. *Nat. Rev. Cancer*, 1(2):157–62, 2001.
- [390] L. Kopfstein and G. Christofori. Metastasis: cell-autonomous mechanisms versus contributions by the tumor microenvironment. *Cell. Mol. Life Sci.*, 63:449–468, 2006.
- [391] D. V. Krysko, T. V. Berghe, K. D’Herde, and P. Vandenabeele. Apoptosis and necrosis: Detection, discrimination and phagocytosis. *Methods*, 44:205–21, 2008.
- [392] E. Kuhl and G. Holzapfel. A continuum model for remodeling in living structures. *J. Mater. Sci.*, 42:8811–8823, 2007.
- [393] R. Kuiper, J. Schellens, G. Blijham, J. Beijnen, and E. Voest. Clinical research on antiangiogenic therapy. *Pharmacol. Res.*, 37:1–16, 1998.
- [394] P. Kunkel, U. Ulbricht, P. Bohlen, R. F. M.A. Brockmann, D. Stavrou, M. Westphal, and K. Lamszus. Inhibition of glioma angiogenesis and growth in vivo by systemic treatment with a monoclonal antibody against vascular endothelial growth factor receptor-2. *Cancer Res.*, 61:6624–6628, 2001.
- [395] L. Kunz-Schughart, J. P. Freyer, F. Hofstaedter, and R. Ebner. The use of 3-d cultures for high-throughput screening: The multicellular spheroid model. *J. Biomol. Screening*, 9:273–85, 2004.
- [396] R. Küppers and R. Dalla-Favera. Mechanisms of chromosomal translocations in B cell lymphomas. *Oncogene*, 20(40):5580–94, 2001.

- [397] A. Lal, C. Glazer, H. Martinson, H. Friedman, G. Archer, J. Sampson, and G. Riggins. Mutant epidermal growth factor receptor up-regulates molecular effectors of tumor invasion. *Cancer Res.*, 62:3335–3339, 2002.
- [398] C. R. Lamb. Diagnosis of calcification on abdominal radiographs. *Vet. Rad. Ultrasound*, 32(5):211–20, 1991.
- [399] O. T. Lampejo, D. M. Barnes, P. Smith, and R. R. Millis. Evaluation of infiltrating ductal carcinomas with a DCIS component: correlation of the histologic type of the in situ component with grade of the infiltrating component. *Semin. Diagn. Pathol.*, 11(3):215–22, 1994.
- [400] K. Lamszus, P. Kunkel, and M. Westphal. Invasion as limitation to anti-angiogenic glioma therapy. *Acta Neurochir Suppl.*, 88:169–177, 2003.
- [401] K. Landman and C. Please. Tumour dynamics and necrosis: Surface tension and stability. *IMA J. Math. Appl. Medicine Biol.*, 18:131–158, 2001.
- [402] M. C. Lane, M. A. Koehl, F. Wilt, and R. Keller. A role for regulated secretion of apical extracellular matrix during epithelial invagination in the sea urchin. *Development*, 117(3):1049–60, 1993.
- [403] H. Larjava, T. Salo, K. Haapasalmi, R. H. Kramer, and J. Heino. Expression of integrins and basement membrane components by wound keratinocytes. *J. Clin. Invest.*, 92(3):1425–35, 1993.
- [404] D. A. Lauffenburger. Cell signaling pathways as control modules: complexity for simplicity? *Proc. Natl. Acad. Sci. USA*, 97(10):5031–3, 2000.
- [405] C. Le Clainche and M. F. Carlier. Regulation of actin assembly associated with protrusion and adhesion in cell migration. *Physiol. Rev.*, 88(2):489–513, 2008.
- [406] D. Lee, H. Rieger, and K. Bartha. Flow correlated percolation during vascular remodeling in growing tumors. *Phys. Rev. Lett.*, 96:058104, 2006.
- [407] J. S. Lee, D. M. Basalyga, A. Simionescu, J. C. Isenburg, D. T. Sinionescu, and N. R. Vyavahare. Elastin calcification in the rat subdermal model is accompanied by up-regulation of degradative and osteogenic cellular responses. *Am. J. Pathol.*, 168:490–8, 2006.
- [408] S. Lee, S. K. Mohsin, S. Mao, S. G. Hilsenbeck, D. Medina, and D. C. Allred. Hormones, receptors, and growth in hyperplastic enlarged lobular units: early potential precursors of breast cancer. *Breast Canc. Res.*, 8(1):R6, 2006.
- [409] S. Lehoux and A. Tedgui. Signal transduction of mechanical stresses in the vascular wall. *Hypertension*, 32(2):338–345, 1998.
- [410] J. Less, T. Skalak, E. Sevick, and R. Jain. Microvascular architecture in a mammary carcinoma: branching patterns and vessel dimensions. *Cancer Res.*, 51:265–273, 1991.
- [411] H. Levine and M. Nilsen-Hamilton. Angiogenesis- a biochemical/mathematical perspective. *Tutorials in Math. Biosci. III*, 1872:23–76, 2006.
- [412] H. Levine, S. Pamuk, B. Sleeman, and M. Nilsen-Hamilton. Mathematical modeling of capillary formation and development in tumor angiogenesis: penetration into the stroma. *Bull. Math. Biol.*, 63:801–863, 2001.
- [413] H. Levine and B. Sleeman. Modelling tumour-induced angiogenesis. In L. Preziosi, editor, *ancer modelling and simulation*, pages 147–184. Chapman&Hall/CRC, Boca Raton, Florida, 2003.
- [414] H. Levine, B. Sleeman, and M. Nilsen-Hamilton. Mathematical modeling of the onset of capillary formation initiating angiogenesis. *J. Math. Biol.*, 42:195–238, 2001.

- 
- [415] H. Levine, M. Smiley, A. Tucker, and M. Nilsen-Hamilton. A mathematical model for the onset of avascular tumor growth in response to the loss of p53 function. *Cancer Informatics*, 2:163–188, 2006.
- [416] H. Levine, A. Tucker, and M. Nilsen-Hamilton. A mathematical model for the role of cell signal transduction in the initiation and inhibition of angiogenesis. *Growth Factors*, 20:155–175, 2002.
- [417] J. Li, P. Kevrekidis, C. Gear, and I. Kevrekidis. Deciding the nature of the coarse equation through microscopic simulations: The baby-bathwater scheme. *SIAM Review*, 49:469–487, 2007.
- [418] X. Li, V. Cristini, Q. Nie, and J. Lowengrub. Nonlinear three-dimensional simulation of solid tumor growth. *Disc. Dyn. Contin. Dyn. Syst. B*, 7:581–604, 2007.
- [419] S.-Y. Lin, W. Xia, J. C. Wang, K. Y. Kwong, and B. Spohn.  $\beta$ -Catenin, a novel prognostic marker for breast cancer: Its roles in Cyclin D1 expression and cancer progression. *Proc. Natl. Acad. Sci. USA*, 97(8):4262–6, 2000.
- [420] L. Liotta and E. Kohn. The microenvironment of the tumour-host interface. *Nature*, 411:375–379, 2001.
- [421] A. Lipton. Pathophysiology of Bone Metastases: How This Knowledge May Lead to Therapeutic Intervention. *J. Support. Oncol.*, 2(3):205–220, 2004.
- [422] X. D. Liu, S. Osher, and T. Chan. Weighted essentially non-oscillatory schemes. *J. Comput. Phys.*, 115:200–212, 1994.
- [423] B. Lloyd, D. Szczerba, M. Rudin, and G. Szekely. A computational framework for modeling solid tumour growth. *Phil. Trans. Roy. Soc. A*, 366:3301–3318, 2008.
- [424] B. Lloyd, D. Szczerba, and G. Szekely. A coupled finite element model of tumor growth and vascularization. In N. Ayache, S. Ourselin, and A. Maeder, editors, *Medical image computing and computer-assisted intervention-MICCA 2007: 10th international conference*, volume 4792 of *Lecture Notes in Computer Science*, pages 874–881. Springer, New York, 2007.
- [425] J. Lotem and L. Sachs. Epigenetics and the plasticity of differentiation in normal and cancer stem cells. *Oncogene*, 25(59):7663–7672, 2006.
- [426] R. M. B. Loureiro and P. A. D’Amore. Transcriptional regulation of vascular endothelial growth factor in cancer. *Cytokine Growth Factor Rev.*, 16(1):77–89, 2005.
- [427] J. Lowengrub, H. Frieboes, F. Jin, Y. Chuang, X. Li, P. Macklin, S. Wise, and V. Cristini. Nonlinear modeling of cancer: Bridging the gap between cells and tumors. *Nonlinearity*, 2010. in press.
- [428] V. Lubarda and A. Hoger. On the mechanics of solids with a growing mass. *Int. J. Solids Structures*, 39:4627–4664, 2002.
- [429] P. J. Lucio, M. T. Faria, A. M. Pinto, M. R. da Silva, M. E. Correia Jr., R. J. da Costa, and A. B. Parreira. Expression of adhesion molecules in chronic B-cell lymphoproliferative disorders. *Haematologica*, 83(2):104–11, 1998.
- [430] B. Lustig, B. Jerchow, M. Sachs, S. Wiler, T. Pietsch, U. Karsten, M. van de Wetering, H. Clevers, P. M. Schlag, W. Birchmeier, and J. Behrens. Negative feedback loop of Wnt signaling through upregulation of conductin/axin2 in colorectal and liver tumors. *Mol. Cell. Biol.*, 22:1184–93, 2002.
- [431] B. MacArthur and C. Please. Residual stress generation and necrosis formation in multi-cell tumour spheroids. *J. Math. Biol.*, 49:537–552, 2004.
- [432] P. Macklin. Numerical Simulation of Tumor Growth and Chemotherapy. M.S. thesis, University of Minnesota School of Mathematics, September 2003.



- 
- [433] P. Macklin. *Toward Computational Oncology: Nonlinear Simulation of Centimeter-Scale Tumor Growth in Complex, Heterogeneous Tissues*. Ph.D. dissertation, University of California, Irvine Department of Mathematics, June 2007.
- [434] P. Macklin, L. Carreras, J. Kim, S. Sanga, V. Cristini, and M. E. Edgerton. Mathematical analysis of histopathology indicates comparable oxygen uptake rates for quiescent and proliferating breast cancer cells in DCIS. (in preparation), 2009.
- [435] P. Macklin, J. Kim, G. Tomaiuolo, M. E. Edgerton, and V. Cristini. Agent-Based Modeling of Ductal Carcinoma in Situ: Application to Patient-Specific Breast Cancer Modeling. In T. Pham, editor, *Computational Biology: Issues and Applications in Oncology*, chapter 4, pages 77–112. Springer, New York, NY, 2009.
- [436] P. Macklin, J. Kim, G. Tomaiuolo, M. E. Edgerton, and V. Cristini. An agent-based cell model, with application to patient-specific ductal carcinoma in situ modeling. (in preparation), 2010.
- [437] P. Macklin and J. Lowengrub. Evolving interfaces via gradients of geometry-dependent interior Poisson problems: Application to tumor growth. *J. Comput. Phys.*, 203:191–220, 2005.
- [438] P. Macklin and J. Lowengrub. An improved geometry-aware curvature discretization for level set methods: Application to tumor growth. *J. Comput. Phys.*, 215:392–401, 2006.
- [439] P. Macklin and J. Lowengrub. Nonlinear simulation of the effect of microenvironment on tumor growth. *J. Theor. Biol.*, 245:677–704, 2007.
- [440] P. Macklin and J. Lowengrub. A new ghost cell/level set method for moving boundary problems: Application to tumor growth. *J. Sci. Comp.*, 35(2-3):266–99, 2008.
- [441] P. Macklin, S. McDougall, A. Anderson, M. Chaplain, V. Cristini, and J. Lowengrub. Multiscale modeling and nonlinear simulation of vascular tumour growth. *J. Math. Biol.*, 58(4-5):765–98, 2009.
- [442] A. D. C. Macknight, D. R. DiBona, A. Leaf, and M. C. Mortimer. Measurement of the composition of epithelial cells from the toad urinary bladder. *J. Membrane Biol.*, 6(2):108–26, 1971.
- [443] S. Madsen, E. Angell-Petersen, S. Spetalen, S. Carper, S. Ziegler, and H. Hirschberg. Photodynamic therapy of newly implanted glioma cells in the rat brain. *Lasers Surg. Med.*, 38:540–548, 2006.
- [444] S. Maggelakis and J. Adam. Mathematical model of prevascular growth of a spherical carcinoma. *Math. Comput. Modelling*, 13:23–38, 1990.
- [445] E. Maher, F. Furnari, R. Bachoo, D. Rowitch, D. Louis, W. Cavenee, and R. De-Pinho. Malignant glioma: genetics and biology of a grave matter. *Genes Dev.*, 15:1311–1333, 2001.
- [446] G. Majno and I. Joris. *Cells, Tissues, and Disease: Principles of General Pathology*. Oxford University Press, New York, second edition, 2004.
- [447] A. G. Makeev, D. Maroudas, and I. G. Kevrekidis. ‘Coarse’ stability and bifurcation analysis using stochastic simulators: Kinetic Monte Carlo examples. *J. Chem. Phys.*, 116:10083–10091, 2002.
- [448] A. G. Makeev, D. Maroudas, A. Z. Panagiotopoulos, and I. G. Kevrekidis. Coarse bifurcation analysis of kinetic Monte Carlo simulations: A lattice-gas model with lateral interactions. *J. Chem. Phys.*, 117:8229–8240, 2002.
- [449] R. Malladi, J. A. Sethian, and B. C. Vemuri. Shape Modeling with Front Propagation: A Level Set Approach. *IEEE Trans. Pattern Anal. Mach. Intell.*, 17(2), 1995.

- 
- [450] R. Malladi, J. A. Sethian, and B. C. Vemuri. A fast level set based algorithm for topology-independent shape modeling. *J. Math. Imaging Vision*, 6(2-3):269–289, 1996.
- [451] M. Malumbres and M. Barbacis. RAS oncogenes: the first 30 years. *Nat. Rev. Cancer*, 3(6):459–465, 2001.
- [452] L. Malvern. *Introduction of the mechanics of a continuous medium*. Prentice Hall, Englewood Cliffs, 1969.
- [453] D. Manoussaki, S. Lubkin, R. Vernon, and J. Murray. A mechanical model for the formation of vascular networks in vitro. *Acta Biotheor.*, 44:271–282, 1996.
- [454] N. Mantzaris, S. Webb, and H. Othmer. Mathematical modeling of tumor-induced angiogenesis. *J. Math. Biol.*, 49:111–187, 2004.
- [455] B. Marchant, J. Norbury, and J.A. Sherratt. Travelling wave solutions to a haptotaxis-dominated model of malignant invasion. *Nonlinearity*, 14:1653–1671, 2001.
- [456] A. F. Maree, A. Jilkin, A. Dawes, V. A. Grieneisen, and L. Edelstein-Keshet. Polarization and movement of keratocytes: a multiscale modelling approach. *Bull. Math. Biol.*, 68(5):1169–1211, 2006.
- [457] D. Martin and P. Colella. A cell-centered adaptive projection method for the incompressible Euler equations. *J. Comput. Phys.*, 163:271–312, 2000.
- [458] M. Marusic, Z. Baizer, J. Freyer, and S. Vuk-Pavlovic. Analysis of growth of multicellular tumour spheroids by mathematical models. *Cell Prolif.*, 27:73–94, 1994.
- [459] K. Matsumoto and T. Nakamura. Hepatocyte growth factor and the Met system as a mediator of tumor-stromal interactions. *Int. J. Cancer*, 119(3):477–483, 2006.
- [460] B. McArthur and C. Please. Residual stress generation and necrosis formation in multicell tumor spheroids. *J. Math. Biol.*, 49:537–552, 2004.
- [461] S. McDougall, A. Anderson, and M. Chaplain. Mathematical modelling of dynamic adaptive tumour-induced angiogenesis: Clinical applications and therapeutic targeting strategies. *J. Theor. Biol.*, 241:564–589, 2006.
- [462] S. McDougall, A. Anderson, M. Chaplain, and J. Sherratt. Mathematical modelling of flow through vascular networks: implications for tumour-induced angiogenesis and chemotherapy strategies. *Bull. Math. Biol.*, 64:673–702, 2002.
- [463] D. McElwain and L. Morris. Apoptosis as a volume loss mechanism in mathematical models of solid tumor growth. *Math. Biosci.*, 39:147–157, 1978.
- [464] C. Medrek, G. Landberg, and K. Andersson, T adn Leandersson. Wnt-5a-CKI $\alpha$  signaling promotes  $\beta$ -Catenin/E-Cadherin complex formation and intercellular adhesion in human breast epithelial cells. *J. Biol. Chem.*, 284:10968–79, 2009.
- [465] A. Menzel. Modelling of anisotropic growth in biological tissues- a new approach and computational aspects. *Biomech. Model. Mechanobiol.*, 3:147–171, 2005.
- [466] R. Merks, S. Brodsky, M. Goligorsky, S. Newman, and J. Glazier. Cell elongation is key to in silico replication of in vitro vasculogenesis and subsequent remodeling. *Dev. Biol.*, 289:44–54, 2006.
- [467] R. Merks and J. Glazier. Dynamic mechanisms of blood vessel growth. *Nonlinearity*, 19:C1–C10, 2006.
- [468] R. Merks, E. P. A. Shirinifard, and J. Glazier. Contact-inhibited chemotaxis in de novo and sprouting blood-vessel growth. *PLoS Comp. Biol.*, 4:e1000163, 2008.
- [469] A. Merlo. Genes and pathways driving glioblastomas in humans and murine disease models. *Neurosurg. Rev.*, 26:145–158, 2003.
- [470] N. Metropolis, A. Rosenbluth, M. Rosenbluth, A. Teller, and E. Teller. Equation of state calculations by fast computing machines. *J. Chem. Phys.*, 21:1087–1092, 1953.

- [471] P. Michieli, C. Basilico, S. Pennacchietti, A. Maffè, L. Tamagnone, S. Giordano, A. Bardelli, and P. Comoglio. Mutant met mediated transformation is ligand-dependent and can be inhibited by hgf antagonists. *Oncogene*, 18:5221–5231, 1999.
- [472] L. P. Middleton, G. Vlastos, N. Q. Mirza, S. Eva, and A. A. Sahin. Multicentric mammary carcinoma: evidence of monoclonal proliferation. *Cancer*, 94(7):1910–6, 2002.
- [473] F. Milde, M. Bergdorf, and P. Koumoutsakos. A hybrid model for three-dimensional simulations of sprouting angiogenesis. *Biophys. J.*, 95:3146–3160, 2008.
- [474] S. Mitran. *BEARCLAW a code for multiphysics applications with embedded boundaries: user's manuel*. Department of Mathematics, University of North Carolina, <http://www.amath.unc.edu/Faculty/mitran/bearclaw.html>, 2006.
- [475] D. F. Moffat and J. J. Going. Three dimensional anatomy of complete duct systems in human breast: pathological and developmental implications. *J. Clin. Pathol.*, 49:48–52, 1996.
- [476] A. Mogilner and L. Edelstein-Keshet. Regulation of actin dynamics in rapidly moving cells: a quantitative analysis. *Biophys. J.*, 83(3):1237–58, 2002.
- [477] R. Montesano, K. Matsumoto, T. Nakamura, and L. Orci. Identification of a fibroblast-derived epithelial morphogen as hepatocyte growth factor. *Cell*, 67:901–908, 1991.
- [478] J. Moreira and A. Deutsch. Cellular automaton models of tumor development: A critical review. *Adv. Complex Sys.*, 5:247–267, 2002.
- [479] A. Morotti, S. Mila, P. Accornero, E. Tagliabue, and C. Ponzetto. K252a inhibits the oncogenic properties of met, the hgf receptor. *Oncogene*, 21:4885–4893, 2002.
- [480] K. Morton and D. Mayers. *Numerical Solution of Partial Differential Equations*. Cambridge, Cambridge, UK, second edition, 2005.
- [481] B. Mosadegh, W. Saadi, S. J. Wang, and N. L. Jeon. Epidermal growth factor promotes breast cancer cell chemotaxis in CXCL12 gradients. *Biotech. Bioeng.*, 100(6):1205–13, 2008.
- [482] W. Mueller-Klieser. Multicellular spheroids: a review on cellular aggregates in cancer research. *J. Cancer Res. Clin. Oncol.*, 113:101–122, 1987.
- [483] W. Mueller-Klieser. Three-dimensional cell cultures: from molecular mechanisms to clinical applications. *Am. J. Physiol. Cell Physiol.*, 273:C1109–C1123, 1997.
- [484] W. Mueller-Klieser, J. Freyer, and R. Sutherland. Influence of glucose and oxygen supply conditions on the oxygenation of multicellular spheroids. *Br. J. Cancer*, 53:345–353, 1986.
- [485] G. Müller and J.-J. Métois. *Crystal Growth: From Fundamentals to Technology*. Elsevier, 2004.
- [486] W. Mullins and R. Sekerka. Morphological instability of a particle growing by diffusion or heat flow. *J. Appl. Phys.*, 34:323–329, 1963.
- [487] G. R. Mundy. Metastasis to bone: causes, consequences and therapeutic opportunities. *Nat. Rev. Cancer*, 2(8):584–93, 2002.
- [488] J. Murray and G. Oster. Cell traction models for generation of pattern and form in morphogenesis. *J. Math. Biol.*, 33:489–520, 1984.
- [489] V. R. Muthukkaruppan, L. Kubai, and R. Auerbach. Tumor-induced neovascularization in the mouse eye. *J. Natl. Cancer Inst.*, 69(3):699–705, 1982.
- [490] K. Nabeshima, T. Moriyama, Y. Asada, N. Komada, T. Inoue, H. Kataoka, A. Sumiyoshi, and M. Koono. Ultrastructural study of TPA-induced cell motility: human well-differentiated rectal adenocarcinoma cells move as coherent sheets via localized modulation of cell–cell adhesion. *Clin. Exp. Med.*, 13(6):499–508, 1995.

- 
- [491] M. Nagane, A. Levitzki, A. Gazit, W. Cavenee, and H. Huang. Drug resistance of human glioblastoma cells conferred by a tumor-specific mutant epidermal growth factor receptor through modulation of bcl-x l and caspase-3-like proteases. *Proc. Natl. Acad. Sci. USA*, 95:5724–5729, 1998.
- [492] H. Naganuma, R. Kimurat, A. Sasaki, A. Fukamachi, H. Nukui, and K. Tasaka. Complete remission of recurrent glioblastoma multiforme following local infusions of lymphokine activated killer cells. *Acta Neurochir.*, 99:157–160, 1989.
- [493] J. Nagy. The ecology and evolutionary biology of cancer: A review of mathematical models of necrosis and tumor cell diversity. *Math. Biosci. Eng.*, 2:381–418, 2005.
- [494] M. N. Nakatsu, R. C. A. Sainson, J. N. Aoto, K. L. Taylor, M. Aitkenhead, S. Prez-del Pulgard, P. M. Carpenter, and C. C. W. Hughes. Angiogenic sprouting and capillary lumen formation modeled by human umbilical vein endothelial cells (HUVEC) in fibrin gels: the role of fibroblasts and Angiopoietin-1. *Microvasc. Res.*, 66:102–112, 2003.
- [495] M. A. H. Navarrete, C. M. maier, R. Falzoni, L. G. d. A. Quadros, E. C. Baracat, and A. C. P. Nazário. Assessment of the proliferative, apoptotic, and cellular renovation indices of the human mammary epithelium during the follicular and luteal phases of the menstrual cycle. *Breast Cancer Res.*, 7:R306–13, 2005.
- [496] C. Nelson and M. Bissell. Of extracellular matrix, scaffolds, and signaling: tissue architecture regulates development, homeostasis, and cancer. *Annu. Rev. Cell Dev. Biol.*, 22:287–309, 2006.
- [497] P. Netti, L. Baxter, Y. Boucher, R. Skalak, and R. Jain. Time dependent behavior of interstitial fluid pressure in solid tumors: Implications for drug delivery. *Cancer Res.*, 55:5451–5458, 1995.
- [498] A. Neville, P. Matthews, and H. Byrne. Interactions between pattern formation and domain growth. *Bull. Math. Biol.*, 68:1975–2003, 2006.
- [499] G. Ngwa and P. Maini. Spatio-temporal patterns in a mechanical model for mesenchymal morphogenesis. *J. Math. Biol.*, 33:489–520, 1995.
- [500] M. Nichols and T. Foster. Oxygen diffusion and reaction kinetics in the photodynamic therapy of multicell tumour spheroids. *Phys. Med. Biol.*, 39:2161–2181, 1994.
- [501] R. Nishikawa, X. Ji, R. Harmon, C. Lazar, G. Gill, W. Cavenee, and H. Huang. A mutant epidermal growth factor receptor common in human glioma confers enhanced tumorigenicity. *Proc. Natl. Acad. Sci. USA*, 91:7727–7731, 1994.
- [502] J. Nor, J. Christensen, J. Liu, M. Peters, D. Mooney, R. Strieter, and P. Polverini. Up-regulation of bcl-2 in microvascular endothelial cells enhances intratumoral angiogenesis and accelerates tumor growth. *Cancer Res.*, 61:2183–2188, 2001.
- [503] M. A. Nowak, N. L. Komarova, A. Sengupta, J. V. Prasad, I.-M. Shih, B. Vogelstein, and C. Lengauer. The role of chromosomal instability in tumor initiation. *Proc. Natl. Acad. Sci. USA*, 99(25):16226–16231, 2002.
- [504] J. O’Connor, A. Jackson, G. Parker, and G. Jayson. Dce-mri biomarkers in the clinical evaluation of antiangiogenic and vascular disrupting agents. *Br. J. Cancer*, 96:189–195, 2007.
- [505] K. Oda, Y. Matsuoka, A. Funahashi, and H. Kitano. A comprehensive pathway map of epidermal growth factor receptor signaling. *Mol. Syst. Biol.*, 1, 2005.
- [506] T. Ohtake, I. Kimijima, T. Fukushima, M. Yasuda, K. Sekikawa, S. Takenoshita, and R. Abe. Computer-assisted complete three-dimensional reconstruction of the mammary ductal/lobular systems. *Cancer*, 91:2263–72, 2001.

- 
- [507] B. Øksendal. *Stochastic Differential Equations: An Introduction with Applications*. Springer, New York, 6th edition, 2007.
- [508] M. Orme and M. Chaplain. Two-dimensional models of tumour angiogenesis and anti-angiogenesis strategies. *Math. Med. Biol.*, 14:189–205, 1997.
- [509] S. Osher and R. Fedkiw. Level Set Methods: An Overview and Some Recent Results. *J. Comput. Phys.*, 169(2):463–502, 2001.
- [510] S. Osher and R. Fedkiw. *Level Set Methods and Dynamic Implicit Surfaces*. Springer, New York, NY, 2002.
- [511] S. Osher and J. Sethian. Fronts propagating with curvature-dependent speed: algorithms based on hamilton-jacobi formulation. *J. Comput. Phys.*, 79:12, 1988.
- [512] H. Othmer and A. Stevens. Aggregation, blowup, and collapse: the abc’s of taxis in reinforced random walks. *Siam. J. Appl. Math.*, 57:1044–1081, 1997.
- [513] H. G. Othmer, S. R. Dunbar, and W. Alt. Models of dispersal in biological systems. *J. Math. Biol.*, 26:263–298, 1988.
- [514] M. Owen, T. Alarcón, P. Maini, and H. Byrne. Angiogenesis and vascular remodeling in normal and cancerous tissues. *J. Math. Biol.*, 58:689–721, 2009.
- [515] M. R. Owen, H. M. Byrne, and C. E. Lewis. Mathematical modelling of the use of macrophages as vehicles for drug delivery to hypoxic tumour sites. *J. Theor. Biol.*, 226(4):377–91, 2004.
- [516] T. Padera, B. Stoll, J. Tooredman, D. Capen, E. di Tomaso, and R. Jain. Cancer cells compress intratumour vessels. *Nature*, 427:695, 2004.
- [517] D. Page, T. Anderson, and G. Sakamoto. *Diagnostic Histopathology of the Breast*. Churchill Livingstone, New York, 1987.
- [518] D. L. Page, W. D. Dupont, L. W. Rogers, and M. Landenberger. Intraductal carcinoma of the breast: follow-up after biopsy only. *Cancer*, 49(4):751–8, 1982.
- [519] S. Paget. The distribution of secondary growths in cancer of the breast. *Lancet*, 133(3421):571–573, 1889.
- [520] S. Paku. First step of tumor-related angiogenesis. *Lab. Invest.*, 65:334–346, 1991.
- [521] D. Palmieri, C. E. Horak, J.-H. Lee, D. O. Halverson, and P. S. Steeg. Translational approaches using metastasis suppressor genes. *J. Bioenerg. Biomembr.*, 38(3-4):151–161, 2006.
- [522] E. Palsson and H. Othmer. A model for individual and collective cell movement in dictyostelium discoideum. *Proc. Nat. Acad. Sci. USA*, 97:10338–10453, 2000.
- [523] S. Pamuk. Qualitative analysis of a mathematical model for capillary formation in tumor angiogenesis. *Math. Models Meth. Appl. Sci.*, 13:19–33, 2003.
- [524] P. Panorchan, M. S. Thompson, K. J. Davis, Y. Tseng, K. Konstantopoulos, and D. Wirtz. Single-molecule analysis of cadherin-mediated cell-cell adhesion. *J. Cell Sci.*, 119:66–74, 2006.
- [525] W. Pao, T. Y. Wang, G. J. Riely, et al. KRAS mutations and primary resistance of lung adenocarcinomas to gefitinib or erlotinib. *PLoS Med.*, 2:e17, 2005.
- [526] S. Parnuk. A mathematical model for capillary formation and development in tumor angiogenesis: A review. *Chemotherapy*, 52:35–37, 2006.
- [527] S. Patan, S. Tanda, S. Roberge, R. Jones, R. Jain, and L. Munn. Vascular morphogenesis and remodeling in a human tumor xenograft: blood vessel formation and growth after ovariectomy and tumor implantation. *Circ. Research*, 89:732–739, 2001.
- [528] N. Patani, B. Cutuli, and K. Mokbel. Current management of DCIS: a review. *Breast Cancer Res. Treat.*, 111(1):1–10, 2008.

- 
- [529] A. Patel, E. Gawlinski, S. Lemieux, and R. Gatenby. A cellular automaton model of early tumor growth and invasion: The effects of native tissue vascularity and increased anaerobic tumor metabolism. *J. Theor. Biol.*, 213:315–331, 2001.
- [530] N. Paweletz and M. Knierim. Tumor-related angiogenesis. *Crit. Rev. Oncol. Hematol.*, 9:197–242, 1989.
- [531] S. Peirce. Computational and mathematical modeling of angiogenesis. *Microcirculation*, 15(8):739–751, 2008.
- [532] S. Pennacchietti, P. Michieli, M. Galluzzo, S. Giordano, and P. Comoglio. Hypoxia promotes invasive growth by transcriptional activation of the met protooncogene. *Cancer Cell*, 3:347–361, 2003.
- [533] C. Peskin. The immersed boundary method. *Acta Numer.*, 11:479–517, 2002.
- [534] C. S. Peskin. Flow patterns around heart valves: A numerical method. *J. Comput. Phys.*, 10(2):252–71, 1972.
- [535] J. Peterson, G. Carey, D. Knezevic, and B. Murray. Adaptive finite element methodology for tumour angiogenesis modelling. *Int. J. Num. Meth. Eng.*, 69:1212–1238, 2007.
- [536] G. Pettet, C. Please, M. Tindall, and D. McElwain. The migration of cells in multicell tumor spheroids. *Bull. Math. Biol.*, 63:231–257, 2001.
- [537] M. Plank and B. Sleeman. A reinforced random walk model of tumour angiogenesis and anti-angiogenic strategies. *Math. Med. Biol.*, 20:135–181, 2003.
- [538] M. Plank and B. Sleeman. Lattice and non-lattice models of tumour angiogenesis. *Bull. Math. Biol.*, 66:1785–1819, 2004.
- [539] C. Please, G. Pettet, and D. McElwain. A new approach to modeling the formation of necrotic regions in tumors. *Appl. Math. Lett.*, 11:89–94, 1998.
- [540] C. Please, G. Pettet, and D. McElwain. Avascular tumour dynamics and necrosis. *Math. Models Appl. Sci.*, 9:569–579, 1999.
- [541] N. Poplawski, U. Agero, J. Gens, M. Swat, J. Glazier, and A. Anderson. Front instabilities and invasiveness of simulated avascular tumors. *Bull. Math. Biol.*, 71:1189–1227, 2009.
- [542] L. Postovit, M. Adams, G. Lash, J. Heaton, and C. Graham. Oxygen-mediated regulation of tumor cell invasiveness. involvement of a nitric oxide signaling pathway. *J. Biol. Chem.*, 277:35730–35737, 2002.
- [543] J. Pouyssegur, F. Dayan, and N. Mazure. Hypoxia signalling in cancer and approaches to enforce tumour regression. *Nature*, 441:437–443, 2006.
- [544] F. Prall. Tumour budding in colorectal carcinoma. *Histopathology*, 50:151–162, 2007.
- [545] M. Preusser, H. Heinzl, E. Gelpi, K. Schonegger, C. Haberler, P. Birner, C. Marosi, M. Hegi, T. Gorlia, and J. Hainfellner. Histopathologic assessment of hot-spot microvessel density and vascular patterns in glioblastoma: Poor observer agreement limits clinical utility as prognostic factors: a translational research project of the european organization for research and treatment of cancer brain tumor group. *Cancer*, 107:162–170, 2006.
- [546] L. Preziosi. *Cancer Modelling and Simulation*. Chapman and Hall/CRC, London, 2003.
- [547] L. Preziosi and S. Astanin. Modelling the formation of capillaries. In A. Quarteroni, L. Formaggia, and A. Veneziani, editors, *Complex Systems in Biomedicine*. Springer, Milan, 2006.
- [548] L. Preziosi and A. Tosin. Multiphase modeling of tumor growth and extracellular matrix interaction: Mathematical tools and applications. *J. Math. Biol.*, 58:625–656, 2009.

- [549] A. Pries, B. Reglin, and T. Secomb. Structural adaptation and stability of microvascular networks: functional roles of adaptive responses. *Am. J. Physiol. Heart Circ. Physiol.*, 281:H1015–H1025, 2001.
- [550] A. Pries, B. Reglin, and T. Secomb. Structural adaptation of vascular networks: the role of pressure response. *Hypertension*, 38:1476–1479, 2001.
- [551] A. Pries and T. Secomb. Control of blood vessel structure: insights from theoretical models. *Am. J. Physiol. Heart Circ. Physiol.*, 288:1010–1015, 2005.
- [552] A. Pries and T. Secomb. Modeling structural adaptation of microcirculation. *Microcirculation*, 15(8):753–64, 2008.
- [553] A. Pries, T. Secomb, and P. Gaehtgens. Structural adaptation and stability of microvascular networks: theory and simulations. *Am. J. Physiol. Heart Cir. Physiol.*, 275:H349–H360, 1998.
- [554] W. C. Prozialeck, P. C. Lamar, and D. M. Appelt. Differential expression of E-cadherin, N-cadherin and beta-catenin in proximal and distal segments of the rat nephron. *BMC Physiol.*, 4(10), 2004.
- [555] V. Quaranta, K. Rejniak, P. Gerlee, and A. Anderson. Invasion emerges from cancer cell adaptation to competitive microenvironments: Quantitative predictions from multiscale mathematical models. *Sem. Cancer Biol.*, 18(5):338–48, 2008.
- [556] V. Quaranta, A. Weaver, P. Cummings, and A. Anderson. Mathematical modeling of cancer: The future of prognosis and treatment. *Clinica Chimica Acta*, 357:173–179, 2005.
- [557] C. M. Quick, W. L. Young, E. F. Leonard, S. Joshi, E. Gao, and T. Hashimoto. Model of structural and functional adaptation of small conductance vessels to arterial hypotension. *Am. J. Physiol. Heart Circ. Physiol.*, 279(4):H1645–H1653, 2000.
- [558] K. C. Quon and A. Berns. Haplo-insufficiency? Let me count the ways. *Genes Dev.*, 15(22):2917–21, 2001.
- [559] A. Ramanathan, C. Wang, and S. Schreiber. Perturbational profiling of a cell-line model of tumorigenesis by using metabolic measurements. *PNAS*, 102:5992–5997, 2005.
- [560] I. Ramis-Conde, M. Chaplain, and A. Anderson. Mathematical modelling of cancer cell invasion of tissue. *Math. Comp. Model.*, 47:533–545, 2008.
- [561] I. Ramis-Conde, D. Drasdo, A. Anderson, and M. Chaplain. Modeling the influence of the e-cadherin-beta-catenin pathway in cancer cell invasion: A multiscale approach. *Biophys. J.*, 95:155–165, 2008.
- [562] A. Rätz, A. Ribalta, and A. Voigt. Surface evolution of elastically stressed films under deposition by a diffuse interface model. *J. Comput. Phys.*, 214:187–208, 2006.
- [563] K. Rejniak. A single-cell approach in modeling the dynamics of tumor microregions. *Math. Biosci. Eng.*, 2:643–655, 2005.
- [564] K. Rejniak. An immersed boundary framework for modeling the growth of individual cells: An application to the early tumour development. *J.Theor. Biol.*, 247:186–204, 2007.
- [565] K. Rejniak and A. Anderson. A computational study of the development of epithelial acini: I. sufficient conditions for the formation of a hollow structure. *Bull. Math. Biol.*, 70:677–712, 2008.
- [566] K. Rejniak and A. Anderson. A computational study of the development of epithelial acini: II. necessary conditions for structure and lumen stability. *Bull. Math. Biol.*, 70:1450–1479, 2008.

- 
- [567] K. Rejniak and R. Dillon. A single cell-based model of the ductal tumor microarchitecture. *Comp. Math. Meth. Med.*, 8(1):51–69, 2007.
- [568] K. Rennstam and I. Hedenfalk. High-throughput genomic technology in research and clinical management of breast cancer. molecular signatures of progression from benign epithelium to metastatic breast cancer. *Breast Canc. Res.*, 8(4):213ff, 2006.
- [569] B. Ribba, T. Alarcón, K. Marron, P. Maini, and Z. Agur. The use of hybrid cellular automaton models for improving cancer therapy. In B. C. P.M.A. Slood and A. Hoekstra, editors, *ACRI, LNCS*, pages 444–453. Springer, Berlin, 2004.
- [570] B. Ribba, O. Saut, T. Colin, D. Bresch, E. Grenier, and J. P. Boissel. A multiscale mathematical model of avascular tumor growth to investigate the therapeutic benefit of anti-invasive agents. *J. Theor. Biol.*, 243(4):532–41, 2006.
- [571] A. Ridley, M. Schwartz, K. Burridge, R. Firtel, M. Ginsberg, G. Borisy, J. Parsons, and A. Horwitz. Cell migration: Integrating signals from front to back. *Science*, 302:1704–1709, 2003.
- [572] E. Robinson, K. Zazzali, S. Corbett, and R. Foty.  $\alpha 5b1$  integrin mediates strong tissue cohesion. *J. Cell. Sci.*, 116:377–386, 2003.
- [573] E. Rofstad and E. Halsør. Hypoxia-associated spontaneous pulmonary metastasis in human melanoma xenographs: involvement of microvascular hotspots induced in hypoxic foci by interleukin. *Br. J. Cancer*, 86:301–308, 2002.
- [574] E. Rofstad, H. Rasmussen, K. Galappathi, B. Mathiesen, K. Nilsen, and B. Graff. Hypoxia promotes lymph node metastasis in human melanoma xenografts by up-regulating the urokinase-type plasminogen activator receptor. *Cancer Res.*, 62:1847–1853, 2002.
- [575] J. Rohzin, M. Sameni, G. Ziegler, and B. Sloane. Pericellular pH affects distribution and secretion of cathepsin b in malignant cells. *Cancer Res.*, 54:6517–6625, 1994.
- [576] T. Roose, S. J. Chapman, and P. Maini. Mathematical models of avascular tumor growth. *SIAM Review*, 49:179–208, 2007.
- [577] T. Roose, P. Netti, L. Munn, Y. Boucher, and R. Jain. Solid stress generated by spheroid growth using a linear poroelastic model. *Microvascular Res.*, 66:204–212, 2003.
- [578] B. Rubenstein and L. Kaufman. The role of extracellular matrix in glioma invasion: A cellular potts model approach. *Biophys. J.*, 95:5661–5680, 2008.
- [579] J. Rubenstein, J. Kim, T. Ozawa, K. Zhang, M. Westphal, D. Deen, and M. Shuman. Anti-vegf antibody treatment of glioblastoma prolongs survival but results in increased vascular cooption. *Neoplasia*, 2:306–314, 2000.
- [580] K. Rygaard and M. Spang-Thomsen. Quantitation and gompertzian analysis of tumor growth. *Breast Cancer Res. Treat.*, 46:303–312, 1997.
- [581] Y. Saad and M. Schultz. Gmres: A generalized minimal residual algorithm for solving nonsymmetric linear systems. *SIAM J. Sci. Stat. Comput.*, 7:856–869, 1986.
- [582] E. Sahai. Mechanisms of cancer cell invasion. *Curr. Opin. Genet. Dev.*, 15:87–96, 2005.
- [583] T. Sairanen, R. Szepesi, M.-L. Karjalainen-Lindsberg, J. Saksi, A. Paetau, and P. J. Lindsberg. Neuronal caspase-3 and PARP-1 correlate differentially with apoptosis and necrosis in ischemic human stroke. *Acta Neuropathologica*, 118(4):541–52, 2009.
- [584] G. Sakamoto. Infiltrating carcinoma: major histological types. In D. Page and T. Anderson, editors, *Diagnostic Histopathology of the Breast*. Churchill-Livingstone, London, 1987.
- [585] M. E. Sanders, P. A. Schuyler, W. D. Dupont, and D. L. Page. The natural history of low-grade ductal carcinoma in situ of the breast in women treated by biopsy only



- revealed over 30 years of long-term follow-up. *Cancer*, 103(12):2481–4, 2005.
- [586] S. Sanga, M. E. Edgerton, P. Macklin, and V. Cristini. From receptor dynamics to directed cell motion: A predictive multiscale model of cell motility in complex microenvironments. in press, 2009.
- [587] S. Sanga, H. Frieboes, X. Zheng, R. Gatenby, E. Bearer, and V. Cristini. Predictive oncology: a review of multidisciplinary, multiscale in silico modeling linking phenotype, morphology and growth. *NeuroImage*, 37:S120–S134, 2007.
- [588] S. Sanga, J. Sinek, H. Frieboes, M. Ferrari, J. Fruehauf, and V. Cristini. Mathematical modeling of cancer progression and response to chemotherapy. *Expert Rev. Anticancer Ther.*, 6:1361–1376, 2006.
- [589] B. Sansone, P. D. Santo, M. Magnano, and M. Scalerandi. Effects of anatomical constraints on tumor growth. *Phys. Rev. E*, 64:21903, 2002.
- [590] B. Sansone, M. Scalerandi, and C. Condat. Emergence of taxis and synergy in angiogenesis. *Phys. Rev. Lett.*, 87:128102, 2001.
- [591] M. Santini, G. Rainaldi, and P. Indovina. Apoptosis, cell adhesion and the extracellular matrix in three-dimensional growth of multicellular tumor spheroids. *Crit. Rev. Oncol. Hematol.*, 36:75–87, 2000.
- [592] M. Sarntinoranont, F. Rooney, and M. Ferrari. Interstitial stress and fluid pressure within a growing tumor. *Ann. Biomed. Eng.*, 31:327–335, 2003.
- [593] J. Satulovsky, R. Lui, and Y. L. Wang. Exploring the control circuit of cell migration by mathematical modeling. *Biophys. J.*, 94(9):3671–83, 2008.
- [594] N. Savill and P. Hogeweg. Modeling morphogenesis: From single cells to crawling slugs. *J. Theor. Biol.*, 184:229–235, 1997.
- [595] J. L. Scarlett, P. W. Sheard, G. Hughes, E. C. Ledgerwood, H.-K. Ku, and M. P. Murphy. Changes in mitochondrial membrane potential during staurosporine-induced apoptosis in Jurkat cells. *FEBS Letters*, 475(3):267–72, 2000.
- [596] J. Schlessinger. Ligand-induced, receptor-mediated dimerization and activation of EGF receptor. *Cell*, 110(6):669–72, 2002.
- [597] K. Schmeichel, V. Weaver, and M. Bissel. Structural cues from the tissue microenvironment are essential determinants of the human mammary epithelial cell phenotype. *J. Mammary Gland Biol. Neoplasia*, 3:201–213, 1998.
- [598] L. S. Schulman and P. E. Seiden. Statistical mechanics of a dynamical system based on conway’s game of life. *J. Stat. Phys.*, 19(3):293–314, 1978.
- [599] E. Seftor, P. Meltzer, D. Kirshmann, J. Pe’er, A. Maniotis, J. Trent, R. Folberg, and M. Hendrix. Molecular determinants of human uveal melanoma invasion and metastasis. *Clin. Exp. Metastasis*, 19:233–246, 2002.
- [600] M. J. Seidensticker and J. Behrens. Biochemical interactions in the wnt pathway. *Biochim. Biophys. Acta*, 1495:168–82, 2000.
- [601] B. Selam, U. A. Kayisli, J. A. Garcia-Velasco, and A. Arici. Extracellular matrix-dependent regulation of FAS ligand expression in human endometrial stromal cells. *Biol. Reprod.*, 66(1):1–5, 2002.
- [602] G. L. Semenza. HIF-1, O<sub>2</sub>, and the 3 PHDs: How animal cells signal hypoxia to the nucleus. *Cell*, 107(1):1–3, 2001.
- [603] G. Serini, D. Ambrosi, E. Giraudo, A. Gamba, L. Preziosi, and F. Bussolino. Modeling the early stages of vascular network assembly. *EMBO J.*, 22:1771–1779, 2003.
- [604] S. Setayeshgar, C. Gear, H. Othmer, and I. Kevrekidis. Application of coarse integration to bacterial chemotaxis. *SIAM Multiscale Model. Sim.*, 4:307–327, 2005.

- 
- [605] J. A. Sethian. *Level Set Methods and Fast Marching Methods*. Cambridge University Press, New York, NY, 1999.
- [606] J. A. Sethian and P. Smereka. Level set methods for fluid interfaces. *Ann. Rev. of Fluid Mech.*, 35(1):341–372, 2003.
- [607] M. Shannon and B. Rubinsky. The effect of tumor growth on the stress distribution in tissue. *Adv. Biol. Heat Mass Transfer*, 231:35–38, 1992.
- [608] N. Sharifi, B. T. Kawasaki, E. M. Hurt, and W. L. Farrar. Stem Cells in Prostate Cancer: Resolving the Castrate-Resistant Conundrum and Implications for Hormonal Therapy. *Cancer Biol. Ther.*, 5(8):910–906, 2006.
- [609] C. J. Sherr. Cancer Cell Cycles. *Science*, 274(5293):1672–1677, 1996.
- [610] J. Sherratt. Traveling wave solutions of a mathematical model for tumor encapsulation. *SIAM J. Appl. Math.*, 60:392–407, 1999.
- [611] J. Sherratt and M. Chaplain. A new mathematical model for avascular tumour growth. *J. Math. Biol.*, 43:291–312, 2001.
- [612] A. N. Shiryaev. *Probability*. Springer, New York, NY, 2nd edition, 1995.
- [613] B. I. Shraiman. Mechanical feedback as a possible regulator of tissue growth. *Proc. Natl. Acad. Sci. USA*, 102(9):3318–23, 2005.
- [614] C.-W. Shu and S. Osher. Efficient implementation of essentially non-oscillatory shock-capturing schemes. *J. Comput. Phys.*, 77:439–471, 1988.
- [615] C.-W. Shu and S. Osher. Efficient implementation of essentially non-oscillatory shock capturing schemes, II. *J. Comput. Phys.*, 83:32–78, 1989.
- [616] D. Shweiki, A. Itin, D. Soffer, and E. Keshet. Vascular endothelial growth factor induced by hypoxia may mediate hypoxia-initiated angiogenesis. *Nature*, 359:843–845, 1992.
- [617] A. Sierra. Metastases and their microenvironments: linking pathogenesis and therapy. *Drug Resist. Updates*, 8:247–257, 2005.
- [618] S. A. Silver and F. A. Tavassoli. Ductal carcinoma in situ with microinvasion. *Breast J.*, 4(5):344–8, 1998.
- [619] M. J. Silverstein. Predicting residual disease and local recurrence in patients with ductal carcinoma in situ. *J. Natl. Cancer Inst.*, 89(18):1330–1, 1997.
- [620] M. J. Silverstein. Recent advances: diagnosis and treatment of early breast cancer. *BMJ*, 314(7096):1736ff, 1997.
- [621] M. J. Silverstein. Ductal carcinoma in situ of the breast. *Annu. Rev. Med.*, 51:17–32, 2000.
- [622] P. T. Simpson, J. S. Reis-Filho, T. Gale, and S. R. Lakhani. Molecular evolution of breast cancer. *J. Pathol.*, 205(2):248–54, 2005.
- [623] J. Sinek, H. Frieboes, X. Zheng, and V. Cristini. Two-dimensional chemotherapy simulations demonstrate fundamental transport and tumor response limitations involving nanoparticles. *Biomedical Microdevices*, 6:297–309, 2004.
- [624] J. Sinek, S. Sanga, X. Zheng, H. Frieboes, M. Ferrari, and V. Cristini. Predicting drug pharmacokinetics and effect in vascularized tumors using computer simulation. *J. Math. Biol.*, 58:485–510, 2009.
- [625] S. Skinner. Microvascular architecture of experimental colon tumors in the rat. *Cancer Res.*, 50:2411–2417, 1990.
- [626] V. I. F. Slettenaar and J. L. Wilson. The chemokine network: A target in cancer biology? *Adv. Drug Deliv. Rev.*, 58(8):962–974, 2006.
- [627] K. Smallbone, R. Gatenby, R. Gillies, P. Maini, and D. Gavaghan. Metabolic changes during carcinogenesis: Potential impact on invasiveness. *J. Theor. Biol.*, 244:703–713,

- 2007.
- [628] K. Smallbone, R. Gatenby, and P. Maini. Mathematical modelling of tumour acidity. *J. Theor. Biol.*, 255:106–112, 2008.
- [629] K. Smallbone, D. Gavaghan, R. Gatenby, and P. Maini. The role of acidity in solid tumor growth and invasion. *J. Theor. Biol.*, 235:476–484, 2005.
- [630] K. Smallbone, D. Gavaghan, P. Maini, and J. M. Brady. Quiescence as a mechanism for cyclical hypoxia and acidosis. *J. Math. Biol.*, 55:767–779, 2007.
- [631] S. A. A. Sohaib and R. H. Reznick. MR imaging in ovarian cancer. *Canc. Img.*, 7(Special Issue A):S119–29, 2007.
- [632] V. Spencer, R. Xu, and M. Bissell. Extracellular matrix, nuclear and chromatin structure, and gene expression in normal tissues and malignant tumors: a work in progress. *Adv. Cancer Res.*, 97:275–294, 2007.
- [633] T. A. Springer. Adhesion receptors of the immune system. *Nature*, 346(6283):425–34, 1990.
- [634] P. Steeg. Angiogenesis inhibitors: motivators of metastasis? *Nature Med.*, 9:822–823, 2003.
- [635] A. Stein, T. Demuth, D. Mobley, M. Berens, and L. Sander. A mathematical model of glioblastoma tumor spheroid invasion in a three-dimensional in vitro experiment. *Biophys. J.*, 92:356–365, 2007.
- [636] M. S. Steinberg and M. Takeichi. Experimental specification of cell sorting, tissue spreading, and specific spatial patterning by quantitative differences in cadherin expression. *Proc. Natl. Acad. Sci. USA*, 91:206–9, 1994.
- [637] A. Stephanou, S. McDougall, A. Anderson, and M. Chaplain. Mathematical modelling of flow in 2d and 3d vascular networks: Applications to anti-angiogenic and chemotherapeutic drug strategies. *Math. Comput. Model.*, 41:1137–1156, 2005.
- [638] A. Stephanou, S. McDougall, A. Anderson, and M. Chaplain. Mathematical modelling of the influence of blood rheological properties upon adaptative tumour-induced angiogenesis. *Math. Comput. Model.*, 44:96–123, 2006.
- [639] J. Stewart, P. Broadbridge, and J. Goard. Symmetry analysis and numerical modelling of invasion by malignant tumour tissue. *Nonlinear Dyn.*, 28:175–193, 2002.
- [640] C. Stokes and D. Lauffenburger. Analysis of the roles of microvessel endothelial cell random motility and chemotaxis in angiogenesis. *J. Theor. Biol.*, 152:377–403, 1991.
- [641] P. C. Stomper and F. R. Margolin. Ductal carcinoma in situ: the mammographer’s perspective. *Am. J. Roentgenology*, 162:585–91, 1994.
- [642] E. Stott, N. Britton, J. Glazier, and M. Zajac. Simulation of benign avascular tumour growth using the potts model. *Math. Comput. Model.*, 30:183–198, 1999.
- [643] D. Stupack and D. Cheresh. Get a ligand, get a life: Integrins, signaling and cell survival. *J. Cell. Sci.*, 115:3729–3738, 2002.
- [644] C. Sun and L. Munn. Lattice-boltzmann simulation of blood flow in digitized vessel networks. *Comp. and Math. Appl.*, 55:1594–1600, 2008.
- [645] S. Sun, M. Wheeler, M. Obeyesekere, and C. P. Jr. A deterministic model of growth factor-induced angiogenesis. *Bull. Math. Biol.*, 67:313–337, 2005.
- [646] S. Sun, M. Wheeler, M. Obeyesekere, and C. P. Jr. Multiscale angiogenesis modeling using mixed finite element methods. *Multiscale Model. Simul.*, 4:1137–1167, 2005.
- [647] X.-F. Sun and H. Zhang. Clinicopathological significance of stromal variables: angiogenesis, lymphangiogenesis, inflammatory infiltration, MMP and PINCH in colorectal carcinomas. *Mol. Cancer*, 5:43, 2006.

- 
- [648] K. Sundfor, H. Lyng, and E. Rofstad. Tumour hypoxia and vascular density as predictors of metastasis in squamous cell carcinoma of the uterine cervix. *Br. J. Cancer*, 78:822–827, 1998.
- [649] M. Sussman, P. Smereka, and S. Osher. A level set approach for computing solutions to incompressible two-phase flow. *J. Comput. Phys.*, 114(1):146–159, 1994.
- [650] R. Sutherland. Cell and environment interactions in tumor microregions: the multicell spheroid model. *Science*, 240:177–184, 1988.
- [651] R. Sutherland, J. Carlsson, R. Durand, and J. Yuhas. Spheroids in cancer research. *Cancer Res.*, 41:2980–2994, 1981.
- [652] K. Swanson, C. Bridge, J. Murray, and E. A. Jr. Virtual and real brain tumors: Using mathematical modeling to quantify glioma growth and invasion. *J. Neuro. Sci.*, 216:1–10, 2003.
- [653] L. A. Taber. An optimization principle for vascular radius including the effects of smooth muscle tone. *Biophys. J.*, 74(1):109–114, 1998.
- [654] P. J. Tannis, O. E. Nieweg, R. A. Valdés Olmos, and B. B. R. Kroon. Anatomy and physiology of lymphatic drainage of the breast from the perspective of sentinel node biopsy. *J. Am. Coll. Surg.*, 192(3):399–409, 2001.
- [655] Y. Tao and M. Chen. An elliptic-hyperbolic free boundary problem modelling cancer therapy. *Nonlinearity*, 19:419–440, 2006.
- [656] Y. Tao, N. Yoshida, and Q. Guo. Nonlinear analysis of a model of vascular tumour growth and treatment. *Nonlinearity*, 17:867–895, 2004.
- [657] M. J. Terol, M. Tormo, J. A. Martinez-Climent, I. Marugan, I. Benet, A. Ferrandez, A. Teruel, R. Ferrer, and J. Garcia-Conde. Soluble intercellular adhesion molecule-1 (s-ICAM-1/s-CD54) in diffuse large B-cell lymphoma: association with clinical characteristics and outcome. *Ann. Oncol.*, 14(3):467–74, 2003.
- [658] R. Thomlinson and L. Gray. The histological structure of some human lung cancers and the possible implications of radiotherapy. *Br. J. Cancer*, 9:539–549, 1955.
- [659] B. Thorne, A. Bailey, and S. Peirce. Combining experiments with multi-cell agent-based modeling to study biological tissue patterning. *Briefings in Bioinformatics*, 8:245–257, 2007.
- [660] M. Tindall, C. Please, and M. Peddie. Modelling the formation of necrotic regions in avascular tumours. *Math. Biosci.*, 211:34–55, 2008.
- [661] S. Tong and F. Yuan. Numerical simulations of angiogenesis in the cornea. *Microvasc. Res.*, 61:14–27, 2001.
- [662] A. Tosin. Multiphase modeling and qualitative analysis of the growth of tumor cords. *Networks Heterogen. Media*, 3:43–84, 2008.
- [663] A. Tosin, D. Ambrosi, and L. Preziosi. Mechanics and chemotaxis in the morphogenesis of vascular networks. *Bull. Math. Biol.*, 68:1819–1836, 2006.
- [664] P. Tracqui. Biophysical models of tumor growth. *Rep. Prog. Phys.*, 72:056701, 2009.
- [665] U. Trottenberg, C. Oosterlee, and A. Schüller. *Multigrid*. Academic Press, New York, 2005.
- [666] C. Truesdell and R. Toupin. Classical field theories. In S. Flugge, editor, *Handbuch der Physik, Vol III/I*. Springer-Verlag, Berlin, 1960.
- [667] S. Turner and J. Sherratt. Intercellular adhesion and cancer invasion: A discrete simulation using the extended potts model. *J. Theor. Biol.*, 216:85–100, 2002.
- [668] B. Tysnes and R. Mahesparan. Biological mechanisms of glioma invasion and potential therapeutic targets. *J. Neurooncol.*, 53:129–147, 2001.

- [669] P. Vajkoczy, M. Farhadi, A. Gaumann, R. Heidenreich, R. Erber, A. Wunder, J.C., M. M. Tonn, and G. Breier. Microtumor growth initiates angiogenic sprouting with simultaneous expression of vegf, vegf receptor-2, and angiopoietin-2. *J. Clin. Invest.*, 109:777–785, 2002.
- [670] L. van Kempen, D. Ruiter, G. van Muijen, and L. Coussens. The tumor microenvironment: a critical determinant of neoplastic evolution. *Eur. J. Cell. Biol.*, 82:539–548, 2003.
- [671] I. van Leeuwen, C. Edwards, M. Ilyas, and H. Byrne. Towards a multiscale model of colorectal cancer. *World Gastroenterol.*, 13:1399–1407, 2007.
- [672] V. V. Vasko and M. Saji. Molecular mechanisms involved in differentiated thyroid cancer invasion and metastasis. *Curr. Opin. Oncol.*, 19(1):11–17, 2007.
- [673] P. Vaupel, H. Haugland, T. Nicklee, A. Morrison, and D. Hedley. Hypoxia-inducible factor-1 alpha is an intrinsic marker for hypoxia in cervical cancer xenografts. *Cancer Res.*, 61:7394–7398, 2001.
- [674] R. Vernon, J. Angello, M. Iruela-Arispe, and T. Lane. Reorganization of basement membrane matrices by cellular traction promotes the formation of cellular networks in vitro. *Lab. Invest.*, 66:536–547, 1992.
- [675] E. Villa-Cuesta, E. Gonz/’alez-P/’erez, and J. Modolell. Apposition of *iroquois* expressing and non-expressing cells leads to cell sorting and fold formation in *Drosophila* imaginal wing disc. *BMC Devel. Biol.*, 7(106), 2007.
- [676] B. Vollmayr-Lee and A. Rutenberg. Stresses in growing soft tissues. *Acta Biomaterialia*, 2:493–504, 2006.
- [677] J. von Neumann. *Theory of Self-Replicating Automata*. University of Illinois Press, 1966. Edited by Arthur W. Burks.
- [678] C. Walker and G. Webb. Global existence of classical solutions for a haptotaxis model. *SIAM J. Math. Anal.*, 38(5):1694–1713, 2007.
- [679] T. Walles, M. Weimer, K. Linke, J. Michaelis, and H. Mertsching. The potential of bioartificial tissues in oncology research and treatment. *Onkologie*, 30:388–394, 2007.
- [680] R. Wang, L. Jimming, K. Lyte, N. K. Yashpal, F. Fellows, and C. G. Goodyer. Role for  $\beta 1$  integrin and its associated  $\alpha 3$ ,  $\alpha 5$ , and  $\alpha 6$  subunits in development of the human fetal pancreas. *Diabetes*, 54:2080–9, 2005.
- [681] Z. Wang, L. Zhang, J. Sagotsky, and T. Deisboeck. Simulating non-small cell lung cancer with a multiscale agent-based model. *Theor. Biol. Med. Model.*, 4:50, 2007.
- [682] J. Ward and J. King. Mathematical modelling of avascular tumour growth. *IMA J. Math. Appl. Medicine Biol.*, 14:36–69, 1997.
- [683] J. Ward and J. King. Mathematical modelling of avascular-tumour growth ii: modelling growth saturation. *Math. Med. Biol.*, 16:171–211, 1999.
- [684] J. Ward and J. King. Modelling the effect of cell shedding on avascular tumour growth. *J. Theor. Med.*, 2:155–174, 2000.
- [685] J. Ward and J. King. Mathematical modelling of drug transport in tumour multicell spheroids and monolayer cultures. *Math. Biosci.*, 181:177–207, 2003.
- [686] R. Wcislo and W. Dzwinel. Particle based model of tumor progression stimulated by the process of angiogenesis. In J. Adam and N. Bellomo, editors, *Computational Science - ICCS 2008*, pages 177–186. Springer, Heidelberg, 2008.
- [687] A. M. Weaver. Invadopodia: specialized cell structures for cancer invasion. *Clin. Exp. Metastasis*, 23(2):97–105, 2006.

- 
- [688] C. Wei, M. Larsen, M. P. Hoffman, and K. M. Yamada. Self-organization and branching morphogenesis of primary salivary epithelial cells. *Tissue Eng.*, 13(4):721–35, 2007.
- [689] O. D. Weiner, W. A. Marganski, L. F. Wu, S. J. Altschuler, and M. W. Kirschner. An actin-based wave generator organizes cell motility. *PLoS Biol.*, 5(9):e221, 08 2007.
- [690] S. R. Wellings, H. M. Jensen, and R. G. Marcum. An atlas of subgross pathology of the human breast with special reference to possible precancerous lesions. *J. Natl. Cancer Inst.*, 55(2):231–73, 1975.
- [691] A. Wells, B. Harms, A. Iwabu, L. Koo, K. Smith, L. Griffith, et al. Motility signaled from the EGF receptor and related systems. *Meth. Mol. Biol.*, 327:159–77, 2006.
- [692] A. Wells, J. Kassis, J. Solava, T. Turner, and D. A. Lauffenburger. Growth factor-induced cell motility in tumor invasion. *Acta Oncol.*, 41(2):124–30, 2002.
- [693] M. Welter, K. Bartha, and H. Rieger. Emergent vascular network inhomogeneities and resulting blood flow patterns in a growing tumor. *J. Theor. Biol.*, 250:257–80, 2008.
- [694] M. Welter, K. Bartha, and H. Rieger. Hot spot formation in tumor vasculature during tumor growth in an arterio-venous-network environment. arXiv.org  $\dot{\iota}$  q-bio  $\dot{\iota}$  arXiv:0801.0654v2, 2008.
- [695] N. Wentzensen, S. Vinokurova, and M. von Knebel Doeberitz. Systematic review of genomic integration sites of human papillomavirus genomes in epithelial dysplasia and invasive cancer of the female lower genital tract. *Canc. Res.*, 64(11):3878–84, 2004.
- [696] K. Wiesenfeld and F. Moss. Stochastic resonance and the benefits of noise: from ice ages to crayfish and squids. *Nature*, 373:33, 1995.
- [697] H. S. Wiley, S. Y. Shvartsman, and D. A. Lauffenburger. Computational modeling of the EGF-receptor system: a paradigm for systems biology. *Trends Cell. Biol.*, 13(1):43–50, 2003.
- [698] S. Wise, J. Kim, and J. Lowengrub. Solving the regularized, strongly anisotropic Cahn-Hilliard equation by an adaptive nonlinear multigrid method. *J. Comput. Phys.*, 226:414–446, 2007.
- [699] S. Wise, J. Lowengrub, and V. Cristini. An adaptive algorithm for simulating solid tumor growth using mixture models. in prep.
- [700] S. Wise, J. Lowengrub, H. Frieboes, and V. Cristini. Three-dimensional multispecies nonlinear tumor growth– i. model and numerical method. *J. Theor. Biol.*, 253:524–543, 2008.
- [701] E. K. Wolf, A. C. Smidt, and A. E. Laumann. Topical sodium thiosulfate therapy for leg ulcers with dystrophic calcification. *Arch. Dermatol.*, 144(12):1560–2, 2008.
- [702] K. Wolf and P. Friedl. Molecular mechanisms of cancer cell invasion and plasticity. *Br. J. Dermatology*, 154:11–15, 2006.
- [703] K. Wolf, R. Müller, S. Borgmann, E.-B. Bröcker, and P. Friedl. Amoeboid shape change and contact guidance: T-lymphocyte crawling through fibrillar collagen is independent of matrix remodeling by MMPs and other proteases. *Blood*, 102(9):3262–9, 2003.
- [704] J. Wu and S. Cui. Asymptotic behavior of solutions of a free boundary problem modeling the growth of tumors with stokes equations. *Discr. Contin. Dyn. Sys.*, 24(2):625–51, 2009.
- [705] J. Wu, F. Zhou, and S. Cui. Simulation of microcirculation in solid tumors. *IEEE/ICME Int. Conf. on Complex Med. Eng.*, pages 1555–1563, 2007.
- [706] M. Wurzel, C. Schaller, M. Simon, and A. Deutsch. Cancer cell invasion of brain tissue: guided by a prepattern? *J. Theor. Medicine*, 6:21–31, 2005.

- [707] Y. Xiong, P. Rangamani, B. Dubin-Thaler, M. Sheetz, and R. Iyengar. A three-dimensional stochastic spatio-temporal model of cell spreading. *Nat. Proc.*, 2007. Available from Nature Proceedings: <http://10.1038/npre.2007.62.2>.
- [708] R. Xu, V. Spencer, and M. Bissell. Extracellular matrix-regulated gene expression requires cooperation of swi/snf and transcription factors. *J. Biol. Chem.*, 282:14992–14999, 2007.
- [709] S. Xu. Hopf bifurcation of a free boundary problem modeling tumor growth with two time delays. *Chaos Solitons Fractals*, 41(5):2491–4, 2009.
- [710] Y. Xu and R. Gilbert. Some inverse problems raised from a mathematical model of ductal carcinoma in situ. *Math. Comp. Model.*, 49(3-4):814–28, 2009.
- [711] H. Yamaguchi, J. Wyckoff, and J. Condeelis. Cell migration in tumors. *Curr. Op. Cell Biol.*, 17:559–564, 2005.
- [712] K. Yamauchi, M. Yang, P. Jiang, M. Xu, N. Yamamoto, H. Tsuchiya, K. Tomita, A. R. Moossa, M. Bouvet, and R. M. Hoffman. Development of real-time subcellular dynamic multicolor imaging of cancer-cell trafficking in live mice with a variable-magnification whole-mouse imaging system. *Canc. Res.*, 66:4028–4214, 2006.
- [713] S. Young and R. Hill. Effects of reoxygenation of cells from hypoxic regions of solid tumors: anticancer drug sensitivity and metastatic potential. *J. Natl. Cancer Inst.*, 82:338–339, 1990.
- [714] S. Young, R. Marshall, and R. Hill. Hypoxia induces dna overreplication and enhances metastatic potential of murine tumor cells. *Proc. Natl. Acad. Sci. USA*, 85:9533–9537, 1988.
- [715] J. Yu, J. Rak, B. Coomber, D. Hicklin, and R. Kerbel. Effect of p53 status on tumor response to antiangiogenic therapy. *Science*, 295:1526–1528, 2002.
- [716] A. Zagorska and J. Dulak. HIF-1: the knowns and unknowns of hypoxia sensing. *Acta Biochimica Polonica*, 51(3):563–585, 2004.
- [717] D. Zagzag, A. R., M. Greco, H. Yee, J. Holash, S. Wiegand, S. Zabski, G. Yancopoulos, and M. Grumet. Vascular apoptosis and involution in gliomas precede neovascularization: a novel concept for glioma growth and angiogenesis. *Lab. Invest.*, 80:837–849, 2000.
- [718] M. Zajac, G. Jones, and J. Glazier. Model of convergent extension in animal morphogenesis. *Phys. Rev. Lett.*, 85:2022–2025, 2000.
- [719] M. H. Zaman, R. D. Kamm, P. Matsudaira, and D. A. Lauffenburger. Computational model for cell migration in three-dimensional matrices. *Biophys. J.*, 89(2):1389–97, 2005.
- [720] A. Zetterberg, O. Larsson, and K. G. Wilman. What is the restriction point? *Curr. Opin. Cell Biol.*, 7(6):835–842, 1995.
- [721] L. Zhang, C. Athale, and T. Deisboeck. Development of a three-dimensional multiscale agent-based tumor model: simulating gene-protein interaction profiles, cell phenotypes and multicellular patterns in brain cancer. *J. Theor. Biol.*, 244:96–107, 2007.
- [722] L. Zhang, C. Strouthos, Z. Wang, and T. Deisboeck. Simulating brain tumor heterogeneity with a multiscale agent-based model: Linking molecular signatures, phenotypes and expansion rate. *Math. Comp. Model.*, 49:307–319, 2009.
- [723] L. Zhang, Z. Wang, J. Sagotsky, and T. Deisboeck. Multiscale agent-based cancer modeling. *J. Math. Biol.*, 58(4-5):545–59, 2009.
- [724] G. Zhao, J. Wu, S. Xu, M. Collins, Q. Long, C. Koenig, Y. Jiang, J. Wang, and A. Padhani. Numerical simulation of blood flow and interstitial fluid pressure in solid

- 
- tumor microcirculation based on tumor-induced angiogenesis. *Mech. Sinica*, 23:477–483, 2007.
- [725] X. Zheng, S. Wise, and V. Cristini. Nonlinear simulation of tumor necrosis, neo-vascularization and tissue invasion via an adaptive finite-element/level-set method. *Bull. Math. Biol.*, 67:211–259, 2005.
- [726] F. Zhou and S. Cui. Bifurcation for a free boundary problem modeling the growth of multi-layer tumors. *Nonlinear Analysis Theory Meth. Appl.*, 68:2128–2145, 2008.
- [727] F. Zhou, J. Escher, and S. Cui. Well-posedness and stability of a free boundary problem modeling the growth of multi-layer tumors. *J. Diff. Eq.*, 244:2909–2933, 2008.
- [728] D. Zipori. The mesenchyme in cancer therapy as a target tumor component, effector cell modality and cytokine expression vehicle. *Cancer Metastasis Rev.*, 25(3):459–467, 2006.